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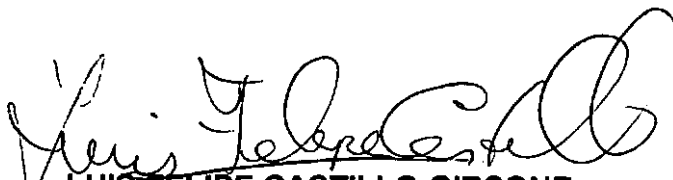
“En relación con la solicitud contenida en el expediente de la referencia y, en particular, para dar respuesta al Concepto Técnico No. 1583, contenido en su último Oficio proferido dentro del mismo, dentro del término conferido, por medio del presente escrito me permito manifestarle:

– Acompaño copia de los siguientes documentos solicitados por el señor Examinador conforme con el Artículo 46 de la Decisión 486:

1. “The Total Synthesis of *dl*- Δ^9 – Tetrahydrocannabinol and Four of Its Isomers”, K.E. Fahrenholtz et al, Journal of the American Chemical Society, vol. 89, no. 23, Noviembre 8, 1967, páginas 5934 – 5941.
2. “The stability of cannabis and its preparations on storage”, J.W. Fairbairn et al, J. Pharm. Pharmac. 1976, páginas 1 – 7.
3. “Cannabis XV. Preparation and Stability of Δ^9 Tetrahydrocannabinol – β -cyclodextrin inclusión complex”, Y. Shoyama et al, Vol. 46, Octubre 1983, páginas 633-637.

Le solicito se sirva agregar este escrito y sus anexos al Expediente.

De Usted Atentamente,


LUIS FELIPE CASTILLO GIBSON
T.P. No. 68.995 del C.S.J.

The Total Synthesis of *dl*- Δ^8 -Tetrahydrocannabinol and Four of Its Isomers¹

XP-002274651

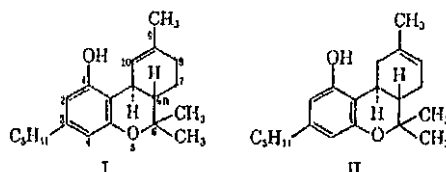
K. E. Fahrenholtz, M. Lurie, and R. W. Kierstead

Contribution from the Chemical Research Department, Hoffmann-La Roche Inc., Nutley, New Jersey 07110. Received June 2, 1967

Abstract: The total syntheses of the racemic modifications of two psychotomimetically active components of marihuana, Δ^8 -tetrahydrocannabinol (I) and Δ^9 -tetrahydrocannabinol (II), are described. The syntheses of three unnatural isomers, XII, XXI, and XXIII, are also presented. Cyclization of the coumarin V gave the dibenzopyran VII, which was converted by stereochemically controlled reactions to the *trans*-ketone Xa and the *cis*-ketone XI. The *trans*-ketone Xa was converted through the chloride XIV to a mixture rich in the Δ^8 isomer I, from which I was isolated in crystalline form. The *trans*-ketone Xa was also converted *via* the carbinol XIIIa to the Δ^9 isomer II. The *cis*-ketone XI was converted *via* the carbinol XXII to the *cis* analog XXIII of Δ^8 -tetrahydrocannabinol. The ketones Xa and XI were converted to the corresponding exocyclic methylene derivatives XII and XXI, respectively. The structure of the key intermediate IX was proven by its alternate synthesis from the chromanone XVII.

The crude resin obtained from the flowering tops of female plants of several *Cannabis sativa* L. varieties has been known and used as a psychotomimetic agent for many years.²⁻⁴ The structure of this material, known variously as marihuana, hashish, etc., had evaded elucidation for many years. Finally, the groups of Cahn,⁵ Todd,⁶ and Adams⁷ elucidated the gross structure of many of the components of the crude resin and several of the natural materials (physiologically inactive) were synthesized. The complete structure of the psychotomimetically active components, however, defied elucidation until more powerful modern physical methods became available. Recently the components of marihuana have received renewed attention. In 1964, Gaoni and Mechoulam,⁸ on the basis of nmr studies, and Santavy,⁹ on the basis of infrared studies and molecular rotation differences, determined the complete structure of the major physiologically active constituent, *l*- Δ^8 -tetrahydrocannabinol (I).¹⁰ One of these groups has also described^{11a} the first total synthesis of *dl*-I, the preparation requiring repeated column chromatography to effect purification of an intermediate. Early this year an elegant synthesis of *l*-cannabinidiol was published^{11b} in which this naturally occurring material was prepared in one step from olivetol. In view of the reports^{8,11c} that *l*-cannabinidiol can be con-

verted in unspecified yield to *l*- Δ^9 -tetrahydrocannabinol, this new synthesis of *l*-cannabinidiol may provide a convenient procedure for the synthesis of *l*- Δ^9 -tetrahydrocannabinol. Last year the isolation of another psychotomimetically active component, *l*- Δ^9 -tetrahydrocannabinol (II), from some varieties of Mexican hemp was an-



nounced¹² and a total synthesis of *dl*-II has also been reported,¹³ the isolation of *dl*-II requiring column chromatography followed by a preparative vapor phase chromatographic separation.^{13a}

The present paper reports the synthesis of *dl*- Δ^8 -tetrahydrocannabinol (I), *dl*- Δ^9 -tetrahydrocannabinol (II), and other tetrahydrocannabinols by procedures amenable to large-scale preparation of these compounds.

The von Pechmann condensation of olivetol¹⁴ (III) and diethyl α -acetoglutarate¹⁵ (IV) in the presence of phosphorus oxychloride gave only one isolated product in spite of the fact that two coumarins and two chromones are possible¹⁶ (Scheme 1). That the product isolated was the desired compound V was supported by its spectra and its eventual conversion to IX which was also prepared by another, unambiguous route (see below). The next step in the reaction sequence required an intramolecular condensation involving a methyl group vinylogous to a lactone carbonyl. A few similar intermolecular^{17,18} and intramolecular¹⁹ con-

(1) Part of these results have been reported in a preliminary communication: K. E. Fahrenholtz, M. Lurie, and R. W. Kierstead, *J. Am. Chem. Soc.*, **88**, 2079 (1966).

(2) (a) D. F. Downing, *Quart. Rev. (London)*, **16**, 133 (1962); (b) D. F. Downing in "Psychopharmacological Agents," Vol. 1, M. Gordon, Ed., Academic Press Inc., New York, N. Y., 1964, pp 585-607.

(3) R. Adams, *Harvey Lectures*, **37**, 168 (1941).

(4) A. R. Todd, *Experientia*, **2**, 55 (1946).

(5) R. S. Cahn, *J. Chem. Soc.*, 1400 (1933), and earlier papers.

(6) F. Berge, A. L. Morisson, H. Rinderknecht, A. R. Todd, A. D. Macdonald, and G. Woolfe, *ibid.*, 286 (1943), and earlier papers.

(7) R. Adams, M. Harfenist, and S. Loewe, *J. Am. Chem. Soc.*, **71**, 1624 (1949), and earlier papers.

(8) Y. Gaoni and R. Mechoulam, *ibid.*, **86**, 1646 (1964).

(9) F. Santavy, *Acta Univ. Palackianae Olomuc. Fac. Med.*, **35**, 5 (1964); *Chem. Abstr.*, **62**, 4057d (1965).

(10) Compound I has been called Δ^8 -tetrahydrocannabinol by other workers. However, we feel the numbering system used here is preferable, conforming to the *Chemical Abstracts* nomenclature of dibenzopyran compounds.

(11) (a) R. Mechoulam and Y. Gaoni, *J. Am. Chem. Soc.*, **87**, 3273 (1965); (b) T. Petržilka, W. Haefliger, C. Sikemeier, G. Ohloff, and A. Eschenmoser, *Helv. Chim. Acta*, **50**, 719 (1967); (c) Y. Gaoni and R. Mechoulam, *Tetrahedron*, **22**, 1481 (1966).

(12) R. L. Hively, W. A. Mosher, and F. W. Hoffmann, *J. Am. Chem. Soc.*, **88**, 1832 (1966).

(13) E. C. Taylor, K. Lenard, and Y. Shvo, *ibid.*, **88**, 367 (1966).

(13a) NOTE ADDED IN PROOF. After the acceptance of this manuscript, two syntheses of (-)- Δ^8 -tetrahydrocannabinol [(-)-II] appeared in print: T. Y. Jen, G. A. Hughes, and H. Smith, *ibid.*, **89**, 4551 (1967), and R. Mechoulam, P. Braun, and Y. Gaoni, *ibid.*, **89**, 4552 (1967). Both of these groups also prepared (+)-II and in addition the latter group converted (-)-II to (-)-I via (-)-XIV.

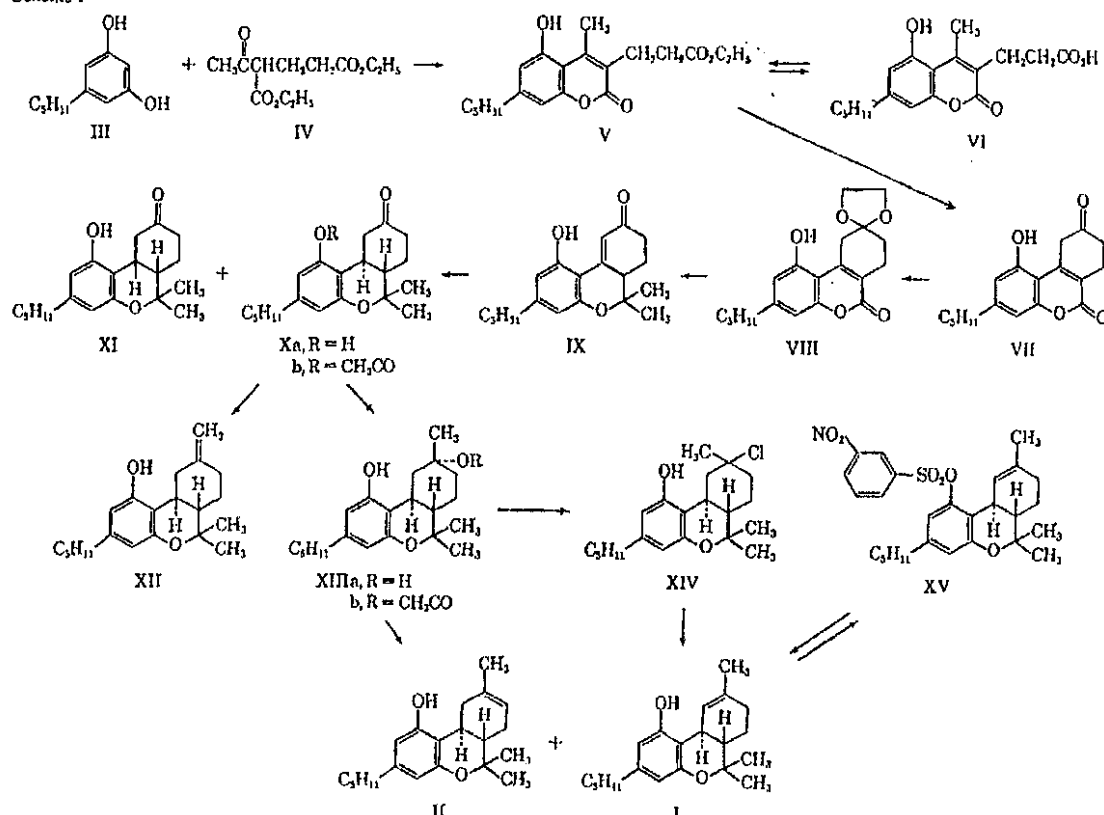
(14) C. M. Suter and A. W. Weston, *ibid.*, **61**, 232 (1939).

(15) H. Henecke, *Ber.*, **81**, 197 (1948).

(16) S. Sethna and R. Phadke, *Org. Reactions*, **7**, 1 (1953).

(17) W. Borscheu and R. Manteuffel, *Ann.*, **505**, 177 (1933).

Scheme 1



densations have been reported on compounds where the most acidic protons were on the methyl group. In V, however, the hydroxylic proton and the protons α to the ester group are all probably more acidic than the methyl protons. Treatment of V with sodium hydroxide in ethanol¹⁹ gave the corresponding acid VI rather than the cyclization product VII. The former compound was readily reconverted to V by treatment with the diethyl acetal of dimethylformamide.^{20,21} Treatment of V with sodium hydride in refluxing benzene gave no reaction. The use of potassium *t*-butoxide in dimethyl sulfoxide gave a low yield of cyclized product whereas sodium hydride in dimethyl sulfoxide gave good yields of VII accompanied by small and varying amounts of VI. The spectral properties of VII suggest that the double bond is conjugated with the lactone group forming a coumarin chromophore. In particular the nmr spectrum clearly shows the absence of olefinic protons. Treatment of VII with ethylene glycol gave the corresponding ketal VIII. The reaction of VIII with methylmagnesium iodide²² followed by acid hydrolysis then gave IX²³ which has also been prepared from XVII (see below). The nmr spectrum of IX

(18) R. Kuhn and C. Grundmann, *Ber.*, **69**, 1757 (1936).
(19) H. Meerwein, *Ann.*, **358**, 71 (1908).
(20) H. Vorbrüggen, *Angew. Chem. Intern. Ed. Engl.*, **2**, 211 (1963).
(21) H. Brechbühler, H. Büchi, E. Hatz, J. Schreiber, and A. Eschenmoser, *ibid.*, **2**, 212 (1963); *Helv. Chim. Acta*, **48**, 1746 (1965).
(22) R. Adams, C. K. Chin, and B. R. Baker, *J. Am. Chem. Soc.*, **62**, 2201 (1940).
(23) All compounds prepared in this paper which contain centers of asymmetry are racemic and for convenience are represented throughout by one enantiomeric series.

indicates the presence of an olefinic proton and the other spectra also confirm the rearranged position of the double bond. The reduction of IX with lithium in liquid ammonia at -70° gave a mixture of two products. The minor product was identical with the *cis*-ketone XI which was also prepared by an alternate and unambiguous route (see below). The major product of this reduction was the desired *trans*-ketone Xa. This ketone was allowed to react with triphenylphosphine-methylene to give the exocyclic analog XII of tetrahydrocannabinol. No deliberate attempts were made to isomerize the double bond to an endocyclic position but it was shown that XII is recovered unchanged under the conditions used for the dehydration of the carbinol XIIIa (see below).

As will be mentioned later the *cis*-ketone XI reacted completely with methylmagnesium iodide. However, when the *trans*-ketone Xa was treated with methylmagnesium iodide, even with a large excess of reagent or at the temperature of refluxing benzene, thin layer chromatography of the reaction mixture showed the presence of a large amount of unreacted starting material. Only small amounts of pure XIIIa could be obtained from these mixtures by crystallization. However, reaction of the corresponding acetate Xb with methylmagnesium iodide gave a product mixture (free of ketone Xa) which could be used directly for subsequent steps. This mixture could also be crystallized to obtain satisfactory yields of the carbinol XIIIa.^{24,25}

(24) The hydroxyl group in XIIIa is assigned the axial configuration because dehydration under conditions not vigorous enough to cause

Table I. Nuclear Magnetic Resonance Spectra of Tetrahydrocannabinols^a

Type of proton	<i>dl</i> -I In CDCl ₃ ^{b,c}	<i>dl</i> -I In CCl ₄ ^{b,d}	<i>dl</i> -II In CDCl ₃ ^c	<i>dl</i> -II In CCl ₄	<i>dl</i> -XII in CDCl ₃	<i>dl</i> -XXIII in CDCl ₃ ^c	<i>dl</i> -XXI in CDCl ₃
Aromatic	6.28 (d, <i>J</i> = 2) (1)	6.13 (d, <i>J</i> = 2) (1)	6.27 (d, <i>J</i> = 2) (1)	6.08 (d, <i>J</i> = 2) (1)	6.28 (d, <i>J</i> = 1.5) (1)	6.27 (br d) (2)*	6.25 (d, <i>J</i> = 1.5) (1)
Olefinic	6.13 (d, <i>J</i> = 2) (1)	5.97 (d, <i>J</i> = 2) (1)	6.08 (d, <i>J</i> = 2) (1)	5.88 (d, <i>J</i> = 2) (1)	6.10 (d, <i>J</i> = 1.5) (1)	6.12 (d, <i>J</i> = 2) (1)	6.07 (d, <i>J</i> = 1.5) (1)
C-10a H	6.32 (br) (1) C-10	6.28 (br) (1) C-10	5.40 (br) (1) C-8	5.32 (br) (1) C-8	4.77 (b) (3)'/ exocyclic	6.27 (br d) (2)* C-10	4.65 (br d, <i>J</i> = 3) (2) exo- cyclic
C-9 CH ₃	3.18 (br d) (1)	3.13 (br d) (1)	3.22 (br d) (1)	3.13 (br d) (1)	and OH	3.58 (br) (1)	
C-6 (CH ₃) ₂	1.68 (s)	1.65 (s)	1.70 (s)	1.65 (s)		1.68 (s)	
ω-CH ₃	1.40 (s)	1.37 (s)	1.37 (s)	1.30 (s)	1.38 (s)	1.38 (s)	1.38 (s)
	1.08 (s)	1.05 (s)	1.10 (s)	1.05 (s)	1.05 (s)	1.27 (s)	1.28 (s)
	0.88 (t) (3)	0.88 (t) (3)	0.88 (t) (3)	0.88 (t) (3)	0.87 (t) (3)	0.87 (t) (3)	0.87 (t) (3)

^a All spectra were determined on a Varian A-60 spectrometer. Values are given in ppm relative to TMS as internal standard. Coupling constants are given in cps. ^b Identical with natural *l*-I.²⁶ ^c Compare ref. 13. ^d Compare ref. 8. ^e The signal at 6.27 includes one aromatic proton and the C-10 olefinic proton. ^f The signal at 4.77 includes the two exocyclic protons and the phenolic OH which exchanges on addition of D₂O.

This apparent difference in reactivity of Xa and XI toward the Grignard reagent can be readily explained on the basis of steric factors. In the reaction of both Xa and XI with methylmagnesium iodide the initial reaction is probably abstraction of the hydroxylic proton to form the corresponding phenolate anions. Further reaction is then a competition between attack of the Grignard reagent on the carbonyl carbon and enolization of the carbonyl initiated by C-10 proton abstraction by the adjacent phenolate anion. Studies²⁵ by Corey and by Zimmerman have shown that enolization of cyclic ketones proceeds preferentially (but not exclusively) by abstraction of axial protons. Measurements on Dreiding models²⁷ of the ketones with ring C as a chair and ring B as a half-chair (most likely close to the most stable conformation) show that in Xa the hydroxylic oxygen is 1.8 and 2.8 Å distant from the equatorial and axial C-10 protons, respectively. In the *cis*-ketone XI these distances are 1.8 and 3.4 Å, respectively, and in addition the axial proton is sterically shielded from the hydroxylic oxygen by the equatorial hydrogen.

In the reaction of the *cis*-ketone XI, therefore, attack on the carbonyl carbon by the Grignard reagent is preferred over internal abstraction of either the equatorial C-10 proton or the sterically inaccessible axial proton, giving reaction mixtures essentially free of XI. With the *trans*-ketone Xa, however, internal proton abstraction competes favorably with Grignard attack and a large amount of Xa is recovered.

Treatment of the carbinol XIIIa with acetic anhydride in pyridine gave the corresponding diacetate which, upon selective hydrolysis, was converted to the monoacetate XIIIb. Pyrolysis of XIIIb gave impure II while dehydration of XIIIa with a catalytic amount of *p*-toluenesulfonic acid in benzene gave an excellent yield of racemic Δ⁸-tetrahydrocannabinol (II) shown by glpc to be different from and free of either I or XII. This

material had spectra compatible with its structure and the nmr spectrum was identical with that reported for *dl*-II¹² (see Table I).

The reaction of the carbinol XIIIa (or more conveniently the total carbinol mixture from the Grignard reaction on the acetate Xb) with Lucas reagent in acetic acid gave a good yield of the chloride XIV. If the Lucas reaction was carried out on the crude Grignard product from the ketone Xa, the resulting mixture could be separated readily into recovered Xa and the desired chloride XIV.

This chloride, on dehydrohalogenation with potassium hydroxide in ethanol, gave impure racemic Δ⁸-tetrahydrocannabinol (II) identical with the product from dehydration of the carbinol XIIIa. However, dehydrohalogenation of XIV with sodium hydride in tetrahydrofuran gave an excellent yield of a mixture of olefins²⁸ containing 74% I and 26% II. Treatment of this mixture with *m*-nitrobenzenesulfonyl chloride gave pure XV in 23% yield. Hydrolysis of XV gave an excellent yield of crystalline racemic Δ⁸-tetrahydrocannabinol (I),²⁹ shown by glpc to be free of either II or other detectable impurities and identical in all respects (except optical rotation and melting point) with *l*-Δ⁸-tetrahydrocannabinol, isolated from marijuana.³⁰

As a proof for the position of substitution in IX (and therefore also in the von Pechmann product V), we prepared IX by another route which is both unambiguous and which provides an alternate approach to the synthesis of these tetrahydrocannabinol isomers.

The reaction of olivetol (III) with 3-methylcrotonic acid (XVI) in the presence of boron trifluoride etherate at 125°³¹ led to the isolation of two products, XVII (50% yield) and XVIII (22% yield) (Scheme II). The ratio of products could be reversed simply by using boron trifluoride etherate at 25°³² which gives XVII (23% yield) and XVIII (63% yield). Attempted con-

isomerization of the exocyclic isomer XII gives the endocyclic product II. In an analogous case,³³ a 3β-methyl-3α-hydroxy steroid (axial hydroxyl) was dehydrated to endocyclic products whereas the isomeric 3α-methyl-3β-hydroxy steroid (equatorial hydroxyl) was dehydrated to the exocyclic product.

(25) C. E. Cook, R. C. Corley, and M. E. Wall, *Tetrahedron Letters*, 891 (1965).

(26) E. L. Eliel, "Stereochemistry of Carbon Compounds," McGraw-Hill Book Co., Inc., New York, N. Y., 1962, p. 241.

(27) A. S. Dreiding, *Helv. Chim. Acta*, 42, 1339 (1959).

(28) The different behavior of XIV to these basic systems can be explained readily by the assumption that with the sodium hydride-tetrahydrofuran system intramolecular dehydrochlorination is promoted by the initially formed phenolate anion.

(29) Previously obtained as an oil.¹⁴

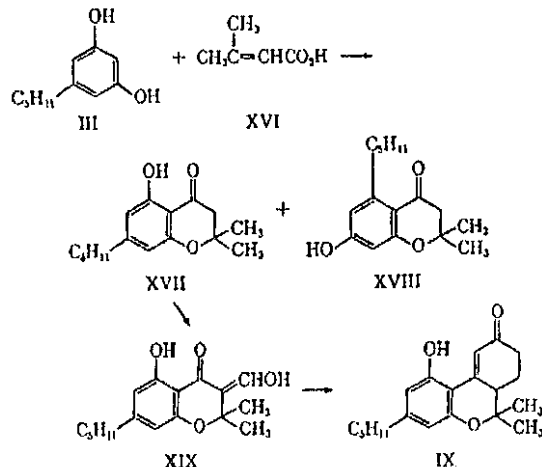
(30) We are indebted to Dr. Nathan B. Eddy of the Department of Health, Education, and Welfare for a sample of *l*-Δ⁸-tetrahydrocannabinol of natural origin.

(31) W. M. McLamore, *J. Am. Chem. Soc.*, 73, 2221 (1951).

(32) H. B. Bhat and K. Venkataraman, *Tetrahedron*, 19, 77 (1963).

condensations with a variety of other systems³²⁻³⁸ were less satisfactory.

Scheme II



Structural assignments of XVII and XVIII were readily made by a study of their spectra. The nmr spectra were most revealing; the hydroxylic proton in XVII appeared at 11.73 ppm while that of XVIII appeared at 8.77 ppm, indicating extensive intramolecular hydrogen bonding in XVII and not in XVIII.³⁷ In addition, the signal associated with the methylene group α to the aromatic ring is shifted downfield by 0.50 ppm in XVIII relative to XVII, indicating extensive deshielding of the methylene protons by the coplanar carbonyl group in XVIII.³⁸ The infrared spectrum of XVII was conspicuous by the absence of absorption characteristic of a free phenol³⁹ while a strong band at 1645 cm^{-1} indicative of strong intramolecular hydrogen bonding⁴⁰ was observed. On the other hand, the infrared spectrum of XVIII exhibited a sharp peak at 3590 cm^{-1} due to an unbonded hydroxyl and the absorption at 1670 cm^{-1} was more normal for an aromatic ketone. Both XVII and XVIII exhibited strong absorption in the ultraviolet region at 280 $\text{m}\mu$ (in ethanol). However, in strong base this band in the spectrum of XVII was shifted to 285 $\text{m}\mu$ and to 328 $\text{m}\mu$ in XVIII, indicative of an *o*-hydroxyaromatic ketone⁴¹ in XVII and a *p*-hydroxyaromatic ketone in XVIII.

The condensation of ethyl formate with XVII in the presence of sodium hydride gave XIX. Ring annelation of XIX with methyl vinyl ketone gave IX, identical in all respects with the material prepared *via* V, thus providing a proof for the position of substitution in V and its subsequent conversion products.

(33) H. R. Snyder and C. T. Elston, *J. Am. Chem. Soc.*, **77**, 364 (1955).

(34) M. Miyano and M. Matsui, *Bull. Chem. Soc. Japan*, **31**, 397 (1958).

(35) L. Smith and V. A. Engelhardt, *J. Am. Chem. Soc.*, **71**, 2671 (1949).

(36) R. D. Desai and M. Ekhtiar, *Proc. Indian Acad. Sci.*, **A8**, 194 (1958).

(37) L. M. Jackman, "Applications of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry," The Macmillan Co., New York, N. Y., 1959, p. 69.

(38) See ref 37, p. 122.

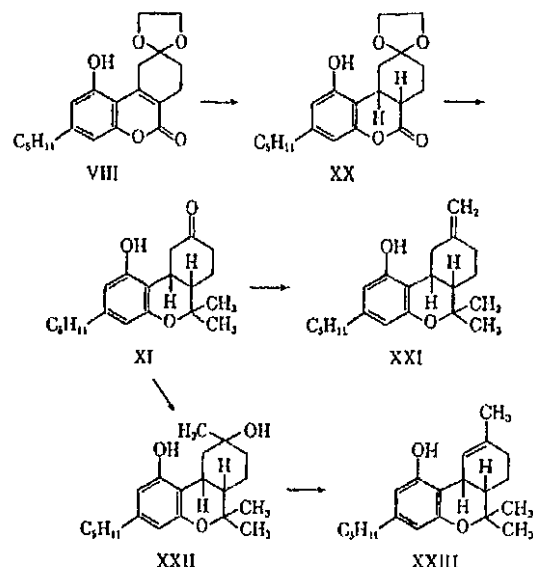
(39) I. J. Bellamy, "The Infra-red Spectra of Complex Molecules," John Wiley and Sons, Inc., New York, N. Y., 1954, p. 90.

(40) See ref 39, p. 124.

(41) A. I. Scott, "Interpretation of the Ultraviolet Spectra of Natural Products," The Macmillan Co., New York, N. Y., 1964, p. 109.

The *cis*-ketone XI was prepared by the unambiguous route given in Scheme III. Catalytic hydrogenation of

Scheme III



VIII in the presence of Raney nickel under high pressure and temperature⁴² gave the *cis*-lactone XX. Treatment of the latter compound with methylmagnesium iodide followed by hydrolysis of the protecting group gave the *cis*-ketone XI which was identical with the minor product of the chemical reduction of IX. The reaction of XI with triphenylphosphinemethylene gave the exocyclic *cis* analog XXI of tetrahydrocannabinol. The reaction of XI with methylmagnesium iodide proceeded to completion, unlike the corresponding reaction with the *trans*-ketone Xa, and the methylcarbinol XXII was readily isolated. This latter compound was then dehydrated with *p*-toluenesulfonic acid to give a single product, clearly an endocyclic olefin with an nmr spectrum identical with that reported¹³ for XXIII (the *cis* analog of Δ^8 -tetrahydrocannabinol).

Experimental Section⁴³

Ethyl 5-Hydroxy-4-methyl-7-pentylcoumarin-3-propionate (V). A mixture of 299 g (1.66 moles) of oilvetol⁴⁴ (II), 420 g (1.84 moles) of diethyl α -acetoglutarate⁴⁵ (IV), and 152 ml (254 g, 1.66 moles) of phosphorus oxychloride, protected from atmospheric moisture, was stirred at room temperature until the mixture solidified. The reaction was then allowed to stand for a total of 10 days. The solid was dissolved in chloroform, washed three times with water, dried (Na_2SO_4), and evaporated to an orange solid. This was recrystallized from benzene to give (in several crops) 481 g (84%) of V as colorless crystals, mp 120–122°. Further recrystallization gave the analytical sample: mp 123–124°; $\lambda_{\text{max}}^{\text{OH}}$ 250 $\text{m}\mu$ (inf) (ϵ 8600), 257 (10,000), 309 (15,000); $\nu_{\text{max}}^{\text{OH}}$ 1700 cm^{-1} (very broad and strong).

(42) P. L. deBenneville and R. Connor, *J. Am. Chem. Soc.*, **62**, 283 (1940).

(43) All melting points were determined in glass capillaries and are corrected. The infrared spectra were determined using a Beckman IR-9 spectrophotometer. The ultraviolet spectra were determined using a Cary 14 spectrophotometer. The nmr spectra were determined using a Varian A-60 spectrometer and are reported as parts per million relative to TMS as internal standard. All compounds prepared in this section had compatible infrared, ultraviolet, and nuclear magnetic resonance spectra, and spectral data are given only where significant. Florisil (Florisil Co.) is a synthetic magnesium silicate adsorbent.

Fahrenholtz, Lurie, Kierstead / Total Synthesis of *dl*- Δ^8 -Tetrahydrocannabinol

Anal. Calcd for $C_{16}H_{14}O_4$: C, 69.34; H, 7.57. Found: C, 69.28; H, 7.21.

5-Hydroxy-4-methyl-7-pentylcoumarin-3-propionic Acid (VI). A small sample of V was mixed with an equal weight of potassium carbonate in 50% aqueous ethanol and heated for 10 min under reflux. The solution was diluted with water, acidified with hydrochloric acid, and filtered. Recrystallization from acetonitrile gave the analytical sample as colorless crystals: mp 230–233°; $\nu_{\text{max}}^{\text{CHCl}_3}$ 1660 and 1710 cm^{-1} .

Anal. Calcd for $C_{18}H_{18}O_5$: C, 67.91; H, 6.97. Found: C, 67.70; H, 7.24.

Esterification of VI to V. A mixture of 1.0 g (3.1 mmoles) of VI, 10 ml of benzene, and 1.5 ml of the diethyl acetal of dimethylformamide^{20,21} was heated for 3 hr under reflux. The cooled reaction was diluted with ether, washed with 1 *N* hydrochloric acid and with water, dried (Na_2SO_4), and concentrated to an orange solid. Recrystallization from benzene gave 790 mg (72%) of V as colorless crystals, mp 121–124°, undepressed upon admixture with an authentic sample.

7,10-Dihydro-1-hydroxy-3-pentyl-6H-dibenzo[*b,d*]pyran-6,9(8H)-dione (VII). To the sodium hydride obtained by washing 72.0 g (1.5 moles) of 50% sodium hydride in mineral oil dispersion with dry hexane was added 120 g (0.35 mole) of V and the two powders were mixed thoroughly. The reaction flask was cooled to 15–17° and 1.2 l. of dimethyl sulfoxide was distilled from calcium hydride directly into the reaction flask. After stirring for an additional hour at 15–17°, the reaction was kept overnight in the refrigerator. After warming to room temperature the reaction mixture was poured into a rapidly stirred mixture of 4 l. of ice and water and 250 ml of concentrated hydrochloric acid, more ice being added as needed to keep the mixture cold. After stirring for an additional hour the slurry was filtered and the solids were washed well with water. The wet filter cake was then heated on the steam bath with excess concentrated sodium bicarbonate solution and, while still warm, was filtered. The filter cake was washed well with water and recrystallized from acetone to give (in several crops) 81 g (78%) of cyclized product VII as colorless crystals, mp 203–206°. Further recrystallization from acetonitrile gave the analytical sample: mp 205.5–207.5°; $\lambda_{\text{max}}^{\text{CHCl}_3}$ 215 m μ (inf) (ϵ 26,000), 250 (inf) (9750), 258 (10,500), 308 (14,550); $\nu_{\text{max}}^{\text{CHCl}_3}$ 1670 and 1710 cm^{-1} ; nmr (DMSO), no olefinic protons.

Anal. Calcd for $C_{24}H_{20}O_4$: C, 71.98; H, 6.71. Found: C, 71.85; H, 6.85.

The combined sodium bicarbonate filtrates from several reactions were acidified with concentrated hydrochloric acid and filtered. The filter cake was washed well with water and, while still damp, was recrystallized from acetonitrile to give from 3 to 10% yields of the acid VI, mp 228–231°, undepressed upon admixture with an authentic sample.

7,8,9,10-Tetrahydro-1-hydroxy-3-pentylspiro[6H-dibenzo[*b,d*]pyran-9,2'-[1',3']dioxolan]-6-one (VIII). A solution of 9.22 g (0.031 mole) of VII in 500 ml of benzene containing 10 ml of ethylene glycol and 10 mg of *p*-toluenesulfonic acid was heated overnight under reflux (Dean-Stark trap). The cooled solution was poured into water containing excess sodium bicarbonate. The organic layer was dried (Na_2SO_4) and evaporated to a yellow oil which, upon crystallization from dichloromethane-ether, gave 9.91 g (94%) of VIII as colorless crystals, mp 112–114°. Further recrystallization from ether gave the analytical sample: mp 114.5–116°; $\lambda_{\text{max}}^{\text{CHCl}_3}$ 228 m μ (inf) (ϵ 8400), 250 (inf) (9200), 257 (10,400), 304 (14,200); $\nu_{\text{max}}^{\text{CHCl}_3}$ 1695 cm^{-1} ; nmr (CDCl_3), δ 6.65 (two protons), no other unsaturated protons.

Anal. Calcd for $C_{26}H_{24}O_4$: C, 69.75; H, 7.02. Found: C, 69.61; H, 7.31.

Occasionally on standing, solutions of VIII deposited crystals, mp ~110–140°. This material, on recrystallization from methanol, was reconverted to material with mp 114.5–116°. However, on recrystallization from ether it was also converted to a polymorphic modification of VIII whose analytical sample had mp 145–148°; the infrared, ultraviolet, and nmr spectra were identical with those of the 114.5–116° modification.

Anal. Calcd for $C_{26}H_{24}O_4$: C, 69.75; H, 7.02. Found: C, 69.69; H, 7.40.

dl-6 α ,7-Dihydro-1-hydroxy-6,6-dimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-9(8H)-one (IX). A slurry of 60.0 g (0.175 mole) of VIII in 1.5 l. of ether was added over 90 min to the Grignard reagent prepared from 44.6 g (1.84 g-atoms) of magnesium and 110 ml (251 g, 1.77 moles) of methyl iodide in 1.8 l. of ether. After refluxing for 2 days the reaction was treated carefully with 200 ml of 1 *N* hydrochloric acid, the color changing from yellow to red-

brown to green, and then with 740 ml of 6 *N* hydrochloric acid. The mixture was stirred vigorously for 1 hr and then the ether layer was washed once with water and once with 5% sodium bicarbonate, the color changing from green to yellow-brown. The ether layer was dried (Na_2SO_4) and concentrated with stirring under a nitrogen stream on the steam bath to 500 ml. The resulting precipitate was filtered to give 38.4 g (70%) of IX as pale yellow crystals, mp 193–195°, suitable for the next step. Recrystallization from dichloromethane-ether gave the analytical sample as pale yellow crystals: mp 198–199°; $\lambda_{\text{max}}^{\text{CHCl}_3}$ 231 m μ (ϵ 16,000), 247 (inf) (5000), 325 (26,800); nmr (CDCl_3), δ 6.53 and 6.25 (two one-proton doublets, *J* = 2 cps, aromatic protons) and 8.07 (one-proton doublet, *J* = 2 cps, olefinic proton).

Anal. Calcd for $C_{28}H_{28}O_4$: C, 76.40; H, 8.34. Found: C, 76.27; H, 8.66.

dl-6 α ,7,10,10 α -Tetrahydro-1-hydroxy-6,6-dimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-9(8H)-one (Xa). A solution of 11.5 g (0.0366 mole) of IX in 100 ml of tetrahydrofuran was added slowly to a rapidly stirred solution of 400 mg of lithium in 500 ml of liquid ammonia (distilled through potassium hydroxide pellets) kept at –70°. Whenever the blue color began to fade, addition of IX was stopped and more lithium was added. Addition of IX was then resumed. This process was repeated until a permanent blue color persisted after the complete addition of IX. A total of 1.04 g (0.15 g-atom) of lithium was added. Stirring was continued for an additional 5 min and the blue color was discharged by the addition of ammonium chloride. The ammonia was allowed to evaporate and the residue was diluted with 500 ml of water, acidified with hydrochloric acid solution, and extracted with dichloromethane. The extracts were dried (Na_2SO_4) and concentrated to an oil. Crystallization from dichloromethane-hexane (until the mother liquors were free of either Xa or XI as shown by tlc) gave a total of 9.13 g of colorless crystals (79%) of a mixture of Xa and XI and little else (as shown by tlc). This material was then recrystallized from methanol-water to give 6.09 g (52.7%) of the *trans*-ketone Xa, free of the *cis*-ketone XI (thin layer chromatography), mp 162–164°. Further recrystallization from dichloromethane-hexane gave the analytical sample of Xa as colorless crystals: mp 163–165°; $\lambda_{\text{max}}^{\text{CHCl}_3}$ 230 m μ (inf) (ϵ 9800), 275 (inf) (1375), 283 (1490); $\nu_{\text{max}}^{\text{CHCl}_3}$ 1695 cm^{-1} .

Anal. Calcd for $C_{30}H_{28}O_4$: C, 75.91; H, 8.92. Found: C, 75.87; H, 8.89.

Several times the *trans*-ketone Xa was isolated as a lower melting polymorph whose analytical sample (from dichloromethane-ether) had mp 148–150°.

Anal. Calcd for $C_{30}H_{28}O_4$: C, 75.91; H, 8.92. Found: C, 76.19; H, 9.22.

The two samples had identical infrared, ultraviolet, and nmr spectra and identical behavior on tlc. In addition, the samples could be interconverted by judicious choice of recrystallization solvents and seeds.

The mother liquors from several of these reductions were combined and chromatographed over silica gel. Later benzene fractions contained essentially pure Xa. Recrystallization from methanol-water then gave an amount of Xa, mp 162–165°, corresponding to a total yield of Xa of 59%. Further elution with benzene and with mixtures of benzene and ether gave mixtures of Xa and XI, until 3:1 benzene-ether gave a small amount of solid which upon recrystallization from ether gave the *cis*-ketone XI as colorless crystals, mp 148–150°, undepressed upon admixture with an authentic sample of *cis*-ketone prepared from XX (see below).

dl-6 α ,7,10,10 α -Tetrahydro-1-hydroxy-6,6-dimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-9(8H)-one Acetate (Xb). A mixture of 1.00 g (3.2 mmoles) of Xa, 5 ml of pyridine, and 5 ml of acetic anhydride was heated for 15 min at 100°, at the end of which time tlc showed the reaction to be complete. The cooled solution was diluted with 100 ml of water, stirred for 30 min, and extracted with ether. The ether layer was extracted four times with water, dried (Na_2SO_4), and evaporated to 1.07 g of a semicrystalline residue. This was crystallized from hexane to give 0.80 g (71%) of Xb as colorless crystals, mp 86–91°. Further recrystallization from hexane gave the analytical sample: mp 88–91°; $\lambda_{\text{max}}^{\text{CHCl}_3}$ 227 m μ (inf) (ϵ 15500), 275 (2000), 282 (2180); $\nu_{\text{max}}^{\text{CHCl}_3}$ 1705 and 1765 cm^{-1} .

Anal. Calcd for $C_{32}H_{30}O_5$: C, 73.71; H, 8.44. Found: C, 73.63; H, 8.35.

dl-6 α ,7,8,9,10 α -Hexahydro-6,6-dimethyl-9-methylene-3-pentyl-6H-dibenzo[*b,d*]pyran-1-ol (XII). A slurry of sodium

(44) Thin layer chromatography of the total product mixtures showed that the reductions at –70° were much cleaner than those at –133°.

hydride (obtained by washing 1.52 g (0.032 mole) of 50% sodium hydride in mineral oil dispersion with dry hexane) in 60 ml of dimethyl sulfoxide was heated at 50° until solution took place (about 3 hr). After addition of 11.86 g (0.034 mole) of triphenylmethylphosphonium bromide, the reaction was heated for 3 hr at 70°. A solution of 1.00 g (0.0032 mole) of Xa in 60 ml of dimethyl sulfoxide was added and the reaction was heated overnight at 70°. The cooled reaction was poured into a mixture of ice and water containing 20 g of sodium bicarbonate. This was extracted with benzene and the organic layer was washed with water, dried (Na₂SO₄), and evaporated to 6.15 g of a tan oil containing triphenylphosphine oxide by thin layer chromatography. This oil was passed over silica gel in 1:1 hexane-benzene to give 1.57 g of colorless oil containing an odorous impurity. This oil was absorbed onto silica gel in hexane and the first hexane eluates contained the impurity. Later hexane eluates were evaporated to give 0.59 g (59%) of XII as a colorless oil: $\lambda_{\text{max}}^{\text{CHCl}_3}$ 230 m μ (inf) (ϵ 9620), 275 (1290), 283 (1300); nmr, see Table I.

Anal. Calcd for C₂₁H₂₆O₂: C, 80.21; H, 9.62. Found: C, 79.95; H, 9.91.

Attempted Isomerization of XII. A solution of 40 mg of XII and 3 mg of *p*-toluenesulfonic acid in 3 ml of benzene was heated for 15 min under reflux. The cooled reaction was poured into water containing an excess of sodium bicarbonate. The benzene layer and one benzene wash were combined, dried, and evaporated to an oil. This was evaporatively distilled at 135° (0.05 mm) to give 28 mg of an oil with an nmr spectrum identical with that of XII.

***dl*-6 α ,7,8,9,10,10 α -Hexahydro-6,6,9 β -trimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-1,9 α -diol (XIIIa).** To a refluxing solution of methylmagnesium iodide prepared from 10.50 g (0.43 g-atom) of magnesium and 27.0 ml (61.5 g, 0.43 mole) of methyl iodide in 650 ml of ether was added a solution of crude Xb prepared from 15.0 g (0.047 mole) of Xa dissolved in 500 ml of ether. The solution was heated for an additional hour under reflux and excess Grignard reagent was decomposed by the cautious addition of water. The reaction mixture was acidified with 3 *N* hydrochloric acid and the ether layer and several ether extracts were washed with water, sodium bicarbonate solution, and finally with water. The ether layer was dried (Na₂SO₄) and evaporated to a semisolid oil which on crystallization from ether-hexane gave 13.93 g (88%) of a mixture of carbinols suitable for conversion to the chloride XIV (see below). Recrystallization from ether-hexane gave 6.82 g (43%) of XIIIa as colorless crystals, mp 159.5–166°. Further recrystallization from ether-hexane gave the analytical sample: mp 162–163°; $\lambda_{\text{max}}^{\text{CHCl}_3}$ 231 m μ (inf) (ϵ 9500), 276 (1220), 282 (1270); $\nu_{\text{max}}^{\text{CHCl}_3}$ 3595 cm⁻¹, no carbonyl absorption.

Anal. Calcd for C₂₁H₂₈O₂: C, 75.86; H, 9.70. Found: C, 75.97; H, 9.88.

The same carbinol (XIIIa) could be obtained by the Grignard reaction with the tetrahydropyranyl ether of Xa followed by acid hydrolysis in 31% yield.

***dl*-9 α -Acetoxy-6 α ,7,8,9,10,10 α -hexahydro-6,6,9 β -trimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-1-ol (XIIIb).** A solution of 100 mg (0.3 mmole) of XIIIa in 1 ml of acetic anhydride and 1 ml of pyridine was heated for 3 hr on the steam bath.⁴⁴ In the cooled reaction was added 5 ml of water and stirring was continued for 15 min. The reaction was diluted with ether, washed four times with water, dried (Na₂SO₄), and evaporated to give the crude diacetate. This was mixed with 300 mg of sodium bicarbonate in 10 ml of methanol and 2 ml of water and heated for 15 min under reflux. The cooled reaction was diluted with 10 ml of water and filtered to give 85 mg (75%) of XIIIb as colorless crystals, mp 130–136°. Recrystallization from hexane gave the analytical sample: mp 137–139.5°; $\nu_{\text{max}}^{\text{CHCl}_3}$ 3600 and 1720 cm⁻¹.

Anal. Calcd for C₂₃H₃₀O₄: C, 73.76; H, 9.15. Found: C, 73.66; H, 9.24.

***dl*-6 α ,7,10,10 α -Tetrahydro-6,6,9-trimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-1-ol (*dl*- Δ^4 -Tetrahydrocannabinol) (II).** A solution

of 500 mg (1.5 mmoles) of XIIIa and 50 mg of *p*-toluenesulfonic acid in 50 ml of benzene was heated for 30 min under reflux (Dean-Stark trap). The cooled solution was passed through a small column of silica gel and the benzene eluates were evaporated to give 454 mg (96%) of II as a colorless oil (only one isomer by glpc and with spectra identical with those of the analytical sample).

To 35 ml of benzene containing two drops of pyridine was added 350 mg (1.1 mmoles) of II and one-third of the solvent was distilled. The solution was cooled to room temperature under a drying tube and 278 mg (1.3 mmoles) of 3,5-dinitrophenyl isocyanate⁴⁵ was added. After stirring overnight a small amount of solid was removed by filtration and the yellow filtrate was adsorbed onto Florisil. Elution with benzene gave a colorless eluate and a light yellow band on the upper portion of the column. Concentration of the eluate gave colorless crystals which, upon one recrystallization from ether-hexane, afforded 330 mg of the 3,5-dinitrophenylurethane of II, mp 208–209.5°. Recrystallization of this material was not successful and gave a product with a lower melting point. Therefore no further attempts were made to characterize II via the 3,5-dinitrophenylurethane and 180 mg of this solid was hydrolyzed by stirring with 1.1 ml of 1 *N* sodium hydroxide solution in 50 ml of methanol overnight. The solution was poured into a mixture of ether and excess dilute hydrochloric acid. The ether layer was washed with water, dried (Na₂SO₄), and concentrated. The residue was dissolved in benzene and passed over Florisil. The upper portion of the column was bright yellow and the colorless benzene eluate was evaporated to give 90 mg of the analytical sample of II as a colorless oil: $\lambda_{\text{max}}^{\text{CHCl}_3}$ 231 m μ (inf) (ϵ 8820), 275 (1160), 283 (1190); $\nu_{\text{max}}^{\text{CHCl}_3}$ 3600, 1620, and 1575 cm⁻¹; nmr, see Table I.

Anal. Calcd for C₂₁H₂₆O₂: C, 80.27; H, 9.62. Found: C, 80.30; H, 9.73.

***dl*-9-Chloro-6 α ,7,8,9,10,10 α -hexahydro-6,6,9-trimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-1-ol (XIV).** To a solution of 7.10 g (0.021 mole) of a mixture of carbinols (see the preparation of XIIIa) in 100 ml of glacial acetic acid was added a solution of 72 g of zinc chloride in 56 ml of concentrated hydrochloric acid. After stirring for 3 hr at room temperature the reaction mixture was poured into a mixture of 2 l. of water and 2 l. of ether. The ether layer was washed five times with 1-l. portions of water, dried (Na₂SO₄), and evaporated to a yellow oil. Crystallization from hexane gave 5.53 g (74%) of XIV as colorless crystals, mp 85–90°. Recrystallization from hexane gave the analytical sample as colorless crystals: mp 87–90°; $\lambda_{\text{max}}^{\text{CHCl}_3}$ 230 m μ (inf) (ϵ 9700), 276 (1200), 277 (1230).

Anal. Calcd for C₂₁H₂₅ClO₂: C, 71.88; H, 8.90; Cl, 10.10. Found: C, 71.92; H, 9.02; Cl, 10.06.

Similarly, purified carbinol XIIIa gave XIV in comparable yields.

The Preparation of XIV Directly from Xa. A slurry of 10.00 g (0.0316 mole) of Xa in 100 ml of ether was added over 15 min to a solution of methylmagnesium iodide prepared from 7.0 g (0.29 g-atom) of magnesium and 18.0 ml (41.0 g, 0.29 mole) of methyl iodide in 500 ml of ether. The reaction was heated for 90 min under reflux and cooled in an ice bath. Water was added carefully to decompose the excess Grignard reagent and enough 6 *N* hydrochloric acid was added to give two clear layers. The organic layer was diluted with 600 ml of ether and washed twice with water and finally with brine. The solution was dried (Na₂SO₄) and concentrated. To the residue dissolved in 150 ml of glacial acetic acid was added a solution of 103 g of zinc chloride in 80 ml of concentrated hydrochloric acid. After stirring for 90 min at room temperature the reaction mixture was poured into a mixture of 3 l. of water and 2.4 l. of ether. The ether layer was washed five times with 1.5-l. portions of water, dried (Na₂SO₄), and evaporated. The residue was dissolved in benzene and adsorbed on a column of silica gel. The column was eluted with benzene until thin layer chromatograms showed that no more XIV was being eluted. The column was then eluted with ether and these ether eluates were worked up to give 3.64 g of recovered Xa. The benzene eluates were evaporated and crystallized from hexane to give 3.45 g (49%, based on unrecovered Xa) of XIV as light tan crystals, mp 85–88°, identical with the material obtained directly from the carbinol XIIIa.

***dl*- Δ^4 -Tetrahydrocannabinol (I) and *dl*- Δ^4 -Tetrahydrocannabinol (II).** To a slurry of the sodium hydride from 1.00 g (0.02 mole) of 50% sodium hydride in mineral oil dispersion (washed free of mineral oil with dry hexane) in 200 ml of tetrahydrofuran (distilled from lithium aluminum hydride) was added 1.00 g (0.00285 mole) of XIV. The reaction was heated overnight under reflux with stirring, cooled to room temperature, and poured carefully into a

(46) C. Naegeli and A. Tyabji, *Helv. Chim. Acta*, 16, 349 (1933).

mixture of water and ether. The ether layer was washed four times with 500-ml portions of water, dried (Na_2SO_4), and concentrated to 875 mg (98%) of a yellow oil. Vapor phase chromatographic analysis of this oil showed the presence of I and II in the ratio 74:26 accompanied by only traces of other components.

dl-6 α ,7,8,10 α -Tetrahydro-6,6,9-trimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-1-ol *m*-Nitrobenzenesulfonate (XV). To a solution of 500 mg (1.6 mmoles) of the 74:26 mixture of I and II in 10 ml of dry pyridine was added 700 mg (3.2 mmoles) of *m*-nitrobenzenesulfonyl chloride. The solution was heated for 8 hr at 60° and then for 4 hr at 75° at which time the indicated reaction was complete. The cooled reaction was poured into a mixture of ether and dilute hydrochloric acid. The ether layer was washed with dilute hydrochloric acid, water, dilute sodium bicarbonate, and finally with water, dried (Na_2SO_4), and evaporated. The residue was recrystallized repeatedly from methanol to give 180 mg (23%) of XV as very pale yellow crystals, mp 106–107.5°. The analytical sample had mp 105.5–107.5°; $\lambda_{\text{max}}^{\text{calc}}$ 230 m μ (inf) (ϵ 11,600), 247 (7650), 280 (inf) (ϵ 3950); nmr (CDCl_3), 6.60 (one-proton doublet, $J = 2$ cps), 6.38 (one-proton doublet, $J = 2$ cps), 6.05 (one-proton broad singlet).

Anal. Calcd for $\text{C}_{21}\text{H}_{23}\text{NO}_6\text{S}$: C, 64.91; H, 6.66; N, 2.80; S, 6.42. Found: C, 64.64; H, 6.36; N, 2.70; S, 6.78.

dl-6 α ,7,8,10 α -Tetrahydro-6,6,9-trimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-1-ol (*dl*- Δ^8 -Tetrahydrocannabinol) (I). A solution of 1.69 g (3.4 mmoles) of XV in 600 ml of methanol containing 25 ml of 1 *N* sodium hydroxide was heated under reflux until the indicated reaction was complete (45 min). The cooled reaction was poured into ether and dilute hydrochloric acid. The organic layer was washed with water, dilute sodium bicarbonate, and finally with water. It was dried and evaporated to a pale tan oil which on crystallization and recrystallization from hexane gave 0.80 g (84%) of I as very light tan crystals, mp (vac) 60–62°. The analytical sample had mp (vac) 64.5–65.5°; the glpc retention time (no detectable impurities) and infrared, ultraviolet, and nmr spectra were identical with natural Δ^8 -tetrahydrocannabinol; for nmr data, see Table I; $\lambda_{\text{max}}^{\text{calc}}$ 230 m μ (inf) (ϵ 10,000), 275 (1250), 282 (1280); $\nu_{\text{max}}^{\text{calc}}$ 3595, 1620, 1575 cm^{-1} .

Anal. Calcd for $\text{C}_{21}\text{H}_{28}\text{O}_2$: C, 80.21; H, 9.62. Found: C, 80.34; H, 9.54.

5-Hydroxy-2,2-dimethyl-7-pentyl-4-chromanone (XVII) and 7-Hydroxy-2,2-dimethyl-5-pentyl-4-chromanone (XVIII). A mixture of 73 g (0.4 mole) of olivetol (III) and 47 g (0.47 mole) of 3-methylcrotonic acid (XVI) was heated in an oil bath to 125°. Through the condenser, 94 ml of boron trifluoride etherate was added and the solution was heated overnight under reflux. The partially cooled reaction was diluted cautiously with 100 ml of water followed by 425 ml of 6 *N* sodium hydroxide and then boiled for 5 min on the steam bath. The solution was cooled, acidified with 6 *N* hydrochloric acid, and extracted with ether. The ether extracts were washed with water, dilute sodium bicarbonate, and water and then dried (Na_2SO_4) and concentrated to 106 g of a tan oil. This was dissolved in ether and extracted repeatedly with 1 *N* sodium hydroxide solution. The ether layer, upon drying and evaporation, gave 53.1 g (50%) of crude XVII as a light tan oil suitable for the next step. Repeated recrystallization from methanol at –70° gave the analytical sample of XVII as tan crystals: mp 23–27°, normally handled as an oil; $\lambda_{\text{max}}^{\text{calc}}$ 229 m μ (inf) (ϵ 11,780), 280 (13,800), 348 (2980); $\lambda_{\text{max}}^{\text{calc}}$ (in ethanolic sodium acetate) 280 (13,450), 348 (2940); $\lambda_{\text{max}}^{\text{calc}}$ 242 (14,200), 285 (11,150), 373 (5250); $\nu_{\text{max}}^{\text{calc}}$ 1645 cm^{-1} ; nmr (CDCl_3), 11.73 (one-proton singlet, hydroxylic proton), 6.33 and 6.25 (two one-proton doublets, $J = \sim 1$ cps, aromatic protons), 2.72 (two-proton singlet, C-3 H's), 2.52 (two-proton triplet, α methylene), 1.45 (six-proton singlet, 2,2-bis CH_3), and 0.88 (three-proton triplet, ω - CH_3).

Anal. Calcd for $\text{C}_{21}\text{H}_{28}\text{O}_3$: C, 73.25; H, 8.45. Found: C, 73.02; H, 8.75.

The 1 *N* sodium hydroxide extracts of the total reaction mixture were acidified with hydrochloric acid and extracted with ether. The extracts were dried and concentrated. Crystallization from acetonitrile gave 23.1 g (22%) of crude XVIII as tan crystals, mp 82–86°. Recrystallization from hexane gave the analytical sample of XVIII as colorless crystals: mp 94.5–96°; $\lambda_{\text{max}}^{\text{calc}}$ 220 m μ

(ϵ 13,640), 233 (9400), 280 (11,300), 315 (4700); $\lambda_{\text{max}}^{\text{calc}}$ (in ethanolic sodium acetate), 285 (6250), 328 (13,600); $\lambda_{\text{max}}^{\text{calc}}$ 252 (6650) and 328 (19,500); $\nu_{\text{max}}^{\text{calc}}$ 3590 and 1670 cm^{-1} ; nmr (CDCl_3), 8.77 (one-proton singlet, hydroxylic proton), 6.35 (two protons, pair of doublets, aromatic protons), 2.70 (two-proton singlet, C-3 H's), 3.02 (two-proton triplet, α methylene), 1.42 (six-proton singlet, 2,2-bis CH_3), and 0.87 (three-proton triplet, ω - CH_3).

Anal. Calcd for $\text{C}_{21}\text{H}_{28}\text{O}_3$: C, 73.25; H, 8.45. Found: C, 72.98; H, 8.72.

5-Hydroxy-3-hydroxymethylene-2,2-dimethyl-7-pentyl-4-chromanone (XIX). To the sodium hydride obtained by washing 78.5 g (1.64 moles) of 50% sodium hydride in mineral oil dispersion with hexane was added (at room temperature) a solution of 46.1 g (0.175 mole) of XVII in 172 ml (159 g, 2.15 moles) of ethyl formate, utilizing a Dry Ice condenser during the addition. This condenser was then replaced by a water-jacketed condenser. Ether (4.4 l.) was added and after heating overnight under reflux the reaction was cooled and acidified with 1 *N* hydrochloric acid. The ether layer and two ether washes of the aqueous layer were washed with 1 *N* hydrochloric acid and twice with water and then extracted five times with 1 *N* sodium hydroxide. These basic extracts were acidified and extracted with ether. The ether was dried (Na_2SO_4) and concentrated to 27.5 g (54%) of crude XIX as a yellow oil suitable for the next step. Crystallization and recrystallization from hexane gave the analytical sample as yellow crystals: mp 31–32.5°; $\lambda_{\text{max}}^{\text{calc}}$ 223 m μ (inf) (ϵ 16,400), 274 (4900), 331 (15,650), 366 (inf), (8480), 386 (inf) (4200).

Anal. Calcd for $\text{C}_{21}\text{H}_{28}\text{O}_4$: C, 70.32; H, 7.64. Found: C, 70.60; H, 7.89.

The Preparation of IX from XIX. To a solution of 8.33 g (0.029 mole) of crude XIX in 43 ml of methanol and 4.25 ml of methyl vinyl ketone was added 1.0 ml of triethylamine. After standing overnight at room temperature, the solution was diluted with 500 ml of ether and extracted four times with 10% sodium carbonate solution. The ether solution was dried (Na_2SO_4) and concentrated. The residue (8.43 g) was heated overnight under reflux with 84.3 ml of ethanol and 84.3 ml of 2 *N* potassium hydroxide solution. The cooled solution was acidified with 6 *N* hydrochloric acid and extracted with dichloromethane. The extracts were dried (Na_2SO_4) and concentrated, and the residue was crystallized from dichloromethane–ether to give 4.11 g (35%) of IX as yellow crystals, mp 195–198°. Further recrystallization gave pale yellow crystals, mp 197.5–199°, undepressed on admixture with a sample of IX prepared from VII.

dl-6 α ,7,8,9,10,10 α -Hexahydro-1-hydroxy-3-pentylspiro[611-dibenzo[*b,d*]pyran-9,2'-[1',3']dioxolan]-6-one (XX). A solution of 5.00 g (0.0145 mole) of VII in 100 ml of ether was subjected to hydrogenation at 2150-lb pressure and 130° for 2 hr in the presence of 0.5 g of Raney nickel. The catalyst-free solution was evaporated to a crystalline residue which, on recrystallization from ether–hexane, gave 3.53 g (70%) of XX as colorless crystals, mp 142.5–143.5°. A subsequent reduction under apparently identical conditions on a 50-g scale gave a 33% yield of XX. Further recrystallization from ether–hexane gave the analytical sample: mp 143–144°; $\lambda_{\text{max}}^{\text{calc}}$ 275 m μ (inf) (ϵ 2400), 282 (2790).

Anal. Calcd for $\text{C}_{20}\text{H}_{26}\text{O}_2$: C, 69.34; H, 7.57. Found: C, 69.02; H, 7.30.

dl-6 α ,7,10,10 α -Tetrahydro-1-hydroxy-6,6-dimethyl-3-pentyl-611-dibenzo[*b,d*]pyran-9(8H)-one (XI). A slurry of 3.60 g (0.0104 mole) of XX in 180 ml of ether was added (argon atmosphere) over 30 min to a methylmagnesium iodide solution prepared from 11.1 ml (25.3 g, 0.18 mole) of methyl iodide and 4.32 g (0.18 g-atom) of magnesium in 180 ml of ether. The reaction was heated overnight under reflux and 1 *N* hydrochloric acid was added carefully until two clear layers were obtained. The aqueous layer was washed with dichloromethane, and the combined organic layers were concentrated. The residue was mixed with 80 ml of methanol and 40 ml of 3 *N* hydrochloric acid and heated for 2 hr under reflux. The cooled reaction was diluted with 500 ml of water and extracted with dichloromethane. The extracts were dried (Na_2SO_4) and concentrated to a crystalline residue which on recrystallization from ether gave 1.73 g (53%) of colorless crystals, mp 148–150°. Further recrystallization from ether–hexane gave the analytical sample: mp 149.5–150.5°; $\lambda_{\text{max}}^{\text{calc}}$ 230 m μ (inf) (ϵ 9800), 277 (1570), 283 (1620).

Anal. Calcd for $\text{C}_{21}\text{H}_{28}\text{O}_2$: C, 75.91; H, 8.92. Found: C, 76.01; H, 8.88.

dl-6 α ,7,8,9,10,10 α -Hexahydro-6,6-dimethyl-9-methylene-3-pentyl-6H-dibenzo[*b,d*]pyran-1-ol (XXI). A slurry of sodium hydride (obtained by washing 3.04 g (0.063 mole) of 50% sodium

(47) The vapor phase chromatographic analyses were done on an F & M Model 810R-19 instrument with dual hydrogen flame detectors. The column was 0.25 in. $\times$ 6 ft packed with 0.5% NPGA + 0.5% PEG4000MS on Anakrom ABS (60–70 mesh). The column flow was 100 cc/min of nitrogen, the column temperature was 220°, and the retention time of I was 20 min and that of II was 18 min.

hydride in mineral oil dispersion with dry hexane) in 120 ml of dimethyl sulfoxide was heated at 50° until solution took place (about 30 min). After addition of 22.6 g (0.063 mole) of triphenylmethylphosphonium bromide the reaction was heated for 3 hr at 70°. A solution of 2.00 g (0.0063 mole) of XI in 120 ml of tetrahydrofuran was added and the reaction was heated overnight under reflux. The cooled reaction was poured into a mixture of ice and water containing 40 g of sodium bicarbonate. This was extracted with benzene and the extracts were dried (Na₂SO₄) and concentrated to a semicrystalline residue. Addition of ether and filtration gave two crops of triphenylphosphine oxide which contained no XXI by thin layer chromatography and were discarded. The evaporated mother liquors were passed over silica gel in 1:1 hexane-benzene. The eluates were concentrated and crystallized from hexane at Dry Ice temperature to give 0.94 g (47%) of XXI as tan crystals, mp 35–50°. Further recrystallization gave the analytical sample as colorless crystals: mp 48–52°; $\lambda_{\text{max}}^{\text{CHCl}_3}$ 230 m μ (inf) (ϵ 9500), 277 (1200), 284 (1230); nmr, see Table I.

Anal. Calcd for C₂₁H₂₀O₂: C, 80.21; H, 9.62. Found: C, 80.26; H, 9.21.

dl-6 α ,7,8,9,10,10 α -Hexahydro-6,6,9-trimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-1,9-diol (XXII). A slurry of 500 mg (1.58 mmoles) of XI in 10 ml of ether was added to a solution of methylmagnesium iodide prepared from 106 mg (4.4 mg-atoms) of magnesium and 0.30 ml (0.68 g, 4.8 mmoles) of methyl iodide in 10 ml of ether to give a clear reaction mixture. After heating for 1 hr under reflux the reaction was cooled in an ice bath and 1 *N* hydrochloric acid was added until two clear layers were formed. The organic layer was diluted with ether and washed twice with water, once with dilute sodium bicarbonate, and again with water. The ether solution was dried (Na₂SO₄) and concentrated to an oil which on crystal-

lization from hexane gave 380 mg (72%) of XXII as colorless crystals, mp 142–145°. Recrystallization from ether-hexane gave the analytical sample: mp 142–145°; $\lambda_{\text{max}}^{\text{CHCl}_3}$ 230 m μ (inf) (ϵ 10,000), 278 (1420), 283 (1450).

Anal. Calcd for C₂₁H₂₂O₂: C, 75.86; H, 9.70. Found: C, 76.07; H, 9.83.

dl-6 α ,7,8,10 α -Tetrahydro-6,6,9-trimethyl-3-pentyl-6H-dibenzo[*b,d*]pyran-1-ol (XXIII). A solution of 828 mg (2.5 mmoles) of XXII and 50 mg of *p*-toluenesulfonic acid in 50 ml of benzene was heated for 30 min under reflux. The cooled solution was washed with dilute sodium bicarbonate, dried (Na₂SO₄), and evaporated to give 770 mg (98%) of XXIII as a colorless oil. A small portion of this oil was evaporatively distilled at 135° (0.05 mm) to give the analytical sample: $\lambda_{\text{max}}^{\text{CHCl}_3}$ 232 m μ (inf) (ϵ 9850), 275 (1470), 282 (1500); nmr, see Table I.

Anal. Calcd for C₂₁H₂₀O₂: C, 80.21; H, 9.62. Found: C, 79.87; H, 9.47.

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Photochemistry of Nucleic Acids. III. The Structure of DNA-Derived Thymine Photodimer¹

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Contribution from the University Chemical Laboratory, Lensfield Road, Cambridge, England. Received June 29, 1967

Abstract: The identity of the major thymine photoproduct derived from DNA is established with the *cis-syn* dimer 1 obtained from irradiation of thymine in ice. The ¹⁴C-labeled photoproduct was mixed with unlabeled ice-dimer and the cocrystallized material treated with bromine in alkali. The resulting stereospecific rearrangement reaction gave *cis,cis,cis*-3-carbamido-1,7-dimethyl-3,5,0-triazatricyclo[5.3.0.0^{2,4}]deca-4,8,10-trione (7) retaining more than 91% of the specific activity of the recrystallized dimer. The mechanism of the rearrangement was investigated by performing the reaction in oxygen-18-enriched solvent. The observed nonincorporation of this isotope into the product 7 is consistent with a mechanism involving isocyanate formation without nucleophilic attack of hydroxide.

Good evidence exists^{3,4} that photoproducts of thymine and of cytosine are responsible for the majority of photobiological effects caused by ultraviolet irradiation of DNA. The formation of pyrimidine dimers is particularly important in this respect and, in principle, they can arise either by photoaddition of adjacent pyrimidines in the same DNA strand—intrastrand dimerization or from bonding between two neighboring

pyrimidines, one in each strand, giving interstrand dimers and thus effecting cross-linking between the complementary DNA chains.

These two modes are expected to lead to dimers of different stereochemistry, and the inspection of molecular models resulted in the prediction⁵ that intrastrand dimers should have *cis-syn* stereochemistry whereas interstrand dimers would give rise to either *cis-anti* or to *trans-anti* products. An alternative opinion⁶ has suggested that such cross-linking would afford *cis-anti* or *trans-syn* interstrand dimers.

(1) Paper II: G. M. Blackburn and R. J. H. Davies, *J. Chem. Soc., C*, 2239 (1966).

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(3) Abbreviations used: DNA, deoxyribonucleic acid; t-RNA, transfer ribonucleic acid; T, thymine; TT, thymine dimer; UT, uracil-thymine dimer.

(4) R. B. Setlow and W. L. Carrier, *J. Mol. Biol.*, 17, 237 (1966).

(5) G. Fraenkel and D. L. Wulff, *Biochim. Biophys. Acta*, 51, 332 (1961).

(6) C. Nagata, A. Inamura, Y. Taniguchi, M. Kodama, and N. Fukada, *J. Theoret. Biol.*, 9, 357 (1965).

Blackburn, Davies / DNA-Derived Thymine Photodimer Structure

The stability of cannabis and its preparations on storage

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Solutions of pure cannabinoids, nine samples of herbal and two of resin cannabis (one freshly prepared) were stored in varying conditions for up to 2 years. Exposure to light (not direct sunlight) was shown to be the greatest single factor in loss of cannabinoids especially in solutions, which should therefore be protected from light during analytical and phytochemical operations. Previous claims that solutions in ethanol were stable have not been substantiated. The effect of temperature, up to 20°, was insignificant but air oxidation did lead to significant losses. These could be reduced if care was taken to minimize damage to the glands which act as "well filled, well closed containers". Loss of tetrahydrocannabinol after exposure to light does not lead to an increase in cannabidiol, but air oxidation in the dark does. It is concluded that carefully prepared herbal or resin cannabis or extracts are reasonably stable for 1 to 2 years if stored in the dark at room temperature.

It has long been recognized that cannabis resin and herbal cannabis lose potency on storage and this instability is said to be one reason against the use of cannabis in medicine in the past. Since the recognition that potency is mainly due to the content of Δ^9 -trans-tetrahydrocannabinol (THC) some work on resin and herb has indeed confirmed that the THC content falls on storage (Lerner, 1969; Maunder, 1970; Mechoulam, 1970; Schou & Nielsen, 1970). Three reports indicate that ethanol may exert a marked stabilizing effect (Eckler & Miller, 1917; Kubena, Barry & others, 1972; Razdan, Puttick & others, 1972). Little of the earlier work was done on carefully dated samples, with adequate methods of analysis, nor were varying conditions of storage examined in detail. In 1971 we therefore began experiments designed to test the effect of the obvious factors of light, oxygen and temperature and to include in our consideration the fact that in the plant the active resin is stored in glands which are virtually "well-filled, well-closed containers". When preparing resin cannabis and some forms of herbal cannabis, such as ganja, many of these glands will be burst and the released resin will be exposed to air oxidation. Extracts and solutions in various solvents were also examined as these preparations are likely to be handled during phytochemical work.

Since commencing our work Turner, Hadley & others (1973a) published results of experiments covering 1970-72; they used a wider range of temperature than we did but did not study the effect of light in detail: also their results were based on one sample of herb only. Work has also been carried out on short exposure of cannabis leaf to relatively high temperatures (Coffman & Gentner, 1974) and the stability of extracts and pure cannabinoids in solvents (Turner, Hadley & Davis, 1973b; Parker, Borke & others, 1974; Turner & Henry, 1975). Garrett & Tsau (1974) reported on the effect of pH during storage; however, they used extremely dilute solutions of THC in water (0.5 ppm) and do not seem to have considered the effect of light.

MATERIALS AND METHODS

Herbal cannabis. Samples were prepared from our own plants grown in England; those labelled "UN etc." were from seeds supplied by the UN Division of Narcotics, Geneva; SP1 was from seeds purchased in Merzifon, Turkey, 1969; SP4, a Nepalese variety; SP5, from a police seizure in London, 1971; SP10, a Mexican strain and SP11, seeds from the Cameroons, 1972. The leaves and tops, freed from stems more than 2 mm in diameter, were dried in the shade in a current of air at about 20°. For analytical purposes it was not possible to sample adequately without powdering the heterogeneous material; unfortunately this obviously burst some of the glands. To get some idea therefore of this effect, coarsely and finely powdered samples from the same material were stored separately.

Cannabis resin. One batch of Pakistani resin which had been seized by the police in August 1968 was used. In addition we prepared a sample from the dried tops of one of our varieties (South African, UNC 335, collected in November, 1973). The tops which contained 3.64% THC were frozen to make the glands hard and brittle and then coarsely powdered. This powder was rapidly sieved through sieves of varying fineness. Those fractions coarser than 180 µm mesh aperture width contained 1.9 to 3.7% THC; that fraction passing completely through 125 µm mesh contained 11.6% THC. Microscopical examination showed the presence of numerous cystoliths and covering trichomes as well as glands, most of which were intact. Some of this fine powder was stored loose with occasional shaking to keep the powder from clumping; another part was compressed into blocks to simulate the method used for making "commercial" resin.

Pure cannabinoids were obtained from Professor R. Mechoulam or from Makor Chemicals, Jerusalem. The Δ⁹-THC showed a single peak on the g.l.c. system used; the Δ⁸-THC showed some impurities representing about 10% of the area of the major peak. The cannabinol (CBN) and cannabidiol (CBD) were pure to g.l.c.

Analysis was carried out by the method of Fairbairn & Liebmann (1973). However, for the work on the effect of light on pure THC, triacontane was used as internal standard because it is more stable than the steroid normally used. The problem of sampling resin has been discussed earlier (Fairbairn & Liebmann 1973); for our present purpose samples 5 × 3 × 3 mm were cut from the block on each occasion. Each piece was then cut into three equal parts by slicing parallel to the original surfaces of the entire block so that two segments contained the outer part and the third the interior of the block only.

Storage conditions

Samples were stored in transparent glass bottles at 5° in the dark (refrigerator) at room temperature (about 20°) in a dark cupboard and at room temperature behind a north-facing window. The samples were kept well mixed and portions were removed periodically for analysis.

RESULTS

The results for cannabinoids in solution are given in Table 1 and Figs 1A and 1B. One striking feature not recorded in Table 1 is the absence of any evidence for the production of cannabinol (CBN). The results for herbal cannabis are given in Tables 2 and 3 and for our own prepared cannabis resin in Table 4. For the commercial

Table 1.

1.	Δ ⁹ -THC
2.	Δ ⁸ -THC
3.	THC in
4.	CBD in
5.	CBN in
6.	CHCl ₃ e
7.	CHCl ₃ e

(a) CBN c
(b) CBN c

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Table 1. Cannabinoids in solution; loss during different conditions of storage. Figures are the percentages of the main cannabinoid retained; absolute values, at the commencement (t_0) are given in brackets, as mg ml^{-1} .

	5° in dark	20° in dark	20° to light
1. $\Delta^9\text{-THC}$ in petroleum t_0 —(0.08) $t_0 + 2$ days $t_0 + 6$ days	94 92	95 94	80 9
2. $\Delta^9(0)\text{-THC}$ in petroleum t_0 —(0.15) $t_0 + 2$ days $t_0 + 6$ days	100 99	100 101	51 1
3. THC in CHCl_3 , t_0 —(1.30) $t_0 + 2$ days $t_0 + 6$ days $t_0 + 20$ days		99 103 91	92 63 9
4. CBD in CHCl_3 , t_0 —(1.0) $t_0 + 2$ days $t_0 + 6$ days $t_0 + 20$ days		69 32 traces	47 5 none
5. CBN in CHCl_3 , t_0 —(1.32) $t_0 + 2$ days $t_0 + 6$ days $t_0 + 20$ days		95 87 87	99 67 48
6. CHCl_3 extract of UNC 255 t_0 —(THC 1.52) $t_0 + 2$ days $t_0 + 6$ days $t_0 + 20$ days		103 ^a 103 101	67 ^b 15 none
7. CHCl_3 extract of SP1 t_0 —(CBD 0.38) $t_0 + 2$ days $t_0 + 6$ days $t_0 + 20$ days		100 95 103	92 61 8

(a) CBN remained at 0.12, 0.12 and 0.09 respectively.

(b) CBN concentration at t_0 was 0.12 mg ml^{-1} and rose to 0.22, 0.39 and 0.31 respectively.

sample of resin the sampling difficulties already referred to made it impossible to carry out adequate storage tests. In general however, (a) the inner layers contained more THC than the surface layers; (b) in any given piece a low THC content corresponded to a high CBN and vice versa; (c) in the inner layers the total THC + CBN was higher than in the surface layers. Thus for the inner layers the average THC and CBN contents were 4.5 and 2.4% respectively giving a total of 6.9%; in the surface layers the corresponding figures were 2.5, 3.4, with a total of 5.9%. In contrast to these fluctuations, the CBD content was surprisingly stable giving the same average of 4.1% for the inner and surface layers.

DISCUSSION

The most important result of this work has been to emphasise the deleterious effect of exposure to light, a point not sufficiently stressed previously. The effect on solutions of cannabinoids is dramatic. Pure THC in light petroleum is almost completely decomposed in 6 days at room temperature, whereas in the dark there is

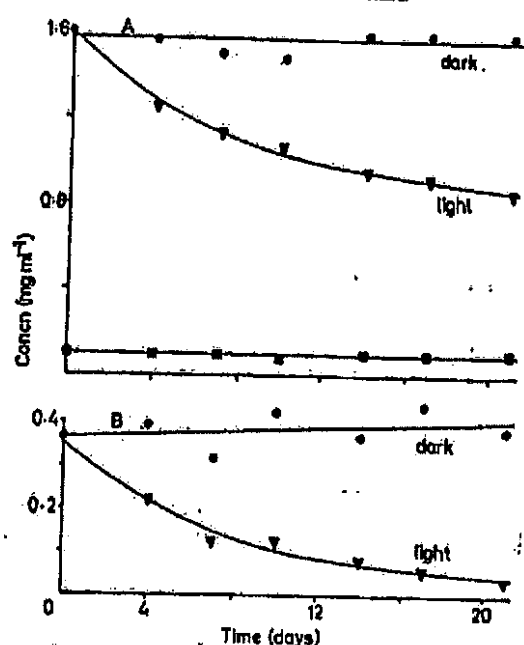


FIG. 1.A. Ethanol extract of UNC 235; effect on THC content of storage in the dark (●) and in the light (▼). CBD content, in dark and light (■). B. Ethanol extract of SP1; effect on CBD content of storage in the dark (●) and in the light (▼).

practically no loss (Table 1, samples 1 and 2). In CHCl₃ solutions decomposition is less marked (sample 3) but the presence of pigments in crude extracts does not seem to have any protective effect against light (sample 6). CBD in CHCl₃ is markedly less stable even in the dark (sample 4), and this confirms the results of Parker & others (1974) which showed significant loss in 1 day and almost total loss in 8 days in the dark. However, in crude samples CBD is very stable in the dark, but once more the pigments present do not protect from the deleterious effect of light (sample 7). This stability in crude extracts in the dark is confirmed by our previous work on commercial resin. Turner & others (1973b) report that THC and CBD in crude extracts in CHCl₃ are stable for 6 days, but unfortunately they do not state whether storage was

Table 2. Herbal cannabis: effect of temperature and light on one sample (SP4). Figures are % remaining in each sample (100 = 1.37% THC).

Storage time	5° in dark	20° in dark	20° in light
t ₀	100	100	100
t ₀ + 31 weeks	92	88	65
t ₀ + 47 "	93	87	64
t ₀ + 73 "	89	80	50
t ₀ + 98 "	91	75	37

Table 3. Herbal cannabis: effect of temperature and light on one sample (SP4). Figures are % remaining in each sample (100 = 1.37% THC).

Sample	5° in dark	20° in dark	20° in light
SP5			
SP10			
SP11			
UNS1			
UNC235			
Sample (a)			
Sample (b)			
UNC234			
UNC335			
UNC238			
Sample (a)			
Sample (b)			
Sample (c)			

n.d. Not determined.

in the light or in the dark. CBD, either in solution and artificial light (1974) may be of dual qualities of chlorophyll and for problem raised by the assay of cannabis procedure is short place, but obviously before completing the ethanol extract: fairly rapid loss of other solvents but of ethanol extract (1 original activity after to 38°. Obviously reference to the U.S. ing tops of pistillate of the fact that flower & Liebmann, 1974), ably from India) w

Table 3. *Herbal cannabis: effect of storage at 20° in the dark of eight varieties.* Figures are % of the main cannabinoid retained, absolute values at the commencement (t_0) are given in brackets.

Variety	Time	Coarse powder (about 1 mm mesh)	Fine powder (about 0.33 mm mesh)
SP5	t_0 (2.65% THC) $t_0 + 1$ year	83	74
SP10	t_0 (1.46% THC) $t_0 + 1$ year	92	80
SP11	t_0 (2.01% THC) $t_0 + 1$ year	74	68
UNSI	t_0 (3.43% THC) $t_0 + 1$ year	91	84
UNC255			
Sample (a)	t_0 (1.90% THC) $t_0 + 2$ years	n.d.	52
Sample (b)	t_0 (4.60%) $t_0 + 1$ year	n.d.	57
UNC254	t_0 (1.05% THC) $t_0 + 2$ years	n.d.	54
UNC335	t_0 (4.50%)	n.d.	69
UNC258			
Sample (a)	t_0 (1.37% CBD) $t_0 + 1$ year	86	78
Sample (b)	t_0 (1.55% CBD) $t_0 + 1$ year	87	73
Sample (c)	t_0 (0.21% CBD) $t_0 + 2$ years	n.d.	76

n.d. Not determined.

in the light or in the dark. Later Turner & Henry (1975) reported that THC and CBD, either in solution or in crude extract form, are stable for 6 days in both natural and artificial light. They suggest that the instability reported by Parker & others (1974) may be due to their having used impure CHCl_3 . We have used varying qualities of chloroform, including spectrograde used by Parker & Turner and their colleagues and found in all cases decomposition is rapid in the light. The main problem raised by our results and those of Parker and others is the use of CHCl_3 in the assay of cannabis (e.g., Fairbairn & Liebmann, 1973). Fortunately the extraction procedure is short (about 1 h at the most) so that little decomposition will take place, but obviously CHCl_3 extracts or solutions should not be left standing for long before completing the assay, nor should they be exposed to strong light.

Ethanollic extracts are also quite stable in the dark but exposure to light leads to fairly rapid loss of cannabinoids (Figs 1 and 2). These results agree with those for other solvents but do not support the claim made by Kubena & others (1972) that an ethanolic extract (Fluid extract of Cannabis U.S.P.X) had lost little or any of its original activity after 43 years storage in amber bottles with temperatures ranging up to 38°. Obviously no figures for the original THC concentration are available but reference to the U.S.P. X shows that the fluid extract is prepared from the "dried flowering tops of pistillate plants" and that 1 ml should represent 1 g of the drug. In view of the fact that flowering tops produced in England contained 2 to 7% THC (Fairbairn & Liebmann, 1974), it would be reasonable to assume that the USP cannabis (probably from India) would contain at least 4% THC. The authors reported that the

Table 4. *Cannabis resin (freshly prepared): changes in cannabinoid content (%) in different conditions of storage for 1 year at room temperature (20°).*

Conditions	THC	CBN
Freshly prepared (1a)	11.6	traces
Light and air		
1. Loose powder	7.3	1.8
2. Compressed powder but mats broken up	5.2	2.4
3. Compressed, unbroken lump		
(a) Surface layer (0.2 mm)	5.2	2.3
(b) Next 2 mm layer	10.7	1.8
(c) Centre	11.4	1.6
Darkness and air		
4. Loose powder	12.0	0.65
5. Compressed but broken up	10.1	1.0
6. Compressed, unbroken lump		
(a) Surface layer	11.1	1.86
(b) Next 2 mm layer	10.3	1.04
(c) Centre	11.0	1.47

fluid extract only contained 0.4% THC, indicating that only about 10% of the original amount remained, a value which would be consistent with our own experience.

The effect of light on one sample of *herbal cannabis* (Table 2) shows once more its deleterious effect. The results in Table 3 on a further 8 samples confirm that in the absence of light carefully dried *herbal cannabis* is quite stable; in fact more so than previously thought. Thus Lerner (1969) surmised a rate of loss of THC of 3 to 5% per month at room temperature, corresponding approximately to 31 to 46% per year. Assuming a linear rate of decomposition Turner & others (1973a) found a loss of THC of 7% per year at room temperature when stored in amber bottles either in the dark or with limited exposure to light. Our seven samples of coarse powder in Tables 2 and 3 give corresponding values ranging from 5 to 26% with a mean of 13%. The results on *cannabis resin* (Table 4) also show the deleterious effects of light but in addition they illustrate other effects which are discussed below.

Although in our experiments the effect of temperature (5° and 20°) was not marked (Tables 1, 2) Turner & others (1973a) showed that at 37 and 50° significant losses occurred. As their experiments were carried out in ovens, presumably light played no part in the deterioration. Coffman & Gentner (1974) also studied the effect of higher temperatures for periods up to 64 h. Little decomposition took place at 65° but considerable losses occurred at 85 and 100°.

The effect of oxygen on *herbal cannabis* seems much less significant than that of light or higher temperatures. This may be due to the fact that, in the carefully dried plant, the cannabinoids are stored in "well-closed containers", the glands. If these are burst by careless preparation or in the making of resin, then loss by oxidation is more likely. The results for *herbal cannabis* in Table 3 are consistent with this suggestion: losses in the fine powder, where more glands will be damaged, are consistently greater than in the coarse powder. On average the 6 samples of fine powder contained 11% less cannabinoids than the coarse powder after 1 year storage. The results of a more ambitious experiment on specially prepared resin are given in Table 4. The loose powder showed less loss in THC after 1 year than compressed powder in which many glands would be burst. However, if the compressed powder was

retained as one lump this loss was mainly confined to the surface layer which clearly protected the inner layers from breakdown. This is also shown in our work on the Pakistani resin sample (see Results) in which the inner layers always contained more THC than the outer. The results (Table 4), do show that the effect of oxygen is much less significant than that of light, in the material we used. Thus in the light there was a loss, for the loose powder, of 37% of the original activity and for the compressed material (more glands broken) of 55% (samples 1 and 2). In the dark the losses in comparable samples 4 and 5, are only zero and 13% respectively. Razdan & others (1972), however, deliberately increased the possible effect of oxygen by spreading THC on filter paper (in the dark at 25°); in these circumstances they report a loss of 75% in 10 months. It can be concluded, that cannabis resin is quite stable if stored in the dark in large lumps which prevent access of air to the interior.

We have already referred to the fact that decomposition of THC in the light does not lead to an increase of CBN, although the latter is not markedly unstable in the light (Table 1, sample 5, and Fig. 1). Conversion to CBN however seems to occur in the resin stored in the dark as well as in the light (Table 4). Levine (1944) also reported a significant conversion of THC to CBN in resin stored in the dark for 3 years. Our results on the Pakistani resin also showed that a low THC content corresponded to a high CBN and vice versa. We make the tentative suggestion therefore that the effect of light may be to convert THC rapidly to polymers which would not be detected by the usual g.l.c. and t.l.c. methods and of oxygen to convert the THC to CBN, some of which will polymerize especially in the light. It would be intriguing to know however how the cannabinoids in the living plant are preserved in the transparent glands which are often exposed for long periods to brilliant sunshine.

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REFERENCES

- COFFMAN, C. B. & GENTNER, W. A. (1974). *Bull. Narcotics*, 26 (1), 67-70.
ECKLER, C. R. & MILLER, F. A. (1917). *J. Am. pharm. Ass. Sci. Ed.*, 6, 872.
FAIRBAIRN, J. W. & LIEBMAN, J. A. (1973). *J. Pharm. Pharmac.*, 25, 150-155.
FAIRBAIRN, J. W. & LIEBMAN, J. A. (1974). *Ibid.*, 26, 413-419.
GARRETT, E. R. & TSAU, J. (1974). *J. pharm. Sci.*, 63, 1563-1574.
KUBENA, R. K., BARRY, H., SEGELMAN, A. B., TREINER, M. & FARNSWORTH, N. R. (1972). *Ibid.*, 61, 144-145.
LERNER, P. (1969). *Bull. Narcotics*, 21, (3), 39-42.
LEVINE, J. (1944). *J. Am. chem. Soc.*, 66, 1868-1870.
MAUNDER, M. J. DE FAUBERT (1970). *J. Assoc. Publ. Analysts*, 8, 42-47.
MECHOULAM, R. (1970). *Science*, 168, 1159-1166.
PARKER, J. M., BORKE, M. L., BLOCK, L. H. & COCHRAN, T. G. (1974). *J. pharm. Sci.*, 63, 970-971.
RAZDAN, R. K., PUTTICK, A. J., ZIKTO, B. A. & HANDRICK, G. R. (1972). *Experientia*, 28, 121-122.
SCHOU, J. & NIELSEN, E. (1970). *Dansk. Tidskr. Farm.*, 44, 359-364.
TURNER, C. E., HADLEY, K. W., FETTERMAN, P. S., DOORENBOS, N. J., QUIMBY, M. W. & WALLER, C. (1973a). *J. pharm. Sci.*, 62, 1601-1605.
TURNER, C. E., HADLEY, K. W. & DAVIS, K. H. (1973b). *Acta Pharm. Jugoslav.*, 23, 89-94.
TURNER, C. E. & HENRY, J. T. (1975). *J. pharm. Sci.*, 64, 357-359.

CANNABIS. XV. ¹ PREPARATION AND STABILITY OF
 Δ^9 -TETRAHYDROCANNABINOL- β -CYCLODEXTRIN
INCLUSION COMPLEX

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ABSTRACT.—Investigation is made of the preparation of the inclusion complex of Δ^9 -tetrahydrocannabinol (1) with β -cyclodextrin (β -CD) in solid form by the precipitation method. In order to estimate the inclusion complex formation, various spectral data (uv, cd, and emission spectra) have been studied. It has been estimated that the inclusion complex is highly stable under various conditions (light of 10,000 lux, oxygen existence, and high temperature (120°)) compared to 1 alone. The extensive application for a drug delivery system is also discussed.

Current interest in the inclusion complex of cyclodextrins (CD) with various kinds of drugs has been centered on their chemical stability, solubility, and absorption properties. The major active principle of marijuana, Δ^9 -tetrahydrocannabinol, is a fragile compound with respect to light (1), auto-oxidation (2), high temperature (1,3), acidic medium (4), and free radical oxidation (5). The major pathway for auto-oxidation of 1 is summarized in figure 1. Compound 1 requires the presence of detergent in order to dissolve in water, thus creating problems in pharmacological studies.

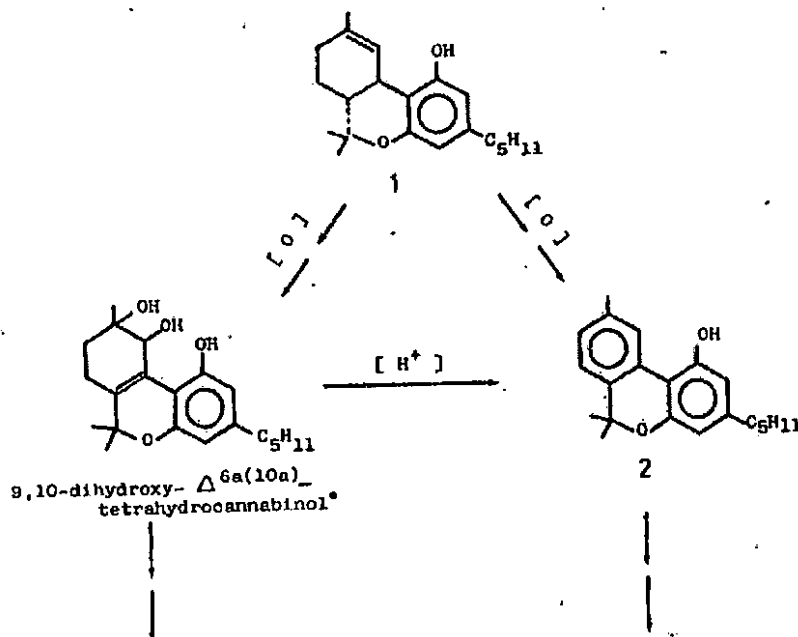


FIGURE 1. Pathway of autoconversion of 1.

*M.A. Elsohly, F.S. El-Ferly and C.E. Turner, *J. Nat. Prod.*, 40, 275 (1977).

¹Previous article: Cannabis XIV. Y. Shoyama, S. Morimoto, and I. Nishioka, *Chem. Pharm. Bull.*, 29, 3720 (1981).

This paper presents the preparation of a Δ^9 -tetrahydrocannabinol- β -cyclodextrin complex (1- β -CD), a more stable compound when compared with 1 alone.

RESULTS AND DISCUSSION

In order to estimate the ability of inclusion complex formation, various spectra data have been studied. Figure 2 shows the uv spectral data of 1 alone and of 1- β -CD in 0.1N NaOH. No variation between the 1 and 1- β -CD spectral profile could be detected; however, the different spectrum in the similar spectra clearly showed the difference of intensity as indicated in Figure 3. The cd spectra of 1 alone and of 1- β -CD are shown in Figure 4 to have similar maximum wave lengths; they differ from each other in intensity. Figure 5 illustrates the emission spectra of the 1- β -CD system in 0.1N NaOH. These spectra indicate that 1 with β -CD increases the intensity and shifts the absorption wave length. These spectra support the existence of the formation of the inclusion complex between 1 and β -CD. The methods for preparation of the inclusion complex have previously been elucidated (6,7); in this study we used the precipitation method.

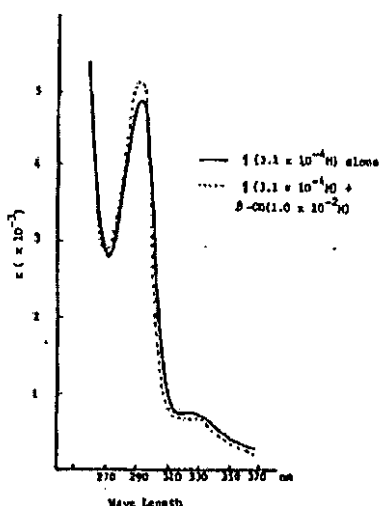


FIGURE 2. The uv spectra of 1 and 1- β -CD inclusion complex in 1N NaOH.

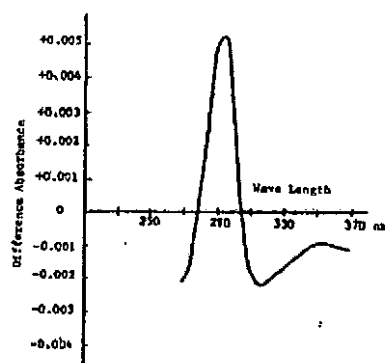


FIGURE 3. Difference spectrum between 1 and 1- β -CD inclusion complex.

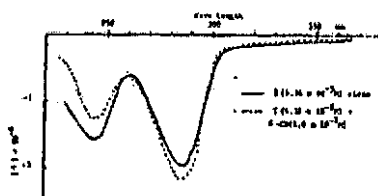


FIGURE 4. The cd spectra of 1- β -CD system in 0.1N NaOH.

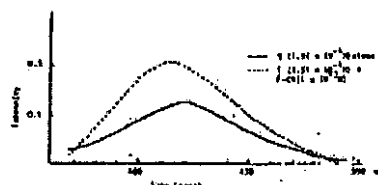
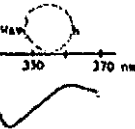


FIGURE 5. Emission spectra of 1- β -CD system in 0.1N NaOH.

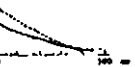
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The inclusion complex of 1- β -CD is a white powder, $C_{103}H_{170}O_{72}$, showing aromatic double bond absorbance (1622 and 1577 cm^{-1}) in the ir spectrum. ^1H -nmr. spectrum shows aromatic protons at δ 5.57 and 5.90. However, it is difficult to confirm the effects of β -CD on each signal of 1. The stoichiometry of the complex in the solid state was analyzed chemically, as indicated, and found to be 2:1 for the β -CD system.

Next, the stabilities of 1 and 1- β -CD were determined. It has been demonstrated that the major artificial product of 1 is cannabinol (2) (8). The major route of 1 oxidation occurs by auto-oxidation under storage conditions. The influences of temperature, light, and the presence of oxygen have been determined (table 1). Degradation of the 1- β -CD complex occurred only in experiment 4, which showed the appearance of 2 after storage for five months. Experiments 1 and 2 indicated the lack of effect by light, if the nitrogen was replaced. The presence of oxygen caused oxidative degradation of 1, producing 2 and other products, as indicated in experiments 3 and 4. The amount of 2 produced varied, as indicated by these experiments. Moreover, the presence of light along with oxygen increased degradation to 2. In addition to the experiments performed at 30° , the influence of oxygen also was observed at -10° (experiment 6). The amount of 1 present was 50.8% and that of 2 was 20.9%, thus indicating that the essential factor for storage of 1 was a decrease in the oxygen concentration. On the other hand, experiments performed at -10° and 30° (experiments 1 and 5) in the presence of nitrogen indicated little variation in the stability of 1.

TABLE 1. Stabilities of 1- β -CD^a and 1^b alone under various conditions.

Experiment Number	Condition ^d	1- β -CD ^a		1 ^b alone	
		1 ^b (%) 2 ^c		1 ^b (%) 2 ^c	
1	$30^\circ\text{N}_2\text{D}$	100.0	0	76.6	11.4
2	$30^\circ\text{N}_2\text{L}$	100.0	0	74.5	10.0
3	$30^\circ\text{O}_2\text{D}$	98.2	0	2.7	30.0
4	$30^\circ\text{O}_2\text{L}$	86.1	2.0	3.3	17.4
5	$-10^\circ\text{N}_2\text{D}$	100.0	0	74.2	11.6
6	$-10^\circ\text{O}_2\text{D}$	98.2	0	50.8	20.9

^a Δ^9 -tetrahydrocannabinol- β -cyclodextrin inclusion complex.

^b Δ^9 -tetrahydrocannabinol.

^cCannabinol.

^d N_2 : replacement of air by N_2 , O_2 : air existence.

D=under dark, L=under light, storage period=5 months.

In order to confirm the stability of the 1- β -CD under more stringent storage conditions, the effects of higher temperatures and darkness were studied to determine differences between the 1- β -CD complex and the 1- β -CD mechanical mixture. The amounts of 1 alone and of the 1- β -CD mechanical mixture gradually decreased over time when heated to 120° in the dark. The explanation for this phenomenon may depend on the exposed surface areas of the different structures. That is, the decomposition of 1 may be increased by the porous surface of β -CD rather than 1 alone. The degradative product from 1 alone and from the 1- β -CD mechanical mixture was 2 in both cases, as identified by glc; the amount of 2 present is listed in figure 6.

On the other hand, the 1- β -CD inclusion complex does not degrade at 120° in the dark over a 24-h period. This finding suggests that the labile part, such as double bond and/or phenol is included by β -CD. Although the direct conformation of the 1- β -CD inclusion complex in a solid state is not yet available, the results thus far support the existence of this structure.

The inclusion complex of 1 with β -CD may have wide applications. For example,

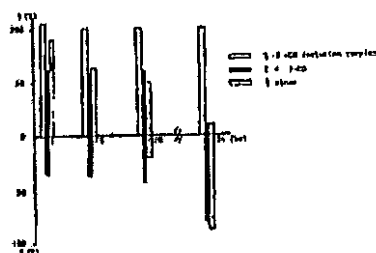


FIGURE 6. Stabilities of 1- β -CD inclusion complex, 1- β -CD mechanical mixture and 1 alone by heating at 120°.

the long-term storage of 1 with increased solubility in water will be possible. Also, CD complexation may be extensively applied to enhance the membrane permeability and bioavailability of slightly soluble 1. Further studies on other cannabinoids are in progress. These results will be presented elsewhere in the near future.

EXPERIMENTAL

The uv spectra were obtained on a Hitachi Model 100-10 Spectrophotometer and the ir spectra were determined on a Nihonbunko Model DS-100 Spectrometer in KBr pellets. The ^1H -nmr spectra were recorded in 1 N NaOH on a JEOL PS-100 Spectrometer; chemical shifts are given on the ppm scale, and signal multiplicities are represented by s(singlet), d(doublet), t(triplet), q(quarter), m(multiplet), and br(broad). The cd spectra were measured on a JASCO J-20 Spectropolarimeter. The emission spectra were measured on a Hitachi MPF-2A Fluorescence Spectrophotometer. The glc analyses were determined on a Shimadzu Gas Chromatograph model GC-4BM under previously reported conditions (9).

Compound 1 and 2 were isolated from the Mexican cannabis as described previously (10).

PREPARATION OF 1- β -CD INCLUSION COMPLEX.—Compound 1 (0.4 g) dissolved in MeOH (8 ml) was added to 4.3% β -CD solution (400 ml) dropwise, while being stirred. After 1 h of stirring, the reaction mixtures were centrifuged (3000 rpm, 25 min). The precipitates were then washed twice with a small amount of water, dried on $\text{c.H}_2\text{SO}_4$, and finally washed twice more with a small amount of ether to remove free 1. Finally, the white powder obtained was completely dried over P_2O_5 at 80° for 5 h. The inclusion complex was analyzed by uv and glc methods, as described below, to determine the ratio of 1- β -CD (1:2). The 1- β -CD complex was a white powder: uv λ max (H_2O) 276 nm; cd (6.72×10^{-3} M 0.1 N NaOH) $[\theta]_{247}^{25} = -1.43 \times 10^4$, $[\theta]_{291}^{25} = -2.35 \times 10^4$; ir (KBr) ν max: 1622, 1577 cm^{-1} ; ^1H -nmr (1 N NaOH) δ 0.99(t, w-Me), 1.33(s, $\text{C}_6\text{-Me}$), 1.36(s, $\text{C}_6\text{-Me}$), 1.90-2.30(m, $\beta\text{-CH}_2$), 3.35(t, glu₄-H), 3.46(q, glu₂-H), 3.74(m, glu_{3,5} and e-H), 4.86(d, glu₁-H), 5.57(bs, arom H), 5.90(bs, arom H), 6.23(bs, C_{10} -H). Anal. Calcd for $\text{C}_{103}\text{H}_{170}\text{O}_{72}$: C, 48.80; H, 6.58. Found: C, 48.64; H, 7.09.

ANALYTICAL SYSTEM.—The uv method: 1- β -CD inclusion complex was dissolved in MeOH- H_2O (1:1) and analyzed by uv photometer at 270 nm to determine the content of 1 using the calibration curves. The glc method: Samples of inclusion complex (20 mg) were suspended in diluted MeOH (1 ml) under sonication. To the solution a drop of cyclohexylamine was added, then extracted with ether (5 ml) five times. After evaporation of ether, the residue was analyzed by glc as described previously (9).

STORAGE OF SAMPLES.—Samples (1 alone, 10.68 mg; 1- β -CD inclusion complex, 20 mg) were sealed in glass ampules with air or with replacement of air by N_2 and were stored either under light of 10,000 lux or in the dark for five months. The storage at 30° was undertaken at the Biotron Institute of this University.

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LITERATURE CITED

1. Y. Shoyama, T. Yamauchi, and I. Nishioke, *Chem. Pharm. Bull.*, **18**, 1327 (1970).
2. R.K. Razdan, A.J. Puttick, B.A. Zicko, and G.R. Handrick, *Experientia*, **28**, 121 (1972).
3. G.S. Lewis and C.E. Turner, *J. Pharm. Sci.*, **67**, 876 (1978).
4. Y. Gaoni and R. Mechoulam, *Tetrahedron*, **22**, 1481 (1966).

5. I.J. Miller, N.K. McCallum, C.M. Kirk, and B.M. Peske, *Experientia*, **38**, 230 (1982).
6. F. Cramer and G.M. Henglein, *Chem. Ber.*, **90**, 2561 (1957).
7. M. Kurozumi, N. Namba, and T. Nagai, *Chem. Pharm. Bull.*, **23**, 3063 (1975).
8. R. Mechoulam, in: "Marijuana, Chemistry, Pharmacology, Metabolism and Clinical Effects," Ed. by R. Mechoulam, Academic Press, New York, 1973, pp 60-75.
9. H. Kushima, Y. Shoyama, and I. Nishioka, *Chem. Pharm. Bull.*, **28**, 594 (1980).
10. Y. Shoyama, S. Morimoto, and I. Nishioka, *Chem. Pharm. Bull.*, **29**, 3720 (1981).

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