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(54) Title: MODIFIED PHOSPHATASES

(57) Abstract: The invention relates to phosphatases and more in specific to (genetically) modified phosphatases, pharmaceutical compositions comprising (genetically) modified phosphatases and the use of (genetically) modified phosphatases for treating or curing for example sepsis, inflammatory bowel disease or other inflammatory diseases, or renal failure. The invention further relates to a method for producing phosphatases.



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Title: Modified phosphatases

The invention relates to phosphatases and more in specific to (genetically) modified phosphatases, pharmaceutical compositions comprising (genetically) modified phosphatases and the use of (genetically) modified phosphatases for treating or curing for example sepsis, inflammatory bowel disease or other inflammatory disease, or renal failure. The invention further relates to a method for producing phosphatases.

10 A phosphatase is an enzyme that dephosphorylates its substrate; i.e. it hydrolyses phosphoric acid monoesters into a phosphate ion and a molecule with a free hydroxyl group. This action is directly opposite to that of phosphorylases and kinases, which attach phosphate groups to their substrates by using energetic molecules like ATP. Phosphatases can be
15 categorised into two main categories: Cysteine-dependent Phosphatases (CDPs) and metallo-phosphatases. The latter ones are dependent on the presence of one or more metal ions in their active site(s) for activity.

CDPs catalyse the hydrolysis of a phosphoester bond via a phospho-cysteine intermediate. The free cysteine nucleophile forms a bond with the
20 phosphorus atom of the phosphate moiety, and the P-O bond linking the phosphate group to the tyrosine is protonated, either by a suitably positioned acidic amino acid residue or a water molecule. The phospho-cysteine intermediate is then hydrolysed by another water molecule, thus regenerating the active site for another dephosphorylation reaction.

25 Metallo-phosphatases co-ordinate 1 or more catalytically essential metal ion(s) within their active site. There is currently some confusion of the identity of these metal ions, as successive attempts to identify them yield different answers. There is currently evidence that these metals could be Magnesium, Manganese, Iron, Zinc, or any combination thereof. It is thought that a
30 hydroxyl ion bridging the two metal ions takes part in nucleophilic attack on the phosphate group

Phosphatases act in opposition to kinases/phosphorylases, which add phosphate groups to proteins. The addition of a phosphate group may activate or de-activate an enzyme (e.g., Kinase signalling pathways) or enable a protein-protein interaction to occur (e.g., SH3 domains); therefore

5 phosphatases are integral to many signal transduction pathways. It should be noted that phosphate addition and removal do not necessarily correspond to enzyme activation or inhibition, and that several enzymes have separate phosphorylation sites for activating or inhibiting functional regulation. CDK, for example, can be either activated or deactivated depending on the specific
10 amino acid residue being phosphorylated. Phosphates are important in signal transduction because they regulate the proteins to which they are attached. To reverse the regulatory effect, the phosphate is removed. This occurs on its own by hydrolysis, or is mediated by protein phosphatases.

15 Without limiting the present invention, alkaline phosphatases are discussed in more detail as an example of the herein described and claimed phosphatases. Alkaline phosphatase (ALP) (EC 3.1.3.1) is a hydrolase enzyme responsible for removing phosphate groups from many types of molecules, including nucleotides, proteins, and alkaloids. The process of removing the
20 phosphate group is called dephosphorylation. As the name suggests, alkaline phosphatases are most effective in an alkaline environment.

Alkaline phosphatase has become a useful tool in molecular biology laboratories, since DNA normally possesses phosphate groups on the 5' end. Removing these phosphates prevents the DNA from ligation (the 5' end
25 attaching to the 3' end of the same or another molecule); also, removal of the phosphate groups allows radiolabeling (replacement by radioactive phosphate groups) in order to measure the presence of the labeled DNA through further steps in the process or experiment. For these purposes, the alkaline phosphatase from shrimp is the most useful, as it is the easiest to inactivate
30 once it has done its job.

Another important use of alkaline phosphatase is as a label for enzyme immunoassays.

Moreover, alkaline phosphatases are used in the treatment of for example sepsis, inflammatory bowel disease, or renal failure.

5 Although the presently available (alkaline) phosphatases are useful in both diagnostics and disease treatment there is a need for alternative phosphatases with for example an altered (for example improved) specific activity, stability (for example in vivo $T_{1/2}$, or stability in respect of storage (shelf-life)) or substrate specificity. Moreover, there is also a need for
10 phosphatases with a different pH or temperature or salt (in)dependency profile.

The present invention provides alternative (genetically) modified phosphatases.

15 In a first embodiment, the invention provides an isolated or recombinant alkaline phosphatase comprising a crown domain and a catalytic domain, wherein said crown domain and said catalytic domain are obtained from different alkaline phosphatases. These mutants are herein further referred to as "domain swapped mutants".

20 Alkaline phosphatase (AP); EC 3.1.3.1 according to IUBMB Enzyme Nomenclature, the common name is alkaline phosphatase (AP), is an enzyme that catalyzes the reaction of a phosphatase monoester and H_2O to an alcohol and phosphate. Other name(s) for AP are alkaline phosphomonoesterase; phosphomonoesterase; glycerophosphatase; alkaline phosphohydrolase;
25 alkaline phenyl phosphatase; orthophosphoric-monoester phosphohydrolase (alkaline optimum). The systemic name of AP is phosphate-monoester phosphohydrolase (alkaline optimum).

AP is a wide specificity enzyme, it also catalyses transphosphorylations. In humans and other mammals at least four distinct, but related alkaline
30 phosphatases are known. In humans these are intestinal, placental, placental-

like, and liver/bone/kidney (or tissue non-specific) alkaline phosphatase. The first three are located together on chromosome 2 while the tissue non-specific form is located on chromosome 1. The exact physiological functions of the APs are not known, but AP appears to be involved in a large number of physiological processes.

The placental alkaline phosphatase is herein abbreviated as ALPP or PLAP. The abbreviations ALPI or IAP refer to intestinal alkaline phosphatase. The placental-like 2 alkaline phosphatase is herein abbreviated as ALPP2, ALPG or GCAP and the abbreviations ALPL, TNSALP, TNAP or BLK are herein used to refer to liver/tissue non-specific alkaline phosphatase. The different abbreviations for one and the same alkaline phosphatase are used interchangeably herein.

From a conformational point of view, an alkaline phosphatase roughly consists of two domains: a crown domain and an active-site domain. The active-site domain can be divided in separate parts like the catalytic residue and the three metal ion sites (Zn1, Zn2 and Mg3). From a primary structure point of view it is clear that the crown domain is flanked by the amino acids that form the active site domain. Hence, in a preferred embodiment, the catalytic domain is not composed of a contiguous sequence of amino acids, but is flanking the crown domain.

The amino acid sequence of alkaline phosphatases and the relative positions of the catalytic and crown domain are known by the skilled person. As an example, reference is made to Figure 1 which shows, amongst others, the amino acid sequence of the four human alkaline phosphatases. The crown domain is underlined in these sequences. The domain swapped mutants of the invention preferably have been made by replacing their own crown domain (as underlined) by a crown domain of another phosphatase (as underlined). For example, the crown domain of ALPP is located between amino acids 366 to 430 and hence in a preferred embodiment reference to the crown domain corresponds to the amino acids 366 to 430 in Figure 1, i.e. in a preferred

embodiment the invention provides an isolated or recombinant alkaline phosphatase comprising a crown domain and a catalytic domain, wherein said crown domain and said catalytic domain are obtained from different alkaline phosphatases and which crown domain in the ALPP of Figure 1 is located
5 between amino acid 366 to 430.

Alkaline phosphatases are present in virtually all organisms from bacteria to humans. In a preferred embodiment, the invention provides an isolated or recombinant alkaline phosphatase comprising a crown domain and a catalytic domain, wherein said crown domain and said catalytic domain are
10 obtained from different alkaline phosphatases and wherein at least one of said different phosphatases is a human phosphatase. The other phosphatase is for example ECAP (*Escherichia coli* alkaline phosphatase) or one of the seven known BIAPs (Bovine Intestinal Alkaline Phosphatase). In a preferred embodiment, the invention provides an isolated or recombinant alkaline
15 phosphatase comprising a crown domain and a catalytic domain, wherein said crown domain and said catalytic domain are obtained from different alkaline phosphatases and wherein the different alkaline phosphatases are human phosphatases. This is especially useful if the modified phosphatase is subsequently used in human therapy, for example in the treatment of sepsis,
20 inflammatory bowel disease or other inflammatory disease, or renal failure. It is expected that such (genetically) modified phosphatases of human origin are not or very little immunogenic. However it is clear to the skilled person that if a modified phosphatase is for example used in "in vitro" or "ex vivo" diagnostics a modified phosphatase may well be composed of for example a
25 human and an *E.coli* alkaline phosphatase or may be composed of a bovine and an *E.coli* alkaline phosphatase.

In yet another preferred embodiment, the invention provides an isolated or recombinant alkaline phosphatase comprising a crown domain and a catalytic domain, wherein said crown domain and said catalytic domain are
30 obtained from different alkaline phosphatases and wherein said crown domain

is the crown domain of ALPP and wherein said catalytic domain is the catalytic domain of ALPI. Preferably, at least one of said different phosphatases is a human phosphatase and in an even more preferred embodiment, both different phosphatases are human phosphatases.

5 Until the present invention it was generally believed that the catalytic domain of an alkaline phosphatase was the most important domain in respect of specific activity. Moreover, it was believed that the crown domain was involved in the stability of an alkaline phosphatase. Thus, upon testing a recombinant alkaline phosphatase comprising the catalytic domain of ALPI
10 and the crown domain of ALPP (further referred to as catALPI/crownALPP) it was expected that the activity of this recombinant alkaline phosphatase would be comparable to the activity of ALPI. However, production of catALPI/crownALPP in a medium with no or very little Zn^{2+} , for example Freestyle™ 293 expression medium (GIBCO), resulted in a specific activity of
15 approximately 600 U/mg whereas ALPI produced by the same cell line and in the same medium resulted in a specific activity of about 30 U/mg.

Even more surprisingly was the effect of adding Zn^{2+} ions to the growth medium of the producer cells: this has little effect on the specific activity of catALPI/crownALPP whereas the specific activity of ALPI increased
20 to approximately 750 U/mg. Addition of similar concentrations of Zn^{2+} after production induced only a 2-fold increase in specific activity of ALPI after 16 h.

A summary of these results is provided in Tables 1 and 2.

Without being bound by theory, it is thought that the crown domain of ALPP provides a conformational change in the produced recombinant
25 catALPI/crownALPP which results in a higher specific activity and a Zn^{2+} independent enzyme, i.e. the presence of the crown domain of ALPP results in relative high specific activity and is considered to be Zn^{2+} independent.

Furthermore, it was not only shown that ALPI required high Zn^{2+} concentrations during production to increase the specific activity of the
30 enzyme, but also that specific activity of ALPI decreased within 24 hours in

Zn^{2+} deficient medium, whereas catALPI/crownALPP retained its initial specific activity under the same conditions. These results imply that the in vivo activity is Zn^{2+} independent. Such an enzyme whose activity is independent of Zn^{2+} could be useful in illnesses where Zn^{2+} depletion is part of the pathology (e.g. nutritional defects, alcohol abuse and intestinal integrity damage, chronic infections including sepsis, or inflammatory diseases in general) or where addition of Zn^{2+} may be contraindicated (e.g. acute phase of sepsis, autoimmune diseases). Apart from production and application advantages, catALPI/crownALPP also has advantages in respect to stability during storage.

It has thus been shown that native AP, such as ALPI, loses its enzymatic activity in environments with low Zn^{2+} concentrations. Thus in diseases, wherein Zn^{2+} depletion is part of the pathology, said native AP is unable to unfold its enzymatic activity at the site where it is thought to be the most beneficial, e.g. at the site of inflammation. In contrast, a recombinant AP not susceptible to low Zn^{2+} concentrations, e.g. catALPI/crownALPP retains its activity in an environment with low Zn^{2+} concentration, e.g. at an inflammation site. In a healthy individual Zn^{2+} serum reference values are between 10 and 20 μM . For instance in alcohol abuse or during malnutrition, these levels can decrease to less than 10 μM or even less than 1 μM . Several enzymes in the human body depend on Zn^{2+} for their activity and for instance immunologic responses are more effective if sufficient levels of Zn^{2+} are present. The innate as well as the specific parts of the immune system are known to be influenced by zinc and it has been established that zinc containing proteins accumulate at sites of inflammation. Furthermore, (sub)chronic inflammation, such as rheumatoid arthritis, sepsis, and Crohn's disease present with serum zinc deficiency. Surprisingly, the invention also provides the insight that catALPI/crownALPP retains its activity in much broader pH range than unmodified (recombinant) alkaline phosphatase. Given the fact that many disorders, such as inflammation and/or ischemia, encompass

disturbances in tissue pH, catALPI/crownALPP is thus particularly useful for the treatment of such diseases. In one embodiment therefore, the invention provides a use of a phosphatase comprising a catalytic domain of ALPI and a crown domain of ALPP as a medicament, preferably for use in treating a disease which is accompanied by disturbed tissue pH, preferably said disease comprises an inflammatory disease and/or a disease accompanied with ischemia.

The invention provides the insight that a recombinant phosphatase comprising the catalytic domain of ALPI and the crown domain of ALPP (catALPI/crownALPP) is especially useful in the treatment of a disease that is accompanied with local or systemic Zn^{2+} deficiency. In another embodiment therefore, the invention provides a use of a phosphatase comprising a catalytic domain of ALPI and a crown domain of ALPP as a medicament, preferably for use in treating a disease which is accompanied by Zn^{2+} deficiency. Preferably said disease comprises an inflammatory disease, more preferably selected from the group consisting of autoimmune diseases, rheumatoid arthritis, asthma, chronic obstructive pulmonary disease, atherosclerosis, inflammatory bowel disease, sepsis, neurodermitis and diseases depicted in Table 10.

In another embodiment, the invention provides the use of a phosphatase comprising a catalytic domain of ALPI and a crown domain of ALPP in the preparation of a medicament for the treatment of a disease which is accompanied by Zn^{2+} deficiency, preferably said disease comprises an inflammatory disease, more preferably a disease selected from the group consisting of autoimmune diseases, rheumatoid arthritis, asthma, chronic obstructive pulmonary disease, atherosclerosis, inflammatory bowel disease, sepsis, neurodermitis and diseases depicted in Table 10.

In yet another embodiment, the invention provides a method for treating a subject (preferably a human) to treat a disease which is accompanied by Zn^{2+} deficiency, comprising administering an effective amount of a phosphatase comprising a catalytic domain of ALPI and a crown domain of

ALPP to a subject in need thereof, wherein said disease preferably comprises an inflammatory disease, more preferably selected from the group consisting of autoimmune diseases, rheumatoid arthritis, asthma, chronic obstructive pulmonary disease, atherosclerosis, inflammatory bowel disease, sepsis, neurodermitis and diseases depicted in Table 10.

In another preferred embodiment, the invention provides an isolated or recombinant alkaline phosphatase comprising a crown domain and a catalytic domain, wherein said crown domain and said catalytic domain are obtained from different alkaline phosphatases and wherein said crown domain is the crown domain of ALPI and wherein said catalytic domain is the catalytic domain of ALPP (further referred to as catALPP/crownALPI). Preferably, at least one of said different phosphatases is a human phosphatase and in an even more preferred embodiment, both different phosphatases are human phosphatases.

Other preferred domain swapped mutants that are based on the human alkaline phosphatases are:

Catalytic domain	Crown domain	Referred to as
ALPI	GCAP	catALPI/crownGCAP
	TNAP	catALPI/crownTNAP
ALPP	GCAP	catALPP/crownGCAP
	TNAP	catALPP/crownTNAP
GCAP	ALPI	catGCAP/crownALPI
	ALPP	catGCAP/crownALPP
	TNAP	catGCAP/crownTNAP
TNAP	ALPI	catTNAP/crownALPI
	ALPP	catTNAP/crownALPP
	GCAP	catTNAP/crownGCAP

For the sake of clarity, ALPI is intestinal AP, ALPP is placental AP, GCAP is placental-like AP and TNAP is tissue non-specific AP.

It is clear that also combinations between the catalytic domain of ECAP or any of the human forms (ALPI, ALPP, GCAP or TNAP) with the crown domain of BIAP can be made. Moreover, combinations of the crown domain of BIAP with the catalytic domain of ECAP or any of the human forms can also be produced.

Throughout the specification, examples and literature in the art, other nomenclature is used to designate the respective isoforms of alkaline phosphatase. For the sake of clarity, in the table below the names and abbreviations commonly used, or used in this application is listed.

ALKALINE PHOSPHATASES	ABBREVIATIONS
Placental alkaline phosphatase	ALPP, PLAP,
Secretable Placental alkaline phosphatase	shPLAP, sALPP
Intestinal alkaline phosphatase	ALPI, IAP hIAP
Secretable Intestinal alkaline phosphatase	shIAP, sALPI
Placental-like alkaline phosphatase	GCAP
Tissue non specific alkaline phosphatase	TNAP, BLK, ALPL, TNSALP
<i>E.coli</i> alkaline phosphatase	ECAP
Bovine intestinal alkaline phosphatase	BIAP
Recombinant alkaline phosphatase comprising the catalytic domain of ALPI and the crown domain of ALPP	catALPI/crownALPP, RecAP, Xinlap
Recombinant alkaline phosphatase comprising the catalytic domain of ALPI and the crown domain of ALPP	catALPP/crownALPI, shPLAP-hIAP-CD

Another class of useful modified phosphatases are phosphatases which under natural conditions are linked to the membrane of a cell via a glycosylphosphatidylinositol (GPI) anchor but which are now modified such that they are no longer attached to the membrane of a cell. Examples of

5 phosphatases that are GPI-anchored are alkaline phosphatase and 5'-nucleotidase. All isoenzymes are functionally active in the cell membrane and GPI-anchor deficient forms are not naturally present at detectable levels. Although serum alkaline phosphate activity has been demonstrated it is generally accepted that the enzyme is still present in shed membrane fractions

10 or membrane vesicles. AP activity in milk is also present in fractions containing membrane vesicles. The GPI anchor is stored as a precursor molecule in the cell where it is attached to the attachment site through a transamidase. The backbone of the GPI-anchor is identical in mammals, but cell-type dependent modifications are known.

15 Alkaline phosphatases are predominantly found in association with plasma-membranes via their GPI anchor. For example, neutrophils present the enzyme against the background of their negatively charged cell membrane instead of releasing it into the inflammatory microenvironment. For this reason it is commonly accepted that for optimal in vivo activity of AP the

20 enzyme should be embedded in a cell membrane or a vesicular membrane. Furthermore it has been observed that poly-anionic substrates can further contribute to favourable anionic conditions in vivo for phosphatase activity of phosphatase enzymes and derivatives thereof normally having an optimum at an alkaline pH, in particular for phosphatase activity of alkaline phosphatase.

25 For pharmaceutical use of AP in human subjects it is for most applications a requirement to apply human forms of the enzyme for medicaments and treatment, as AP forms obtained from other species may be immunogenic in human subjects and treatment could elicit immunological reactions and pathological side effects. In some subjects even lethal side effects

30 i.e. anaphylactic shock (shown in our animal studies) may occur and the risks

of immunological side effects should therefore be minimized. As isolation of AP from humans is practically not feasible, human recombinant forms of the AP proteins can be routinely produced in different recombinant expression platforms. However, expression and purification of GPI containing and
5 membrane-anchored proteins is notoriously difficult; GPI proteins are difficult to separate from membranes and difficult to isolate and purify. However, the GPI anchor and membrane localisation have always been regarded as essential for the biological activity of AP.

This part of the current invention is based on the surprising finding that
10 human AP enzymes lacking a GPI-anchor and which enzymes are thus soluble and easily secreted by recombinant protein expression systems, display significant phosphatase activity at physiological pH levels towards biologically relevant phosphorylated substrates in a liver-cell based biologic assay.

In one of the embodiments, the invention provides an isolated or
15 recombinant phosphatase comprising a modification in the glycosylphosphatidylinositol (GPI) signal sequence, wherein said modification results in a secreted phosphatase, i.e. the phosphatase is not attached to the cell membrane.

In a preferred embodiment, the invention provides an isolated or
20 recombinant phosphatase comprising a modification in the glycosylphosphatidylinositol (GPI) signal sequence, wherein said modification results in a secreted phosphatase that is biological active, i.e. it show activity towards a biological (relevant) substrate.

There is no general sequence responsible for the attachment of a GPI
25 anchor, but there is a clear consensus:

- 1) hydrophobic stretch of amino acids at the C-terminus (at least 11 amino acids, but preferably more than 11 amino acids)
- 2) Upstream of the hydrophobic region, a spacer of hydrophylic amino acids (5-12 amino acids)

3) GPI is attached to a small amino acid: glycine, aspartic acid, asparagine, alanine, serine or cysteine.

4) The 2 subsequent amino acids downstream of the GPI attachment site must be small amino acids and in the majority of cases they are selected from glycine, aspartic acid, asparagine, alanine, serine or cysteine.

Based on this consensus, the skilled person is capable of mutating this consensus, for example by inserting one or multiple amino acids and disrupting part of the consensus. However in a preferred embodiment, the invention provides an isolated or recombinant phosphatase comprising a modification in the glycosylphosphatidylinositol (GPI) signal sequence, wherein said modification results in a secreted phosphatase and wherein said modification comprises a mutation or a deletion of the amino acid sequence encompassing the consensus GPI signal sequence.

For applications in human therapy it is desired that the resultant modified phosphatase is not or very little immunogenic, i.e. that the modified phosphatase is essentially of human origin. In a preferred embodiment, the invention provides an isolated or recombinant phosphatase comprising a modification in the glycosylphosphatidylinositol (GPI) signal sequence, wherein said modification results in a secreted phosphatase (preferably with activity against a biological relevant substrate) and wherein said phosphatase is a human phosphatase.

Examples of phosphatases that are GPI-anchored are alkaline phosphatase and 5'-nucleotidase and hence in a preferred embodiment, the invention provides an isolated or recombinant phosphatase comprising a modification in the glycosylphosphatidylinositol (GPI) signal sequence, wherein said modification results in a secreted phosphatase and wherein said phosphatase is an alkaline phosphatase, for example a human alkaline phosphatase, such as human liver-kidney-bone phosphatase, human intestinal alkaline phosphatase, or human placental-like alkaline phosphatase.

It is clear that any of the described secretable modified phosphatase can for example be produced by introducing into a host cell a nucleic acid capable of encoding said secretable phosphatase in operable linkage with regulatory sequences and allowing said host cell to express said secretable phosphatase and optionally isolating the produced phosphatase from the medium in which the host cell are grown and/or maintained. However, apart from mutations in the above mentioned GPI-attachment sequence, other methods exist that make GPI-anchorless, secreted proteins:

- 1) After expression as membrane anchored proteins, phospholipases may be used to cleave off the GPI anchor. Hence the invention also provides a method for producing a secreted phosphatase comprising culturing a host capable of expressing a membrane anchored phosphatase, allowing said host cell to produce said phosphatase and incubating the obtained cells with a phospholipase and optionally isolating the released phosphatase. The membrane anchored phosphatase is for example a wild type (or natural or non-modified) phosphatase. However, the membrane anchored phosphatase can comprise mutations in other parts of its sequence (for example the crown domain).
- 2) Interference with the production of the GPI anchor or the use of a cell (type) that is deficient in GPI anchor production may also be used to make a secretable form of an otherwise GPI-anchored protein. Examples of cell lines that have been made to be deficient in GPI anchoring biochemistry are e.g. Jurkat, AM-B, C84, BW, S49, CHO and Raji. In yet another embodiment the invention therefore provides a method for producing a secreted phosphatase comprising culturing a host cell capable of expressing a secretable (alkaline) phosphatase (for example a host cell comprising a nucleic acid sequence encoding any of the mentioned modified secreted (alkaline) phosphatases), allowing said host to produce said secretable

phosphatase and optionally isolating the produced phosphatase, wherein said host cell is not capable of biosynthesis of functional GPI anchored proteins. However, the host cell may also produce a phosphatase with a functional GPI signal sequence.

- 5 3) Interference with or the use of a cell deficient in transamidases may be used to inhibit attachment of a GPI anchor to the protein, rendering the protein anchorless and secretable. Such a deficient cell has been obtained through mutagenesis in CHO.

10 It is clear to the skilled person that a modified phosphatase which comprises a crown domain and a catalytic domain, wherein said crown domain and said catalytic domain are obtained from different alkaline phosphatases can be further modified and made secretable. Hence, in a preferred embodiment, the invention provides an isolated or recombinant phosphatase
15 comprising a modification in the glycosylphosphatidylinositol (GPI) signal sequence, wherein said modification results in a secreted phosphatase and wherein said recombinant phosphatase further comprises a crown domain and a catalytic domain that are obtained from different phosphatases. Examples of such (alkaline) phosphatase mutants are provided in Figure 1. Such a
20 combined or "double" mutant results for example in a modified phosphatase with a certain specific activity, stability or substrate specificity and at the same time production of such a product is greatly enhanced by the fact that it can be isolated from the medium surrounding the producer cells.

25 The catalytic domain of alkaline phosphatases is composed of several amino acid sequences that are not contiguous in the primary sequence of the enzyme. The catalytic domain contains the catalytic Serine residue (Ser92 in ALPP) that serves as an acceptor for the phosphate group that is cleaved off the substrate in the dephosphorylation reaction. The catalytic
30 domain of the enzyme further contains one or more metal ions. Specific amino

acid residues contained within the catalytic domain are responsible for the binding and coordination of the metal ions that are involved in the dephosphorylation reaction. In ALPP the metal coordinating residues are: Asp42, His153, Ser155, Glu311, Asp316, His320, Asp357, His358, His360 and
5 His432.

In yet another embodiment, the invention provides an isolated or recombinant phosphatase comprising a mutation in the vicinity of a catalytic residue and/or in a metal ion-coordinating phosphate-binding pocket. The skilled person is very well capable of identifying and mutating amino acids
10 around (i.e. in the, preferably conformational, vicinity of) a catalytic residue and/or in a metal ion-coordinating phosphate pocket. As already has been described above, sequences of phosphatases are known. As an example, Figure 1 shows, amongst others the amino acid sequence of four human alkaline phosphatases.

15 In a preferred embodiment, the invention provides an isolated or recombinant phosphatase comprising a mutation in the vicinity of a catalytic residue and/or in a metal ion-coordinating phosphate-binding pocket wherein said phosphatase is a human phosphatase. This is especially useful if the modified phosphatase is subsequently used in human therapy, for example in
20 the treatment of sepsis, inflammatory bowel disease or other inflammatory disease, or renal failure. It is expected that such (genetically) modified phosphatases of human origin are not or very little immunogenic. However it is clear to the skilled person that if a modified phosphatase is for example used in "in vitro" or "ex vivo" diagnostics a modified phosphatase may well be
25 composed of for example a human and an *E.coli* alkaline phosphatase or may be composed of a bovine and an *E.coli* alkaline phosphatase.

In yet another preferred embodiment, said phosphatase is an alkaline phosphatase.

The metal ion-coordinating phosphate-binding pocket is, at least for
30 the human alkaline phosphatase isoforms, conserved. It consists of two Zn

binding stretches and one Mg binding stretch that contain the amino acids Asp316, His320 and His432 for Zn1, Asp42, Asp 357 and Asp358 for Zn2 and Ser155 and Glu31 for Mg, respectively (reference is made to ALPP Figure 1)

5 Mutations of the coordinating amino acid residues and/or residues located in the vicinity of the coordinating residues are likely to affect the catalytic properties of the resulting mutant enzyme in a positive or negative manner. For example, in ALPP amino acid residues 44, 87, 93, 322, 323 and 429 are located in the vicinity of the of the coordinating residues. Mutagenesis of these residues by substitution with one, two, three or four of the
10 corresponding amino acids of ALPI may affect the catalytic properties of the enzyme. Vice versa, substitution of amino acid residues 44, 87, 93, 322, 323 and 429 of ALPI with the corresponding amino acids of ALPP may affect the catalytic properties of the ALPI enzyme. Tables 4 and 5 show (combinations of) the amino acids that may be substituted.

15 Hence, in a preferred embodiment, the invention provides an isolated or recombinant phosphatase comprising a mutation in the vicinity of a catalytic residue and/or in a metal ion-coordinating phosphate-binding pocket, wherein said mutation is a mutation as depicted in Table 4, 5 or 6.

It is clear to the skilled person that an isolated or recombinant
20 phosphatase comprising a mutation in the vicinity of a catalytic residue and/or in a metal ion-containing phosphate-binding pocket can be further modified to for example comprise a modification in the GPI signal sequence. Such a mutant can even be further modified by domain of the catalytic and crown domain, i.e. such that said crown domain and catalytic domain that are
25 obtained from different phosphatases.

In yet another embodiment, an isolated or recombinant phosphatase comprising a mutation in the vicinity of a catalytic residue and/or in a metal ion-coordinating containing phosphate-binding pocket can also be further modified by domain swapping of the catalytic and crown domain, i.e. such that

said crown domain and catalytic domain that are obtained from different phosphatases.

Moreover, in yet another embodiment the invention provides an isolated or recombinant phosphatase comprising a mutation in the vicinity of a catalytic residue and/or in a metal ion-coordinating phosphate-binding pocket.

Molecular biology techniques to arrive at any of the described (genetically) modified phosphatases are well known by the skilled person and include techniques such as restriction enzyme incubations, ligations, PCR, introduction of mutations etc.

In yet another embodiment, the invention provide a nucleic acid sequence encoding a phosphatase as described herein, for example a nucleic acid sequence encoding a domain swapped mutant or a nucleic acid encoding a secreted phosphatase or a nucleic acid sequence encoding a secreted domain swapped mutant etc. The invention further provides a vector comprising a nucleic acid sequence encoding a phosphatase as described herein. Such a vector preferably comprises additional nucleic acid sequences such as elements necessary for transcription/translation of the nucleic acid sequence encoding a phosphatase (for example promoter and/or terminator sequences). Said vector may also comprise nucleic acid sequences coding for selection markers (for example an antibiotic) to select or maintain host cells transformed with said vector. Examples of suitable vectors are cloning or expression vectors. Any vector suitable for mediating expression in a suitable host cell may be used according to the invention, either integrated or episomally replicating in a host cell. The vector may be a plasmid, a virus (comprising a retrovirus, adenovirus, adeno-associated virus, baculovirus), cosmid, a phage or a phagemid, an episomal vector or an artificial chromosome.

Furthermore the invention also provides a host cell comprising a nucleic acid sequence or vector as described. The cell may be a eukaryotic cell,

preferably a mammalian cell, a plant cell or a yeast cell, that is suitable for production of recombinant proteins. Suitable yeast host cells comprise *Saccharomyces cerevisiae* and *Pichia pastoris*. Preferred host cells are mammalian (or more preferred human) derived cells such as BHK, HEK293,
5 CHO or PerC6™.

A nucleic acid sequence encoding a phosphatase as described herein or a vector comprising said nucleic acid sequence or a host cell comprising said nucleic acid sequence or a vector comprising said nucleic acid sequence are very useful in the production of modified phosphatases. Phosphatases comprise
10 glycosylation sites and hence the phosphatases are preferably produced in cells that provide the desired glycosylation pattern. In a preferred embodiment, the used production system is a mammalian (for example human) in vitro production platform and even more preferably the production involves large-scale production. In another preferred embodiment, the used production
15 system is a plant or yeast or mammalian (preferably non-human) platform in which an artificial human-like glycosylation pattern is introduced.

Upon testing different production methods, the inventors of the present invention surprisingly determined that the presence of Zn^{2+} ions
20 during the production of a wild type or mutant (alkaline) phosphatase can have an impact on the specific activity of the produced phosphatase. For example, the specific activity of ALPI can be increased from 30 U/mg to 750 U/mg by adding Zn^{2+} to the growth medium of the used host cell. Medium normally used for culturing host cells comprises 0.5 – 3 nM Zn^{2+} . Upon the
25 addition of Zn^{2+} up to 1 mM the specific activity drastically improved. It is therefore concluded that ALPI is a Zn^{2+} dependent phosphatase. This is in contrast to the already described catALPI/crownALPP mutant that seems to be independent of Zn^{2+} , i.e. the absence of Zn^{2+} during the culturing of the host cells does not significantly influence the specific activity of the produced

phosphatase nor does the absence of Zn^{2+} during storage and during the reaction decrease specific activity.

In yet another embodiment, the invention provides a method for producing a phosphatase comprising culturing a host cell capable of expressing
5 said phosphatase in a medium comprising Zn^{2+} and allowing the cell to produce said phosphatase. In a preferred embodiment, said host cell is a mammalian cell and in another preferred embodiment, said phosphatase is a human phosphatase. In yet another preferred embodiment, said phosphatase is an alkaline phosphatase. The method of the invention can be used to
10 produce wild type (or natural or non-genetically modified) phosphatase and can equally well be used to produce a genetically modified phosphatase, for example any of the herein described phosphatases.

In a further preferred embodiment, the invention provides a method for producing a phosphatase comprising culturing a host cell capable of
15 expressing said phosphatase in a medium comprising Zn^{2+} and allowing the cell to produce said phosphatase, said method further comprising isolating said phosphatase. The invention further provides a phosphatase obtainable by a method for producing a phosphatase comprising culturing a host cell capable of expressing said phosphatase in a medium comprising Zn^{2+} and allowing the
20 cell to produce said phosphatase.

Whether or not the herein described (genetically) modified phosphatases have a certain specific activity, a certain substrate specificity or a certain stability (e.g. pH, temperature, in vivo half-life time) can easily be
25 tested by the skilled person by using commercially available substrates and measuring with commercially available kits inorganic phosphate release upon incubation with alkaline phosphatase. Moreover, it is also possible to use the tests described herein in the experimental part to determine whether the modified phosphatases have any biological relevant activity.

As already mentioned the herein described (genetically) modified phosphatases are useful in diagnostics and in therapy. In one of the embodiments the invention provides a pharmaceutical composition comprising a modified phosphatase, for example:

- 5 - an isolated or recombinant alkaline phosphatase comprising a crown domain and a catalytic domain and wherein said crown domain and said catalytic domain are obtained from different alkaline phosphatases or
- an isolated or recombinant phosphatase comprising a modification in the glycosylphosphatidylinositol (GPI) signal sequence and wherein said
- 10 modification results in a secreted phosphatase or
- an isolated or recombinant phosphatase comprising a mutation in the vicinity of a catalytic residue;
- Or any combination thereof.

Said pharmaceutical composition optionally comprises a pharmaceutical

15 acceptable carrier, diluent or excipient.

The pharmaceutical composition may be presented in any form, for example as a tablet, as an injectable fluid or as an infusion fluid etc. Moreover, the (genetically) modified phosphatase can be administered via different routes, for example intravenously, rectally, bronchially, or orally. Yet another

20 suitable route of administration is the use of a duodenal drip.

In a preferred embodiment, the used route of administration is intravenously. It is clear for the skilled person, that preferably an effective amount of a (genetically) modified phosphatase is delivered. As a start point 1-5000 U/kg/day can be used. If the intravenous route of administration is used,

25 a (genetically) modified phosphatase (at least for a certain amount of time) is preferably applied via continuous infusion.

The compositions may optionally comprise pharmaceutically acceptable excipients, stabilizers, activators, carriers, permeators, propellants, disinfectants, diluents and preservatives. Suitable excipients are commonly

30 known in the art of pharmaceutical formulation and may be readily found and

applied by the skilled artisan, references for instance Remington's Pharmaceutical Sciences, Mace Publishing Company, Philadelphia PA, 17th ed. 1985.

For oral administration, the secretable AP can, for example, be
5 administered in solid dosage forms, such as capsules, tablets (preferably with an enteric coating), and powders, or in liquid dosage forms, such as elixirs, syrups, and suspensions. AP can be encapsulated in gelatin capsules together with inactive ingredients and powdered carriers, such as glucose, lactose, sucrose, mannitol, starch, cellulose or cellulose derivatives, magnesium
10 stearate, stearic acid, sodium saccharin, talcum, magnesium carbonate and the like. Examples of additional inactive ingredients that may be added to provide desirable colour, taste, stability, buffering capacity, dispersion or other known desirable features are red iron oxide, silica gel, sodium lauryl sulphate, titanium dioxide, edible white ink and the like. Similar diluents can be used
15 to make compressed tablets. Both tablets and capsules can be manufactured as sustained release products to provide for continuous release of medication over a period of hours. Compressed tablets can be sugar coated or film coated to mask any unpleasant taste and protect the tablet from the atmosphere, or enteric-coated for selective disintegration in the gastrointestinal tract. Liquid
20 dosage forms for oral administration can contain colouring and flavouring to increase patient acceptance.

In a preferred embodiment the compositions comprising a source of a (genetically) modified phosphatase are suitable for oral administration and comprise an enteric coating to protect the AP from the adverse effects of
25 gastric juices and low pH. Enteric coating and controlled release formulations are well known in the art. Enteric coating compositions in the art may comprise of a solution of a water-soluble enteric coating polymer mixed with the active ingredient(s) such as a (genetically) modified phosphatase and other excipients, which are dispersed in an aqueous solution and which may
30 subsequently be dried and/or pelleted. The enteric coating formed offers

resistance to attack of a (genetically) modified phosphatase by atmospheric moisture and oxygen during storage and by gastric fluids and low pH after ingestion, while being readily broken down under the alkaline conditions which exist in the lower intestinal tract.

5

The above described pharmaceutical compositions are very useful in the treatment of for example sepsis, inflammatory bowel disease or other inflammatory disease, and/or renal failure.

In another embodiment therefore, the invention provides a
10 (genetically) modified phosphatase as described herein for use as a medicament, preferably for treating sepsis, inflammatory bowel disease, renal failure, and inflammations preferably selected from the group consisting of rheumatoid arthritis, asthma, chronic obstructive pulmonary disease, atherosclerosis, inflammatory bowel disease, sepsis, neurodermitis and
15 diseases depicted in Table 10.

In yet another embodiment, the invention provides the use of a (genetically) modified phosphatase as described herein in the preparation of a medicament for the treatment of sepsis, inflammatory bowel disease or other inflammatory disease, and/or renal failure.

20 Sepsis is considered present if infection is highly suspected or proven and two or more of the following systemic inflammatory response syndrome (SIRS) criteria are met:

- Heart rate > 90 beats per minute
- Body temperature < 36 (96.8 °F) or > 38 °C (100.4 °F)
- 25 • Hyperventilation (high respiratory rate) > 20 breaths per minute or, on blood gas, a P_aCO_2 less than 32 mm Hg
- White blood cell count < 4000 cells/mm³ or > 12000 cells/mm³ (< 4 x 10⁹ or > 12 x 10⁹ cells/L), or greater than 10% band forms (immature white blood cells).

Consensus definitions however continue to evolve with the latest expanding the list of signs and symptoms of sepsis to reflect clinical bedside experience. The more critical subsets of sepsis are severe sepsis (sepsis with acute organ dysfunction) and septic shock (sepsis with refractory arterial hypotension).

5 Alternatively, when two or more of the systemic inflammatory response syndrome criteria are met without evidence of infection, patients may be diagnosed simply with "SIRS." Patients with SIRS and acute organ dysfunction may be termed "severe SIRS." Patients are defined as having "severe sepsis" if they have sepsis plus signs of systemic hypoperfusion; either
10 end organ dysfunction or a serum lactate greater than 4 mmol/dL. Patients are defined as having septic shock if they have sepsis plus hypotension after an appropriate fluid bolus (typically 20 ml/kg of crystalloid). The invention provides the insight that a (genetically) modified phosphatase according to the invention is especially suitable for the treatment of sepsis. In case of sepsis a
15 (genetically) modified phosphatase as described herein is preferably administrated intravenously.

Inflammatory bowel disease (IBD) is a group of inflammatory conditions of the large intestine and, in some cases, the small intestine. The main forms of IBD are Crohn's disease and ulcerative colitis (UC). Accounting for far fewer
20 cases are other forms of IBD: Collagenous colitis, Lymphocytic colitis, Ischaemic colitis, Diversion colitis, Behçet's syndrome, Infective colitis, Indeterminate colitis. The main difference between Crohn's disease and UC is the location and nature of the inflammatory changes in the gut. Crohn's can affect any part of the gastrointestinal tract, from mouth to anus although a
25 majority of the cases start in the terminal ileum. Ulcerative colitis, in contrast, is restricted to the colon and the anus. Microscopically, ulcerative colitis is restricted to the mucosa (epithelial lining of the gut), while Crohn's disease affects the whole bowel wall. Finally, Crohn's disease and ulcerative colitis present with extra-intestinal manifestations (such as liver problems, arthritis,
30 skin manifestations and eye problems) in different proportions. In rare cases,

patients have been diagnosed with both Crohn's disease and ulcerative colitis, though whether it is a combination or simply unidentifiable as one or another is uncertain. Although very different diseases, both may present with any of the following symptoms: abdominal pain, vomiting, diarrhea, hematochezia, weight loss and various associated complaints or diseases (arthritis, pyoderma gangrenosum, primary sclerosing cholangitis). Diagnosis is generally by colonoscopy with biopsy of pathological lesions. In case of IBD a (genetically) modified phosphatase as described herein is preferably administrated via an enteric coated tablet or via duodenal drip.

Next to the group of inflammatory bowel disease, the invention provides the insight that a phosphatase according to the invention is suitable for treating other inflammatory diseases. Inflammatory diseases can affect diverse organs, such as lungs, joints, liver, pancreas, skin, or even nervous tissue. Table 10 gives an unlimiting list of organs which may be inflammatory affected. A wide variety of etiologic agents have been shown to cause or sustain such inflammatory diseases. Non-limiting examples of said etiologic agents are microbes (bacteria, fungi, virii), allergens, autoimmune, trauma, and ischemia / reperfusion. Although the causative agent and etiology may be very diverse, (sub)chronic inflammation results from a deranged immune reaction. It is commonly thought that such derailed immune reaction is sustained through a vicious spiral that includes inflammation-induced tissue damage that in turn activates the immune system. Amongst others, ATP has been shown to play a role in the above mentioned vicious spiral. The invention provides the insight that a (genetically) modified phosphatase as described herein, is able to dephosphorylate ATP and thus break said vicious spiral, which is beneficial to the individual suffering from said inflammatory disease. When treating such an inflammatory disease, a (genetically) modified phosphatase according to the invention is preferably administrated intravenously or, if feasible locally, for instance, intra-articular in the case of rheumatoid arthritis, intrathecal in the case of inflammation of the (central) nervous system, intrabronchial in the case

of (allergic) asthma, or topical in the case of e.g. neurodermitis. Furthermore, (sub)chronic inflammatory reactions, such as sepsis, Crohn's disease, rheumatoid arthritis and the like, have been described to be accompanied with (serum) zinc deficiencies. A phosphatase according to the invention, wherein
5 said phosphatase does not lose its enzymatic activity when provided in a zinc deficient environment, is especially suitable for treating said inflammatory diseases. The invention provides the insight that a phosphatase according to the invention, wherein said phosphatase comprises a catalytic domain of ALPI and a crown domain of ALPP retains its activity at sub-physiological Zn^{2+}
10 concentrations as low as $0.01 \mu\text{M}$.

In one embodiment therefore, the invention provides for the use of a phosphatase comprising a catalytic domain of ALPI and a crown domain of ALPP for the dephosphorylation of a substrate, preferably an adenosine phosphate, in an environment comprising a Zn^{2+} concentration lower than $10 \mu\text{M}$, preferably a Zn^{2+} concentration lower than $1 \mu\text{M}$, more preferably a Zn^{2+}
15 concentration lower than $0.1 \mu\text{M}$.

Acute renal failure (ARF) is defined as an acute loss of kidney function that results in an increase of the serum creatinine level. In acute renal failure,
20 the glomerular filtration rate decreases over days to weeks. As a result, excretion of nitrogenous waste is reduced, and fluid and electrolyte balances cannot be maintained. Patients with acute renal failure are often asymptomatic, and the condition is diagnosed by observed elevations of blood urea nitrogen (BUN) and serum creatinine levels. Complete renal shutdown is
25 present when the serum creatinine level rises by at least $0.5 \text{ mg per dL per day}$ and the urine output is less than 400 mL per day (oliguria). The herein described (genetically) modified phosphatases can not only be used in the treatment of renal failure but also to improve renal function, especially in cases where the renal function is at least partly impaired/reduced. In a
30 preferred embodiment, the used route of administration is intravenously. It is

clear for the skilled person, that preferably an effective amount of a (genetically) modified phosphatase is delivered. As a start point 1-5000 U/kg/day can be used. If the intravenous route of administration is used, a (genetically) modified phosphatase (at least for a certain amount of time) is preferably applied via continuous infusion.

In yet another embodiment, the invention provides a method for treating a subject suffering from sepsis, inflammatory bowel disease or other inflammatory disease, and/or renal failure comprising administering to said subject an effective amount of any of the herein described modified phosphatases.

Besides the fact that a (genetically) modified phosphatase as described herein can be incorporated in a pharmaceutical composition such a phosphatase can also be part of a nutritional composition.

In a preferred embodiment of the current invention the source of a (genetically) modified phosphatase is a (genetically) modified phosphatase which is preferably produced or isolated from milk, preferably bovine milk. The milk may be obtained from animals that have been bred or genetically modified to produce elevated levels of a (genetically) modified phosphatase in their milk as compared to wild-type animals. The preparation of a (genetically) modified phosphatase enriched fractions from milk is known in the art. For instance the milkfat globule membrane enriched or derived fraction is the preferred a (genetically) modified phosphatase enriched milk fraction and may be routinely obtained by conventional skimming of raw milk. A (genetically) modified phosphatase isolated from milk may be formulated in pharmaceutical compositions and in food compositions or in nutraceuticals.

In a preferred embodiment a (genetically) modified phosphatase containing composition for oral administration of a (genetically) modified phosphatase to the mucosa of the gastrointestinal tract according to the

current invention is a food product or nutraceutical enriched for a (genetically) modified phosphatase. In one embodiment the food product may be a plant, fruit or vegetable, optionally genetically modified to contain an enhanced level of a (genetically) modified phosphatase. In another embodiment a (genetically) modified phosphatase containing food product or nutraceutical is a dairy product. In particular preparations and compositions containing non-pasteurised milk or fractions thereof, preferably bovine milk, contain high levels of a (genetically) modified phosphatase and are particularly suited for oral administration as a source of a (genetically) modified phosphatase according to the current invention.

The current invention also pertains to a method for the preparation of a (genetically) modified phosphatase enriched dairy product, preferably milk, a milk fraction or milk product. The method comprises the fractionation of raw milk, preferably bovine milk, pasteurisation of the fractions not containing or not rich in a (genetically) modified phosphatase and reformulating said fractions with the unpasteurised, a (genetically) modified phosphatase rich fractions, to obtain a less perishable and a (genetically) modified phosphatase enriched dairy product. The non pasteurised a (genetically) modified phosphatase rich fractions may be sterilised by other means, such as, but not limited to, irradiation with UV-, X- or gamma-rays, filtration, pressure, osmotic pressure, chemicals or antibiotics, ensuring that the a (genetically) modified phosphatase enzyme remains substantially active and that the milkfraction becomes substantially sterile. This dairy product may be used in compositions or administered directly to subjects suffering from or at risk of developing sepsis, IBD or renal failure. However, a (genetically) modified phosphatase enriched dairy product may also be offered to healthy subjects as a pharmaceutical or nutraceutical product for the preservation of intestinal structural integrity.

Moreover, a modified phosphatase of the invention can also be added to a nutrient (such as milk) instead of being produced in said nutrient. Moreover, tablets and/or capsules can be prepared which are subsequently added to a nutrient or which can be taken directly by a human being.

5

The invention will be explained in more detail in the following, non-limiting examples.

Experimental part

Materials and methods

Example 1

5 Dephosphorylation of the biologically active substrate ATP by different phosphatases

ATPlite™ Kit obtained from Perkin Elmer containing the following reagents: Mammalian lysis solution, Substrate buffer solution, Lyophilized substrate solution and Lyophilized ATP standard.

- 10 *Preparation of lyophilized ATP standard:* Reconstitute a vial of lyophilized ATP standard solution with MiliQ so that a 10 mM stock solution is obtained. After addition of MiliQ, allow ATP to dissolve completely by swirling for one minute.

Determination of ATP dephosphorylating activity:

- 15 - Prepare 6 standard curves with a start concentration of 20 μ M ATP and make a serial dilution. Final volume should be 100 μ l per well.
- Prepare a phosphatase enzyme activity of 1 U/ml. Prepare 3 standard curves with start concentration 1 U/ml and make a
- 20 serial dilution. Final volume should be 50 μ l.
- Prepare a 40 μ M ATP solution
- Add together 50 μ l of the 1U/ml phosphatase enzyme solution standard curve and 50 μ l of the 40 μ M ATP solution in a 96-well black Optiplate™ (Perkin Elmer). The final volume in each well
- 25 will be 100 μ l.
- Shake plate and incubate 90 min. by 37°C.
- Allow the reagents to equilibrate to room temperature
- Reconstitute one lyophilized substrate solution vial by adding the proper amount of substrate solution buffer, according to the

manufacturer's recommendations. Agitate gently until the solution is homogeneous.

- Add 50 µl of the mammalian cell lysis solution and shake the plate for 5 minutes at 700 rpm.
- 5 - Add 50 µl of the substrate solution to the well and shake the plate for 5 minutes at 700 rpm.
- Remove air bubbles by spraying alcohol 70 % over plate
- Dark adapt the plate for 10 minutes and measure the luminescence on the Viktor3™ Multi Label Reader (Perkin
10 Elmer)

Example 2

Liver-slice assay using different isoforms of alkaline phosphatase

Rats were sacrificed under O₂/N₂O/Forene anaesthesia and livers were
15 taken out and stored in University of Wisconsin organ preservation solution (UW) until slice preparation. Cores (diameter, 5 mm) were made from the pieces of liver tissue and stored in ice-cold UW solution until slicing. The slicing was performed with a Krumdieck slicer. Ice-cold KHB supplemented with glucose to a final concentration of 25 mM was used as slice buffer. Rat
20 liver slices (thickness, 200–250 µm; wet weight, ± 3 mg) were prepared with standard settings (cycle speed, 30; interrupted mode). After slicing, the rat liver slices were stored in UW solution until the start of the experiment.

Slices were incubated individually at 37°C in 12-well plates (Greiner, Alphen a/d Rijn, The Netherlands) in 1.3 ml Williams' medium E
25 supplemented with Glutamax I (Gibco BRL, Paisly), Scotland). 50 mg/ml gentamicin (Gibco BRL) and saturated with 95% O₂ / 5% CO₂. Slices were incubated for 24 hr with or without 10 µg/ml LPS and with or without different isoforms of AP in different concentrations. Medium of the slices were stored at -80°C until NO_x measurement.

NO_x (the sum of NO, NO₂ and NO₃) was measured by adding 5 µl of a mixture of 0.275 µl 100 mM NADPH, 2.2 µl 10 U/ml nitrate reductase, and 0.055 µl 10 mM FAD, which was diluted 2-fold in water, to 110 µl of supernatant. After 30 minutes incubation at 37°C, 5 µl of a second mixture; 2.2 µl Na-pyruvate 0.5 M and 0.55 µl of 5 mg/ml LDH, diluted 1:0.82 in water, was added. After 5 minutes incubation at 37°C, 5 µl of 30% ZnSO₄ was added and samples were centrifuged at 2000 rpm. 100 µl of the supernatant was then transferred to a new plate and mixed with 100 µl of Griess-reagents containing 0.1% sulphanilamide, 0.01% n-Naphtyl-Ethylene-Diamin and 2.5% phosphoric acid. Finally, absorbance was measured at 550 nm on a microplate reader. The absorbance was related to the absorbance of a standard curve of sodium nitrate.

Example 3

15 Biological effect of dephosphorylation of extracellular ATP

The murine macrophage cell line RAW264.7 and the human epithelial cell lines T84 (colorectal carcinoma) were obtained from the American Type Culture Collection (ATCC, Rockville, MD, USA) and were maintained in DMEM/F12 (1:1) and DMEM medium containing 4.5 g/l glucose, respectively. Both media were obtained from Invitrogen Corp. (Breda, The Netherlands), contained Glutamax I and were supplemented with 10 % heat –inactivated FBS (Wisent Inc. (Quebec, Canada), 100 U/ml penicillin and 100 µg/ml streptomycin (both from Intvitrogen Corp.). T84 cell line was subcultured at confluence by employing trypsin-EDTA, RAW264.7 cells were scraped using a rubber policeman.

4x10⁵ T84 cells were plated into 12 well plates (Nunc, Roskilde, Denmark) with 2 ml of medium. On reaching confluency, medium was refreshed and cells were incubated with (Figure 3) or without (Figure 2) 1 µg/ml LPS for 2 hours in the presence or absence of different concentrations of ATP and/or AP as depicted in Figures (checkerboard). After 2 hours, the

supernatant was collected and transferred into 24 well plates containing RAW264.7 cells. These RAW264.7 cells were plated the day before into 24 well plates (Nunc, Roskilde, Denmark) at a density of 2×10^5 in one ml of medium. Prior to transfer of 1 ml of the supernatants of the T84 cells, the supernatant
5 of the RAW was aspirated and discarded.

The RAW cells were incubated with the T84 supernatants for 24 hours at 37°C at 5% CO₂, before the supernatants were collected and checked for cytokine (IL-6, TNF α) content using a commercial available ELISA (Biosource Europe SE, Nivelles, Belgium).

10

Example 4

Preparation of mutants

The mutants as herein in described and more specific the mutants as described in Tables 4, 5 and 6 are/were prepared by using standard molecular
15 biology techniques.

The amino acid positions as mentioned in Tables 4 to 6 correspond to the sequences as depicted in Figure 1. Only the mutated positions are indicated, i.e. only the deviations from the wild type sequences are given. For example mutant 1 of Table 4 is, if compared to the given wild type sequence, unchanged
20 at positions 87, 93 and 429, i.e. position 87 is a K, position 93 is a G and position 429 is an E.

The mutants will be prepared and checked by standard molecular biology techniques, such as PCR site-directed mutagenesis, restriction enzyme analysis, and sequence analysis.

25

Example 5

At t=0 different recombinant alkaline phosphatases containing 450 ± 50 Units were diluted 4000x in diluent buffer (0.025M glycine/NaOH pH 9.6 / 1 mM MgCl₂ / 1% mannitol / 0.05% BSA) with different concentrations of Zn²⁺.
30 In a second example, BSA as a possible Zn²⁺ source was omitted from the

buffer. Within 1 minute after dilution ($T=0$), a sample was taken and measured for alkaline phosphatase activity using the pNPP assay described below. 90 min ($T = 1\frac{1}{2}$ h), 180 min ($T = 3$ h) and 22 hour ($T = 22$ h) later, each time a sample was taken and directly measured for alkaline phosphatase activity using the pNPP assay as described below. Activity in U/ml was backcalculated by the specific activity (U/mg) by dividing the obtained result in U/ml through the protein content previously obtained using a commercially available BCA kit (Pierce).

pNPP phosphatase activity measurement:

10 Working substrate:

Temperature of reagents, substrates, samples and incubation chamber were set to 25°C. The spectrophotometer was set to a wavelength of 405 nm and the light path was 1 cm.

In a disposable cuvette 50 μ l of the test substance and 1450 μ l of the working substrate were pipetted, mixed and immediately the cuvette was placed in the spectrophotometer and the increase in absorbance at 405 nm was recorded for 3 minutes.

The activity per volume was calculated using the following equation:

$$\text{activity (U/ml)} = \Delta E_{405}/\text{min} \times 1.6 \times \text{dilution factor}$$

20 Note: measurement range should be between 0.04 and 0.4 U/ml

Example 6

Effect of pH on stability of different phosphatases.

The pH stability of transiently expressed alkaline phosphatases in HEK293 cells was investigated. A pH range of 4.0-9.0 was tested using 5 different 0.1M (sodium acetate, MES (2-(N-morpholino)ethanesulfonic acid), MOPS (3-N-morpholino)propanesulfonic acid), Tris (2-amino-2-hydroxymethyl-propane-1,3-diol) and glycine) buffers adjusted for their specific pH at 25°C. BiAP stabilized with 1% mannitol was used as reference. The alkaline phosphatase solutions were diluted in the respective buffers to

approximately 100U/ml, stored for 24h at room temperature and determined for enzymatic activity using the *pNPP phosphatase activity* assay as described above.

5 Example 7

Effect of zinc on the expression of the different phosphatases.

The influence of different zinc salts on the activity of transiently expressed alkaline phosphatases in HEK293 cells was investigated using small scale transfections in culture medium supplemented with 1mM MgCl₂ +
10 0.1mM Zinc salt (ZnCl₂, or ZnSO₄ or ZnAc₂). HEK293 cells were transfected with shIAP, shPLAP, Xinplap (catALPI/crownALPP) and sALPP-ALPI-CD (catALPP/crownALPI) and at t=144h samples were taken and analysed for enzymatic activity using the *pNPP phosphatase activity* assay as described above.

15

Example 8

Effect of zinc on the stability of the different phosphatases.

Solutions containing approximately 20U/ml of different (recombinant) isoforms of alkaline phosphatase in the presence or absence of 100μM of ZnCl₂
20 were prepared for temperature stability testing. The samples were stored at RT, 37°C and 56°C and enzyme activity (pNPP) was determined at t=0, 2h and 24h using the *pNPP phosphatase activity* assay as described above.

Example 9

25 Generation of fusion proteins and mutants of alkaline phosphatase.

To select a suitable expression system, the human intestinal (hIAP) and human placental (hPLAP) cDNA, coding for the mature proteins, either with or without the GPI anchor sequence, were cloned into several vectors. After small scale infections, the expression vector used was CMV driven, contained
30 the cystatin signal sequence and harbored the N-terminal HIS tag to facilitate

purification. After successfully expressing both secretable hIAP and hPLAP, two fusion human secretable alkaline phosphatases were constructed in silico, one based on the backbone of intestinal AP containing the placental crown domain (aa 360-430 of the mature sequence) and the second based on the
5 backbone of placental AP with the crown domain of intestinal AP (the first referred to as RecAP or catALPI/crownALPP and second referred to as shPLAP-shIAP-CD or catALPP/crownALPI). The two genes were synthesized containing the human cystatin signal sequence and cloned in the CMV promotor containing expression vector.

10

Example 10

Effect of GPI anchor on phosphatase specific activity

Several recombinant forms of AP were produced, purified and evaluated for enzymatic activity using the *pNPP phosphatase activity* assay as described
15 above. Furthermore, protein amount in the samples was measured using SDS-PAGE and GelEval software. Specific activity was calculated by dividing activity (U/ml) by protein concentration (mg/ml) and expressed as U/mg.

Experimental part

Results

Example 1

5 **Dephosphorylation of the biologically active substrate ATP by different phosphatases**

ATP at a final concentration of 20 μ M was incubated with different concentrations of BIAP, sALPP, sALPI or the chimera catALPI/crownALPP. From Table 9 it is obvious that pNPP chemical activity is not 1:1 related to the activity towards a biological substrate, e.g. ATP. Whereas BIAP and sALPI show more than 50 % dephosphorylation of ATP after 90 minutes at 37°C at concentrations of 0.031 and 0.004 pNPP units, respectively, ALPP and catALPP/crownALPP can only dephosphorylate this amount at concentrations of 0.125 and 0.0625 pNPP units, respectively.

15

Example 2

Liver-slice assay using different isoforms of alkaline phosphatase

Upon stimulation with LPS (10 μ g/ml), liver slices produce NO_x. Figure 6 shows that in the presence of different recombinant human alkaline phosphatases (sALPI, sALPP, GPI-anchored ALPI, catALPI/crownALPP) at different concentrations, the NO_x production was significantly inhibited. In this experiment, bovine derived ALPI was used as a positive control and solvent as a negative control.

Without the addition of LPS, NO_x production by the liver slices was less than 10 μ M and was not significantly altered by the presence of the different phosphatases during incubation (data not shown). From this it is concluded that all tested recombinant isoforms of human alkaline phosphatases, regardless of the presence of a GPI anchor have activity towards a biological substrate that is involved in the LPS-induced NO_x production.

30

Example 3

Biological effect of dephosphorylation of extracellular ATP

The production of IL-6 by RAW264.7 cells upon incubation with T84 supernatant was dependent on LPS. Without LPS (Figure 2), low levels of IL-6 were produced. ATP as well as AP could not alter the levels of IL-6 produced by RAW264.7 cells. In contrast, when LPS was present, the production of IL-6 by RAW264.7 incubated with T84 supernatant increased with increasing concentrations of ATP. This increase in cytokine production by ATP could be completely diminished by AP concentrations of 0.1 U/ml and higher (Figure 3).

Similar results (not shown) were obtained for TNF α using the same protocol and for TNF α and IL-6 using HT29 instead of T84 as a source of epithelial cells.

It is concluded that AP dephosphorylates ATP and thereby decreases its LPS-potentiating effect. It is unlikely that AP dephosphorylates LPS as no effect of AP on LPS was seen. The combination of ATP and LPS could only be diminished by AP to levels equal to LPS alone.

Example 5

Determination of Zn²⁺ dependency of different phosphatases

As depicted in Figure 4 and Table 7, Zn²⁺ significantly stabilized the specific activity of sALPI, whereas sALPP and the chimera catALPI/crownALPP retained their initial specific activity independent of the presence of Zn²⁺ in the medium. In this experiment, BSA, routinely used in the pNPP phosphatase activity assay, may be a source of trace amounts of Zn²⁺. Therefore a second experiment was performed. Table 8 and Figure 5 show that in the absence of BSA and in the presence of Zn²⁺ depleting chelator EDTA, all isoforms lose their specific activity after 22 h. However addition of physiological (0.5 – 10 nM) concentrations of Zn²⁺ during storage and during the reaction preserved more than 60, respectively 75% of the initial specific activity of sALPP and

catALPI/crownALPP, whereas more than 95% of the initial specific activity of sALPI was lost. Unphysiologically high concentrations of Zn^{2+} were necessary to preserve sALPI's specific activity.

It is therefore concluded that the chimer catALPI/crownALPP shows
5 high specific activity, comparable with that of sALPI, combined with Zn^{2+} independent specific activity, comparable with that of sALPP. It is concluded that the Zn^{2+} dependency is located in the sALPI crown domain, whereas the high specific activity is located in the sALPI catalytic domain.

10 Example 6

Effect of pH on stability of different phosphatases.

BiAP, shIAP, shPLAP, RecAP were diluted and stored in 0.1M (sodium acetate, MES (2-(N-morpholino)ethanesulfonic acid), MOPS (3-N-morpholino)propanesulfonic acid), Tris (2-amino-2-hydroxymethyl-propane-1,3-
15 diol) and glycine) buffers with pH values of 4.0, 5.0, 6.0, 7.0, 8.0 and 9.0 at RT for 24h. For each pH value the initial enzyme activity and the enzyme activity after storage was determined according to the standard pNPP assay. As expected and indicated by its name, alkaline phosphatase activity is optimal in the high pH range (alkaline environment). However, as Figure 7 shows, the
20 placental type is more stable than the intestinal isoforms. Furthermore, the results demonstrate that this stability, especially in the high pH range, is determined by the crown domain and not by the catalytic domain, as catALPI/crownALPP (recAP) follows the stability profile of the shPLAP and is stable between pH 5-9, whereas the stability of the BIAP and shIAP is
25 restricted to pH 7-8. In contrast, catALPP/crownALPI (shPLAP-hIAP-CD) is less stable than the placental type AP and catALPI/crownALPP at pH 9. This demonstrates that catALPI/crownALPP retains its activity in a much broader range of pH values than the native APs or the reverse chimer (catALPP/crownALPI) tested.

Example 7

Effect of zinc on the expression of different phosphatases.

Using small scale transfections in HEK293 cells, the influence of different zinc salts on the activity of transiently expressed alkaline phosphatases was investigated. In order to do so, shIAP, shPLAP, Xinplap (catALPI/crownALPP) and sALPP-ALPI-CD (catALPP/crownALPI) were transfected and at t=144h samples were taken and analysed for enzymatic activity. The obtained results revealed that there was a concentration dependency for Zn²⁺ ions regarding the enzymatic activity, but that the increase of activity observed was independent for the type of zinc salt used.

From the results presented in figure 8 it is concluded that the obtained enzyme activity is dependent on zinc, but that it is independent of which kind of zinc salt is used. Furthermore, it was shown that addition of zinc is most favorable for the production of shIAP with an induction factor for enzyme activity of >30. For sALPP and Xinplap the activity enhancement was <2 and >4, respectively. The reverse chimera (sALPP-ALPI-CD, also called catALPP/crownALPI) showed negligible activity if produced in the absence of zinc, whereas it obtained activity comparable with sALPP in the presence of zinc.

Furthermore, Xinplap showed a much higher enzymatic activity without the addition of Zn/Mg ions than any of the other tested APs (factor 5) and is less dependent on Zn/Mg as compared to shIAP or the reverse chimera, catALPP/crownALPI.

25 Example 8

Effect of zinc on the stability of the different phosphatases.

Figure 9 shows the stability profile of different APs in time at different temperatures and in the presence or absence of 100 µM ZnCl₂. It is concluded from the results that, in the absence of zinc, the intestinal isoforms (sALPI and BIAP) of alkaline phosphatase are highly temperature sensitive with respect

to the enzyme activity. At 37°C BIAP lost 20% of its activity within 2 hours and after 24h only 20% enzymatic activity remains. shIAP shows a decline in activity of 30% after 24h storage at 37°C. Figure 9C shows that the presence of zinc during stability testing at 37°C protects the enzyme from degradation.

- 5 However, at 56°C, zinc no longer protects the intestinal isoforms and both lose their enzymatic activity almost completely in the first 2 hours. In contrast, RecAP (catALPI/crownALPP) and shPLAP (sALPP) show excellent stability in the presence or absence of 100µM up to 22 hours even at 56°C.

10 Example 9

Generation of fusion proteins and mutants of alkaline phosphatase.

- Secretable hIAP and secretable hPLAP were expressed in HEK293 using a CMV promotor driven, cystatin signal sequence containing vector and contained a HIS tag on the N-terminal part of the protein. The expressed
- 15 proteins were purified through the HIS tag and analyzed by SDS PAGE (see figure 10). The protein yield of 1 Liter of suspension culture for secretable hIAP and hPLAP was 38 and 16 mg, respectively and showed >95% purity. The specific activity of hIAP and hPLAP was 21 and 100 U/mg, respectively.

- Based upon these two secretable recombinant phosphatases, two
- 20 domain-swap variants were designed in silico, and expressed in HEK293 cells. A remarkable result was obtained for RecAP, which consists of an intestinal alkaline phosphatase backbone with the crown domain of placental alkaline phosphatase. RecAP showed a five times higher enzymatic activity as compared with secretable hIAP (14 U/ml vs 2.7 U/ml) under conventional
- 25 culture conditions. However, the addition of zinc salt up to 100 µM during cell culture and phosphatase expression has only minor influence on the enzymatic activity of RecAP (18.8 U/ml vs 14.0 U/ml without addition zinc), whereas it significantly improves activity of secretable hIAP (37.9 U/ml vs 2.7 U/ml without addition of zinc). The domain-swap variant based on the backbone of
- 30 placental AP with the crown domain of intestinal AP (catALPP/crownALPI) on

the other hand, shows zinc dependency comparable with the intestinal form and maximum activity comparable with secretable PLAP. The observed increase in activity of shIAP and catALPP/crownALPI could not be achieved if zinc was added to the enzyme after expression and purification. It is therefore
 5 concluded that the addition of 100 μ M zinc during expression of alkaline phosphatases, preferably those comprising a crown domain of the intestinal form, results in a higher yield of alkaline phosphatase activity.

Example 10

10 Effect of GPI anchor on phosphatase specific activity

Several recombinant forms of AP were produced and evaluated for enzymatic activity. The results presented in Figure 11 demonstrate that bovine intestinal alkaline phosphatase (BIAP) and recAP (catALPI/crownALPP) display similar specific activity that is superior to that
 15 of secretable hIAP, secretable hPLAP and the GPI-anchored hIAP. The purity of the enzyme is important for the specific enzyme activity. It should therefore be noted that the four human enzymes were purified in a laboratory setting, whereas BIAP is obtained as an ultrapure GMP batch. It should further be noticed that the secretable hIAP has a specific activity that is approximately 2
 20 fold higher than that of GPI-anchored hIAP. Without being limited to theory, the higher specific activity can either be structural related or purity related, as secretable AP is more easily purified than GPI-anchored (membrane bound) AP.

25 Tables

Table 1: HEK293 expression levels (U/ml) of alkaline phosphatases after t=144h and the effect on the activity after overnight incubation with 0.1mM ZnCl₂

Construct	Units/ml	Units/ml after o/n with 0.1mM
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		ZnCl₂
sALPP N-his tag	3.34	3.55
sALPP C-his tag	3.75	3.97
sALPI N-his tag	1.55	1.92
sALPI C-his tag	7.52	8.07
Native sALPI	2.84	4.94

Table 2: HEK 293 expression levels (U/ml) of alkaline phosphatases after t=144h and the effect on the activity of different ZnCl₂ and/or MgCl₂ concentrations in culture medium

Construct	Enzyme activity in Units/ml					
	No addition	0.05mM Zn ²⁺	0.1mM Zn ²⁺	1mM Mg ²⁺	5mM Mg ²⁺	0.1mM Zn ²⁺ / 1mM Mg ²⁺
sALPI	2.2	55.8	56.6	6.4	5.9	61.3
sALPP	6.8	8.7	8.9	6.1	5.4	7.7

Table 3: HEK 293 expression levels (U/ml) of chimeric alkaline phosphatases after t=96h and the effect on the activity of ZnCl₂ in culture medium

Construct	Enzyme activity in Units/ml	
	No addition	0.1mM Zn ²⁺ / 1mM Mg ²⁺
catALPI/crownALPP	3.72	2.43

Table 4: Mutation-sites in ALPP with proposed amino acid changes

Name	Position in mature ALPP				Alternative name
	44	87	93	429	
wt	M	K	G	E	-
mut 1	L				M44L
mut 2		R			K87R
mut 3			A		G93A
mut 4				S	E429S
mut 5	L	R			M44L, K87R
mut 6	L		A		M44L, G93A
mut 7	L			S	M44L, E429S
mut 8		R	A		K87R, G93A
mut 9		R		S	K87R, E429S
mut 10			A	S	G93A, E429S
mut 11	L	R	A		M44L, K87R, G93A
mut 12	L	R		S	M44L, K87R, E429S
mut 13	L		A	S	M44L, G93A, E429S
mut 14		R	A	S	K87R, G93A, E429S
mut 15	L	R	A	S	M44L, K87R, G93A, E429S

Table 5: Mutation sites in ALPI with proposed amino acid changes

	Position in mature ALPI				Alternative name
	44	87	93	429	
wt	L	R	A	S	-
mut 16	M				L44M
mut 17		K			R87K
mut 18			G		A93G
mut 19				E	S429E
mut 20	M	K			L44M, R87K
mut 21	M		G		L44M, A93G
mut 22	M			E	L44M, S429E
mut 23		K	G		R87K, A93G
mut 24		K		E	R87K, S429E
mut 25			G	E	A93G, S429E
mut 26	M	K	G		L44M, R87K, A93G
mut 27	M	K		E	L44M, R87K, S429E
mut 28	M		G	E	L44M, A93G, S429E
mut 29		K	G	E	R87K, A93G, S429E
mut 30	M	K	G	E	L44M, R87K, A93G, S429E

Table 6: Proposed double mutations in ALPP and ALPI within the vicinity of the active site

Additional mutants		
mut 31	ALPP	S322G, R323V
mut 32	ALPI	G322S, V323R

Table 7: sALPI, but not sALPP and the chimera catALPI/crownALPP is Zn²⁺ dependent for retaining its specific activity in vitro.

sALPP	T= 0	T= 1½ h	T= 3 h	T= 22 h
0 ^a µM Zn ²⁺	122	103	105	100
10 µM Zn ²⁺	162	123	117	110
100 µM Zn ²⁺	131	128	150	132
1000 µM Zn ²⁺	126	135	125	114
sALPI	T= 0	T= 1½ h	T= 3 h	T= 22 h
0 µM Zn ²⁺	650	500	413	116
10 µM Zn ²⁺	717	648	953	746
100 µM Zn ²⁺	785	733	868	750
1000 µM Zn ²⁺	752	762	770	989
catALPI/crownALPP	T= 0	T= 1½ h	T= 3 h	T= 22 h
0 µM Zn ²⁺	426	407	455	429
10 µM Zn ²⁺	443	500	479	462
100 µM Zn ²⁺	543	560	456	442
1000 µM Zn ²⁺	452	452	441	465

^a No Zn-salt was added, however, the assay was performed in the presence of albumin (0.05%), which maybe a natural source of zinc; see result section.

Table 8: Specific activity of sALPI is decreased by more than 95% after 22h at physiological (0.01 μM) Zn^{2+} concentrations in the absence of albumin, whereas sALPP and the chimera catALPI/crownALPP retain more than 60, respectively 75 % of their initial specific activity under the same conditions.

sALPP	T= 0	T= 1½ h	T= 3 h	T= 22 h
0 Zn + 100 mM EDTA	29	0	0	0
0.01 μM Zn^{2+}	123	74	76	78
10 μM Zn^{2+}	131	94	88	92
1000 μM Zn^{2+}	118	6	6	3
sALPI	T= 0	T= 1½ h	T= 3 h	T= 22 h
0 Zn + 100 mM EDTA	290	4	10	0
0.01 μM Zn^{2+}	693	384	211	31
10 μM Zn^{2+}	785	541	386	324
1000 μM Zn^{2+}	737	505	382	232
catALPI/crownALPP	T= 0	T= 1½ h	T= 3 h	T= 22 h
0 Zn + 100 mM EDTA	161	8	5	0
0.01 μM Zn^{2+}	330	374	316	252
10 μM Zn^{2+}	543	386	315	266
1000 μM Zn^{2+}	531	306	299	196

Table 9: Dephosphorylating properties of different alkaline phosphatases towards the biological substrate ATP.

[illegible]

Table 10

Unlimiting list of inflammatory diseases and organs affected

<u>Inflammation</u>	<u>Body part</u>
<u>Appendicitis</u>	<u>Appendix</u>
<u>Arteritis</u>	<u>Arteries</u>
<u>Arthritis</u>	<u>Joint</u>
<u>Blepharitis</u>	<u>Eyelids</u>
<u>Bronchiolitis</u>	<u>Bronchioles</u>
<u>Bronchitis</u>	<u>Bronchi</u>
<u>Bursitis</u>	<u>Bursa</u>
<u>Cervicitis</u>	<u>Cervix</u>
<u>Cholangitis</u>	<u>Bile duct</u>
<u>Cholecystitis</u>	<u>Gallbladder</u>
<u>Chorioamnionitis</u>	<u>Chorion and amnion (amniotic sac)</u>
<u>Colitis</u>	<u>Colon</u>
<u>Conjunctivitis</u>	<u>Conjunctiva</u>
<u>Cystitis</u>	<u>Bladder</u>
<u>Dacryoadenitis</u>	<u>Lacrimal gland</u>
<u>Dermatitis</u>	<u>Skin</u>
<u>Dermatomyositis</u>	<u>Skin and muscles</u>
<u>Encephalitis</u>	<u>Brain</u>
<u>Endocarditis</u>	<u>Endocardium</u>
<u>Endometritis</u>	<u>Endometrium</u>
<u>Enteritis</u>	<u>Small intestine</u>
<u>Enterocolitis</u>	<u>Small intestine and large intestine</u>
<u>Epicondylitis</u>	<u>Epicondyle</u>
<u>Epididymitis</u>	<u>Epididymis</u>
<u>Fasciitis</u>	<u>Fascia</u>
<u>Fibrositis</u>	<u>Fibrous connective tissue</u>
<u>Gastritis</u>	<u>Stomach</u>
<u>Gastroenteritis</u>	<u>Stomach and small intestine</u>
<u>Gingivitis</u>	<u>Gingiva</u>
<u>Glossitis</u>	<u>Tongue</u>
<u>Hepatitis</u>	<u>Liver</u>
<u>Hidradenitis suppurativa</u>	<u>Apocrine sweat glands</u>
<u>Ileitis</u>	<u>Ileum</u>
<u>Iritis</u>	<u>Iris</u>
<u>Laryngitis</u>	<u>Larynx</u>
<u>Mastitis</u>	<u>Mammary gland</u>

Table 10 continued

<u>Meningitis</u>	<u>Meninges</u>
<u>Myelitis</u>	<u>Spinal cord</u>
<u>Myocarditis</u>	<u>Myocardium</u>
<u>Myositis</u>	<u>Muscle</u>
<u>Nephritis</u>	<u>Kidney</u>
<u>Omphalitis</u>	<u>Umbilical cord</u>
<u>Oophoritis</u>	<u>Ovaries</u>
<u>Orchitis</u>	<u>Testicle</u>
<u>Osteitis</u>	<u>Bone</u>
<u>Otitis</u>	<u>Ear</u>
<u>Pancreatitis</u>	<u>Pancreas</u>
<u>Parotitis</u>	<u>Parotid gland</u>
<u>Pericarditis</u>	<u>Pericardium</u>
<u>Peritonitis</u>	<u>Peritoneum</u>
<u>Pharyngitis</u>	<u>Pharynx</u>
<u>Pleuritis</u>	<u>Pleura</u>
<u>Phlebitis</u>	<u>Veins</u>
<u>Pneumonitis</u>	<u>Lungs (also pneumonia)</u>
<u>Proctitis</u>	<u>Rectum</u>
<u>Prostatitis</u>	<u>Prostate</u>
<u>Pyelonephritis</u>	<u>Kidney</u>
<u>Rhinitis</u>	<u>Nasal lining</u>
<u>Salpingitis</u>	<u>Fallopian tubes</u>
<u>Sinusitis</u>	<u>Sinus of the skull</u>
<u>Stomatitis</u>	<u>Mouth</u>
<u>Synovitis</u>	<u>Synovial membrane</u>
<u>Tendinitis</u>	<u>Tendon</u>
<u>Tonsillitis</u>	<u>Tonsils</u>
<u>Uveitis</u>	<u>Uvea</u>
<u>Urethritis</u>	<u>Urethra</u>
<u>Vaginitis</u>	<u>Vaginal mucosa</u>
<u>Vasculitis</u>	<u>Blood vessels or lymph vessels</u>
<u>Vulvitis</u>	<u>Vulva</u>

Description of figures**Figure 1**

Sequences of the four human alkaline phosphatase iso-enzymes. Note: these
5 are the sequences of the mature proteins (i.e. without signal sequence) but
before addition of the GPI-anchor and concomitant processing of the C-
terminal amino acids with exception of the chimeric AP's

Figure 2

10 ATP alone is not sufficient to stimulate production of IL-6 in RAW cells upon
incubation with T84 supernatant. No effect on IL-6 production is seen upon
addition of AP

Figure 3

15 Increasing ATP concentrations amplify the LPS-induced IL-6 production in
RAW cells upon incubation with T84 supernatant. Alkaline phosphatase
inhibits the amplifying effect of ATP but not the LPS-induced IL-6 production
itself.

Figure 4

20 Specific activity of catALPI/crownALPP is Zn independent

Figure 5

Specific activity of ALPI but not catALPI/crownALPP decreases in time
25

Figure 6

LPS-induced NOx production by liver slices is inhibited by different secretable
isoforms of human alkaline phosphatase

Figure 7

Relative enzyme activities of different secretable human recombinant alkaline phosphatases and bovine intestinal phosphatase (BIAP) stored for 24h in 0.1M buffers of different pH values.

5

Figure 8

Influence of different zinc salts added to the culture medium during production of different recombinant alkaline phosphatases on the enzymatic activity.

10 **Figure 9**

Stability of the enzymatic activity of different alkaline phosphatases determined after storage in the presence of zinc at: **A** room temperature, **C** 37°C and **E** 56°C or in the absence of zinc at **B** room temperature, **D** 37°C and **F** 56°C

15

Figure 10

SDS-PAGE: protein profile of purified secretable hIAP and secretable hPLAP. Lane 1) MW Marker, 4+5) secretable hIAP 10 and 25 times diluted, 12+13) secretable hPLAP 10 and 25 times diluted.

20

Figure 11

Specific enzymatic activities of 4 human and a bovine AP against para-nitrophenylphosphate (pNPP), the standard chemical substrate for AP activity determination.

25

Claims

1. An isolated or recombinant alkaline phosphatase comprising a crown domain and a catalytic domain and wherein said crown domain and said catalytic domain are obtained from different alkaline phosphatases.
2. A phosphatase according to claim 1, wherein at least one of said
5 different phosphatases is a human phosphatase
3. A phosphatase according to claim 1 or 2, wherein said crown domain is the crown domain of ALPP and wherein said catalytic domain is the catalytic domain of ALPI.
4. A phosphatase according to claim 1 or 2, wherein said crown domain
10 is the crown domain of ALPI and wherein said catalytic domain is the catalytic domain of ALPP.
5. An isolated or recombinant phosphatase comprising a modification in the glycosylphosphatidylinositol (GPI) signal sequence and wherein said modification results in a secreted phosphatase.
- 15 6. A phosphatase according to claim 5, wherein said modification comprises a mutation or a deletion of the amino acid sequence encompassing the consensus GPI signal sequence.
7. A phosphatase according to claim 5 or 6, wherein said phosphatase is a human phosphatase.
- 20 8. A phosphatase according to any one of claim 5 to 7, wherein said phosphatase is an alkaline phosphatase.
9. A phosphatase according to any one of claims 5 to 8, wherein said phosphatase is selected from human liver-kidney-bone phosphatase, human intestinal alkaline phosphatase and human placental-like alkaline
25 phosphatase.
10. A phosphatase according to any one of claims 5 to 9, further comprising a crown domain and a catalytic domain that are obtained from different phosphatases.

11. An isolated or recombinant phosphatase comprising a mutation in the vicinity of a catalytic residue and/or in a metal ion-coordinating phosphate-binding pocket.
12. A phosphatase according to claim 11, wherein said phosphatase is a human phosphatase.
13. A phosphatase according to claim 11 or 12, wherein said phosphatase is an alkaline phosphatase.
14. A phosphatase according to any one of claims 11 to 13, wherein said mutation is a mutation as depicted in Table 4, 5 or 6.
15. A phosphatase according to any one of claims 11 to 14, further comprising a modification in the glycosylphosphatidylinositol (GPI) signal sequence wherein said modification results in a non-GPI anchored phosphatase.
16. A phosphatase according to any one of claims 11 to 14, further comprising a crown domain and a catalytic domain that are obtained from different phosphatases.
17. A phosphatase according to claim 15, further comprising a crown domain and a catalytic domain that are obtained from different phosphatases.
18. A nucleic acid sequence encoding a phosphatase according to any one of claims 1 to 17.
19. A vector comprising a nucleic acid according to claim 18.
20. A host cell comprising a nucleic acid according to claim 18 or a vector according to claim 19.
21. A method for producing a phosphatase comprising culturing a host cell capable of expressing said phosphatase in a medium comprising Zn^{2+} and allowing the cell to produce said phosphatase.
22. A method according to claim 21, wherein said host cell is a mammalian cell.
23. A method according to claim 21 or 22, wherein said phosphatase is human phosphatase.

24. A method according to any one of claims 21 to 23, wherein said phosphatase is an alkaline phosphatase.
25. A method according to any one of claims 21 to 24, wherein said phosphatase is a phosphatase as described in any one claims 1 to 17.
- 5 26. A method according to any one of claims 21 to 25, further comprising isolating said phosphatase.
27. A phosphatase obtainable by a method according to any one of claims 21 to 26.
28. A pharmaceutical composition comprising a phosphatase according
10 to any one of claims 1 to 17 or 27.
29. Use of a phosphatase according to any one of claims 1 to 17 or 27 in the preparation of a medicament for the treatment of sepsis, inflammatory bowel disease or other inflammatory disease, and/or renal failure.
- 30 A method for treating a subject (preferably a human) suffering from
15 sepsis, inflammatory bowel disease or other inflammatory disease, and/or renal failure, comprising administering to said subject an effective amount of a phosphatase according to any one of claims 1 to 17 or 27.
- 31 Use of a phosphatase according to claim 3 for the dephosphorylation of a substrate, preferably an adenosine phosphate, in an environment
20 comprising a Zn^{2+} concentration lower than $10\text{ }\mu\text{M}$, preferably a Zn^{2+} concentration lower than $1\text{ }\mu\text{M}$, more preferably a Zn^{2+} concentration lower than $0.1\text{ }\mu\text{M}$.
- 32 A phosphatase according to any one of claims 1 to 17 or 27 for use as a medicament, preferably for the treatment of sepsis, inflammatory bowel
25 disease and/or other inflammatory disease, and/or renal failure.
- 33 A phosphatase according to claim 3 for use in treating a disease which is accompanied by Zn^{2+} deficiency, said disease preferably comprising an inflammatory disease, more preferably selected from the group consisting of autoimmune diseases, rheumatoid arthritis, asthma, chronic obstructive

pulmonary disease, atherosclerosis, inflammatory bowel disease, sepsis, neurodermitis, and diseases depicted in Table 10.

- 34 Use of a phosphatase according to claim 3 in the preparation of a medicament for the treatment of a disease which is accompanied by Zn^{2+}
- 5 deficiency, said disease preferably comprising an inflammatory disease, more preferably selected from the group consisting of autoimmune diseases, rheumatoid arthritis, asthma, chronic obstructive pulmonary disease, atherosclerosis, inflammatory bowel disease, sepsis, neurodermitis, and diseases depicted in Table 10.
- 10 35 A method for treating a subject (preferably a human) suffering from a disease which is accompanied by Zn^{2+} deficiency, comprising administering an effective amount of phosphatase according to claim 3 to a subject in need thereof., said disease preferably comprising an inflammatory disease, more preferably selected from the group consisting of autoimmune diseases,
- 15 rheumatoid arthritis, asthma, chronic obstructive pulmonary disease, atherosclerosis, inflammatory bowel disease, sepsis, neurodermitis, and diseases depicted in Table 10.

Fig. 1

NP_001623 ALPP (Placental)

1 IIPVEEENPDFWNREAAEALGAACKLQPAQTAANKLIIFLGDGMGVSTVTAARILKGQKK 60
 61 DKLGPETPLAMDRFPYVALSKTYNVDRQVPDSAGATATAYLCGVKGNFQTIGLSAAARFNQ 120
 121 CNTTRGNEVISVMNRAKKAGKSVGVVTTTRVQHASPAGTYAHTVNRNWYSADVPASARQ 180
 181 EGCQDIATQLISNMDIDVILGGGRKYMFRMGTPDPEYPDDYSQGGTRLDGKNLVQEWLAK 240
 241 RQGARYVWNRTLMQASLDPSVTHLMGLFEPGDMKYEIHRDSTLDPSLMEMTEAALRLLS 300
 301 RNPRGFFLFVEGGRIDHGHESRAYRALTETIMFDDAIERAGQLTSEEDTLTLVTADHSH 360
 361 VFSFGGYPLRGSSIFGLAPGKARDRKAYTVLLYGNGPGYVLKDGARPDVTESESGSPEYR 420
 421 QOSAVPLDEETHAGEDVAVFARGPQAHLVHGVQEQTFFIAHVMAFAACLEPYTACDLAPPA 480
 481 GTTDAAHPGRSVVPALLPLLAGTLLLLLETATAP 513

AAI32679 ALPI (Intestinal)

1 VIPAEENPAFWNRQAAEALDAAKKLQPIQKVAKNLILFLGDGLGVPTVTATRILKGQKN 60
 61 GKLGPETPLAMDRFPYLALSKTYNVDRQVPDSAGATATAYLCGVKANFQTIGLSAAARFNQ 120
 121 CNTTRGNEVISVMNRAKQAGKSVGVVTTTRVQHASPAGTYAHTVNRNWYSADMPASARQ 180
 181 EGCQDIATQLISNMDIDVILGGGRKYMFRMGTPDPEYPADASQNGIRLDGKNLVQEWLAK 240
 241 HQGAWYVWNRTLMQASLDQSVTHLMGLFEPGDMKYEIHRDPTLDPSLMEMTEAALRLLS 300
 301 RNPRGFYLFVEGGRIDHGHHEGVAYQALTEAVMFDDAIERAGQLTSEEDTLTLVTADHSH 360
 361 VFSFGGYTLRGSSIFGLAPSKAQDSKAYTSILYGNGPGYVFNSGVRPDVNESESGSPDYQ 420
 421 QQAAPLSSETHAGEDVAVFARGPQAHLVHGVQEQSFAHVMAFAACLEPYTACDLAPPA 480
 481 CTTDAHPVAASLPALLPLLAGTLLLLGASAAP 509

P10696 GCAP (Germ-cell or Placental-like)

1 IIPVEEENPDFWNRQAAEALGAACKLQPAQTAANKLIIFLGDGMGVSTVTAARILKGQKK 60
 61 DKLGPETFLAMDRFPYVALSKTYSVDKHVPDSAGATATAYLCGVKGNFQTIGLSAAARFNQ 120
 121 CNTTRGNEVISVMNRAKKAGKSVGVVTTTRVQHASPAGAYAHTVNRNWYSADVPASARQ 180
 181 EGCQDIATQLISNMDIDVILGGGRKYMFRMGTPDPEYPDDYSQGGTRLDGKNLVQEWLAK 240
 241 HQGARYVWNRTELLQASLDPSVTHLMGLFEPGDMKYEIHRDSTLDPSLMEMTEAALLLS 300
 301 RNPRGFFLFVEGGRIDHGHESRAYRALTETIMFDDAIERAGQLTSEEDTLTLVTADHSH 360
 361 VFSFGGYPLRGSSIFGLAPGKARDRKAYTVLLYGNGPGYVLKDGARPDVTESESGSPEYR 420
 421 QOSAVPLDGETHAGEDVAVFARGPQAHLVHGVQEQTFFIAHVMAFAACLEPYTACDLAPRA 480
 481 GTTDAAHPGPSVVPALLPLLAGTLLLLGTATAP 513

AAI10910 (Tissue Non Specific)

1 LVPEKEKDPKYWRDQAEETLKYALELQKLNTNVAKNVIMFLGDGMGVSTVTAARILKGQL 60
 61 HHNPGEETRLEMDKFPFVALSKTYNTNAQVPDSAGTATAYLCGVKANEGTVGVSAATERS 120
 121 RCNTTQGNEVTSILRWAKDAGKSVGIVTTTRVNHATPSAAYAHSDRDWYSDNEMPPEAL 180
 181 SQGCKDIAYQLMHNIRDIDVIMGGGRKYMYPKNKTDVEYESDEKARGTRLDGLDLVDTWK 240
 241 SFKPRHKHSHFIWNRTELLTLDPHNVLDYLLGLFEPGDMQYELNRNNVTDPSLSEMVVVAI 300
 301 QILRKNPKGFFLLVEGGRIDHGHHEGKAKQALHEAVEMDRAIGQAGSLTSSSEDTLTVVTA 360
 361 DHSHVFTFGGYTPRGNSIFGLAPMLSDTDKKPFTAILYGNGPGYKVVGGERENVSMVDYA 420
 421 HNHYQAQSAVPLRHETHAGEDVAVFSKGPMALLHGVHEQNYVPHVMAYAACIGANLGHC 480
 481 APASSAGSLAAGPLLLALALYPLSVLF 507

Fig. 1 continued

Secretable ALPI with Crown-Domain of PLAP (chimera)

1 VIPAEENPAFWNRQAAEALDAAKKLQPIQKVAKNLILFLGDGLGVPTVTATRILKGQKN 60
61 GKLGPETPLAMDRFPYLALSKTYNVDRQVPDSAATATAYLCGVKANFQTIGLSAAARFNQ 120
121 CNTRGNEVISVMNRAKQAGKSVGVVTTTRVQHASPAGTYAHTVNRNWYSADMPASARQ 180
181 EGCQDIATQLISNMDIDVILGGGRKYMFPMPGTPDPEYPADASQNGIRLDGKNLVQEWLAK 240
241 HQGAWYVWNRTELMQASLDQSVTHLMGLFEPGDTKYEIHRDPTLDPSLMEMTEAALRLLS 300
301 RNPRGFYLFVEGGRIDHGHHEGVAYQALTEAVMFDDAIERAGQLTSEEDTLTLVTADHSH 360
361 VFSFGGYPLRGSSIFGLAPGKARDRKAYTVLLYGNGPGYVLKDGARPDVTESESGSPEYR 420
421 QSAVPLDEETHGGEDVAVFARGPQAHLVHGVQEQS FVAHVMAFAACLEPYTACDLAPPA 480
481 CTTD 484

Secretable ALPP with Crown-Domain of ALPI (chimera)

1 IIPVEENPDEFWNREAAEALGAACKLQPAQTAANKLIIFLGDGMGVSTVTAARILKGQKK 60
61 DKLGPETPLAMDRFPYVALSKTYNVDRQVPDSAATATAYLCGVKGNFQTIGLSAAARFNQ 120
121 CNTRGNEVISVMNRAKQAGKSVGVVTTTRVQHASPAGTYAHTVNRNWYSADVPASARQ 180
181 EGCQDIATQLISNMDIDVILGGGRKYMFRMGTDPDPEYPDDYSQGGTRLDGKNLVQEWLAK 240
241 RQGARYVWNRTELMQASLDPSVTHLMGLFEPGDMKYEIHRDSTLDPSLMEMTEAALRLLS 300
301 RNPRGFFLFVEGGRIDHGHHESTRAYRALTETIMFDDAIERAGQLTSEEDTLTLVTADHSH 360
361 VFSFGGYTLRGSSIFGLAPSKAQDSKAYTSILYGNGPGYVFNSGVRPDVNESESGSPDYQ 420
421 QAAVPLSSETHAGEDVAVFARGPQAHLVHGVQEQT FIAHVMAFAACLEPYTACDLAPPA 480
481 GTTD 484

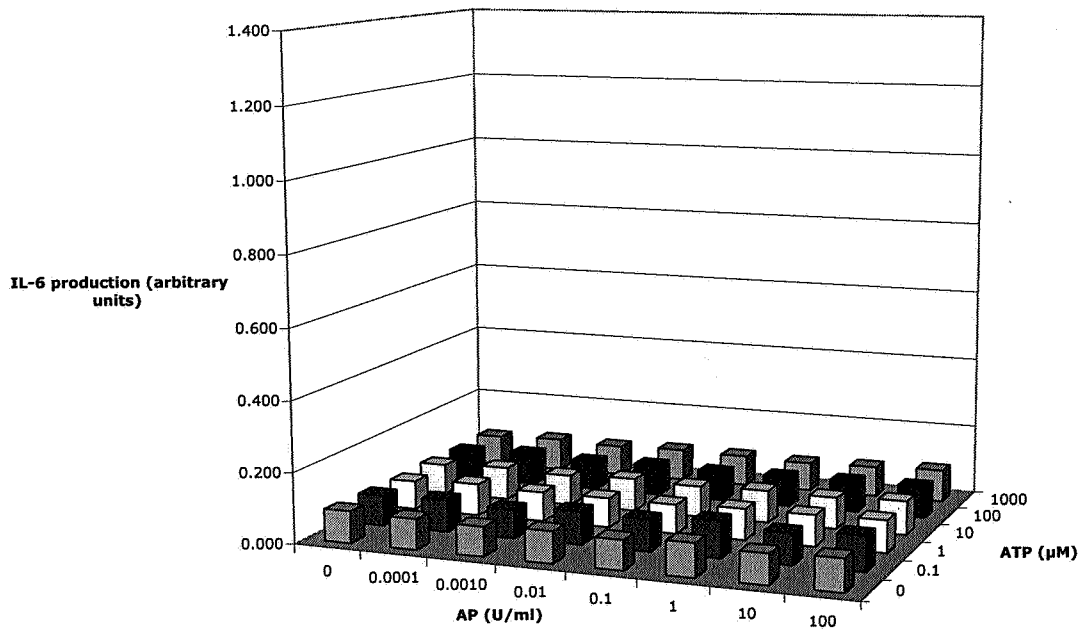


Fig. 2

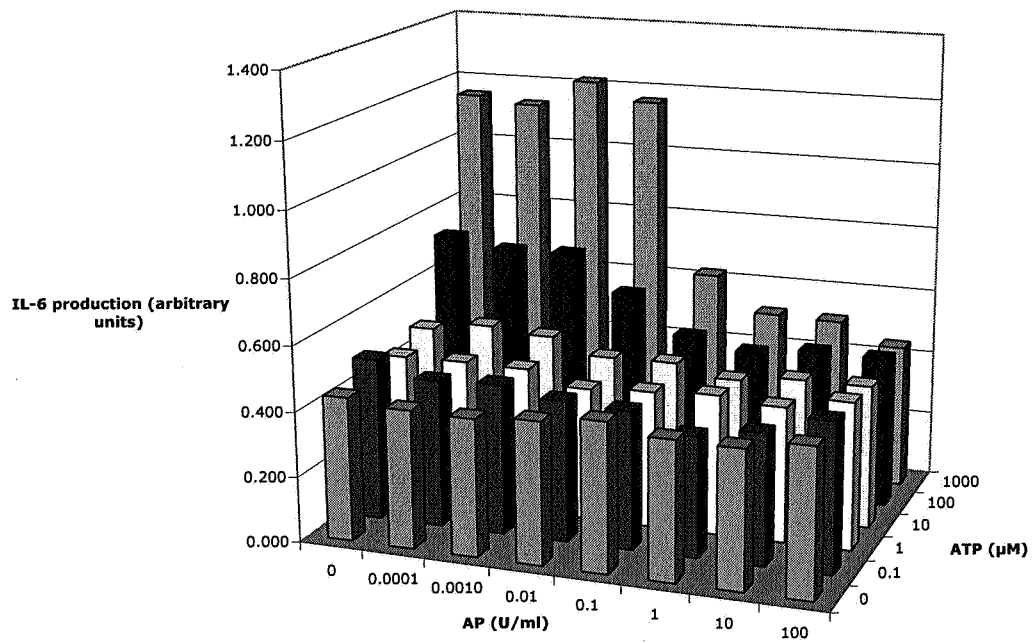


Fig. 3

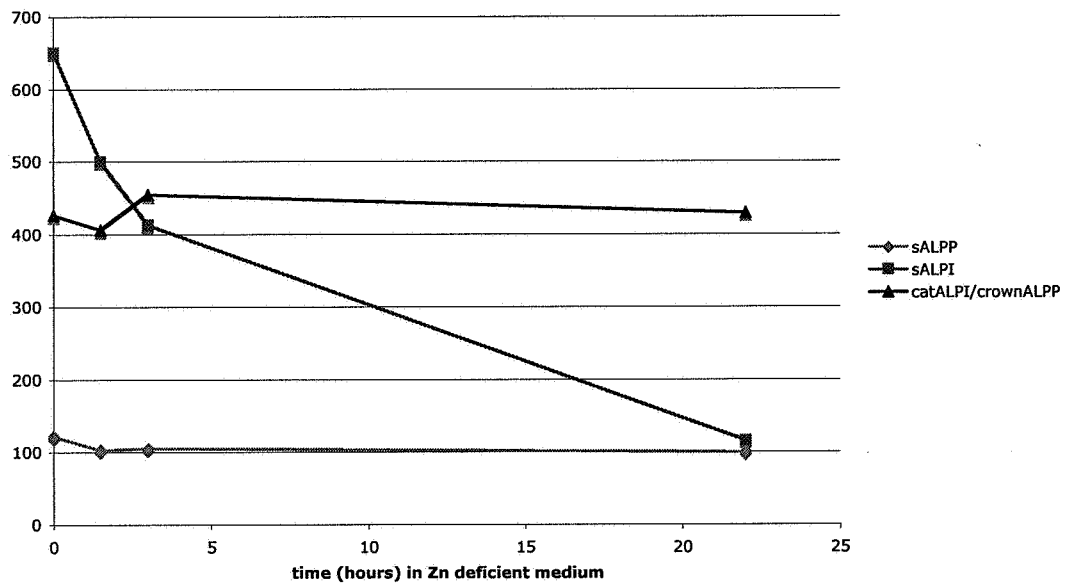
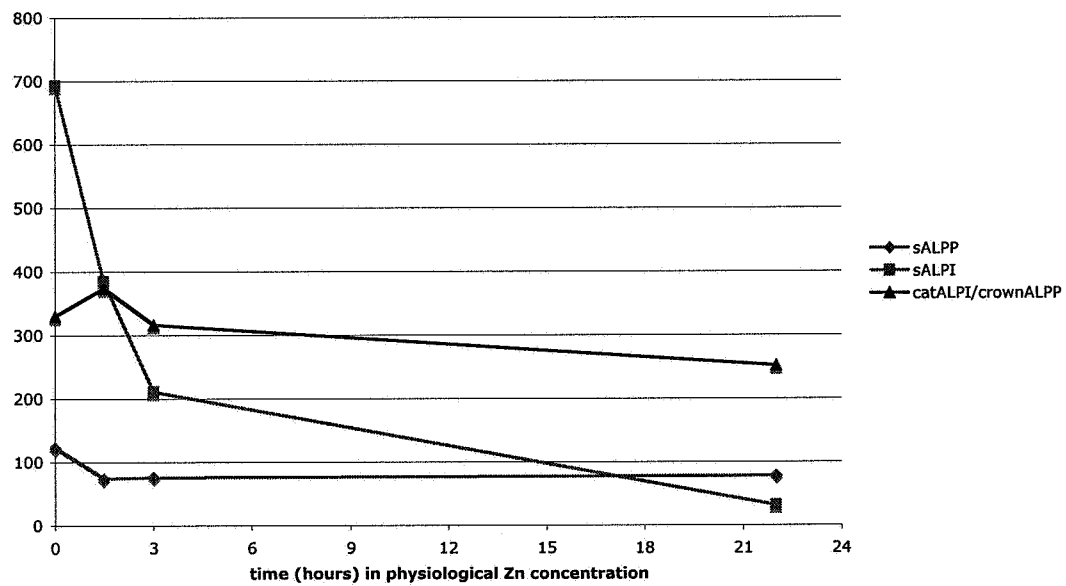
Fig. 4 - Specific activity of catALPI/crownALPP is Zn independent**Fig. 5 - Specific activity of ALPI but not catALPI/crownALPP decreases in time**

Fig 6 - liver slice assay

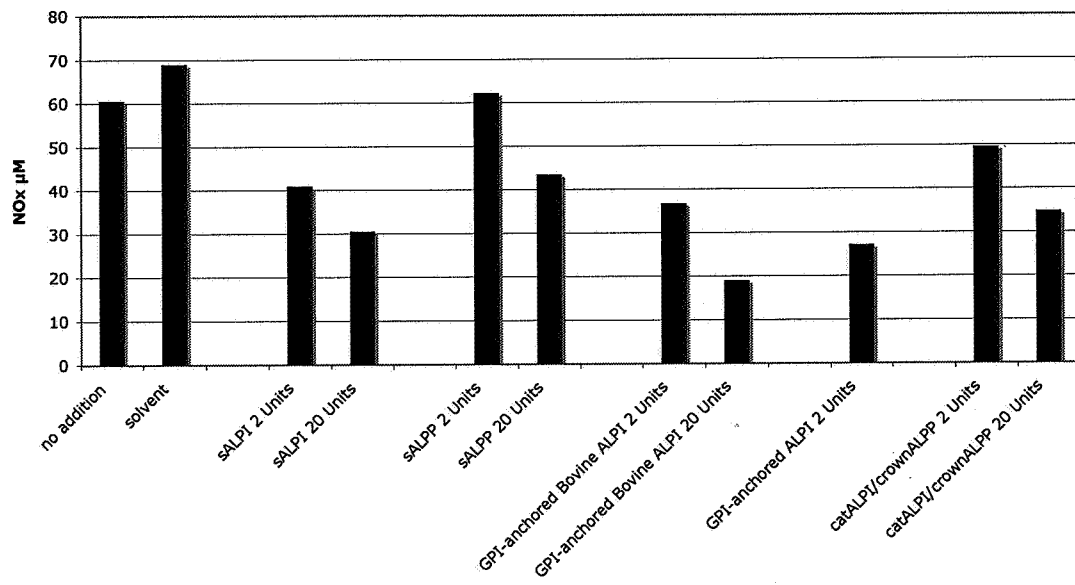


Figure 7

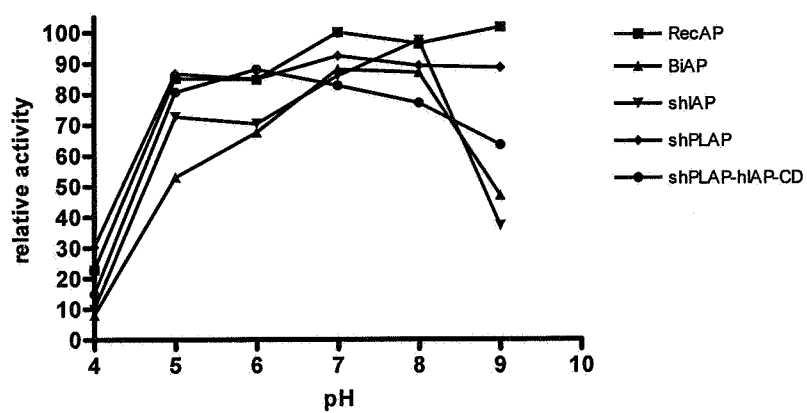


Figure 8

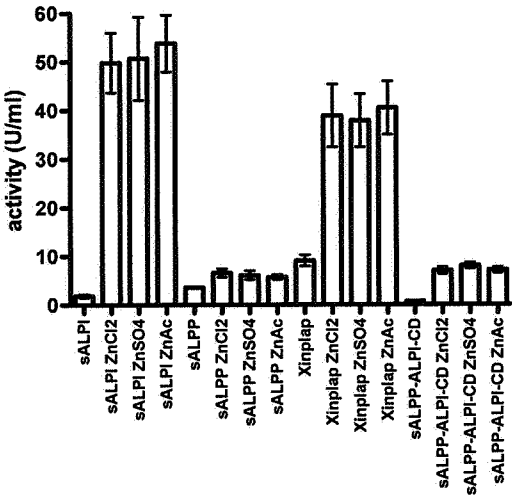
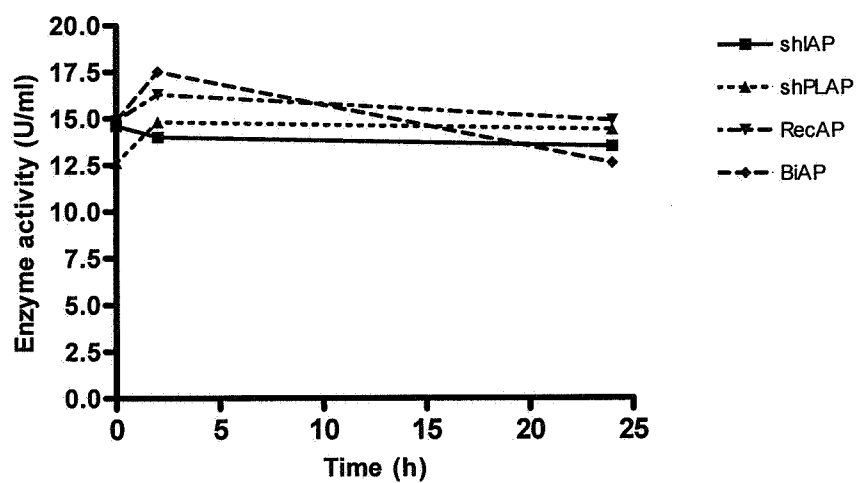
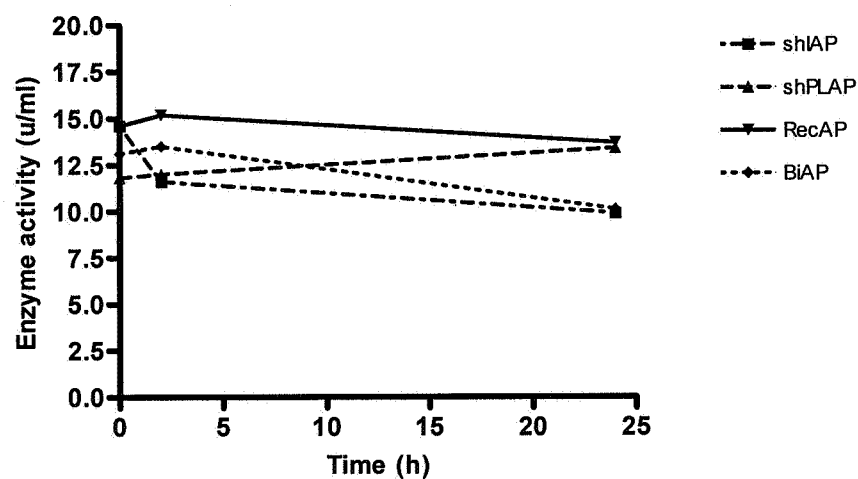
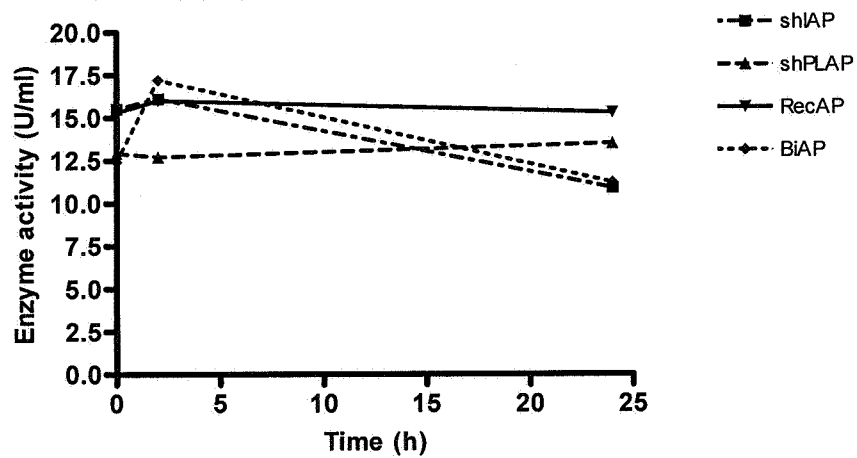


Figure 9

A RT with Zn^{2+} B RT without Zn^{2+} C 37°C with Zn^{2+} 

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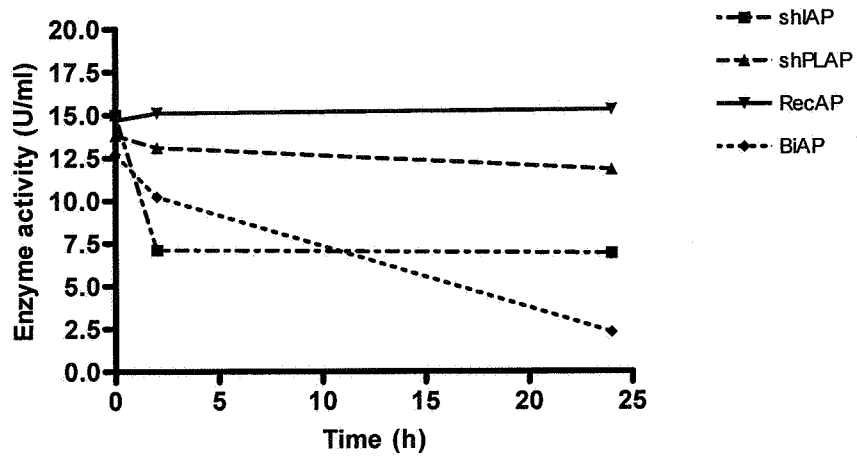
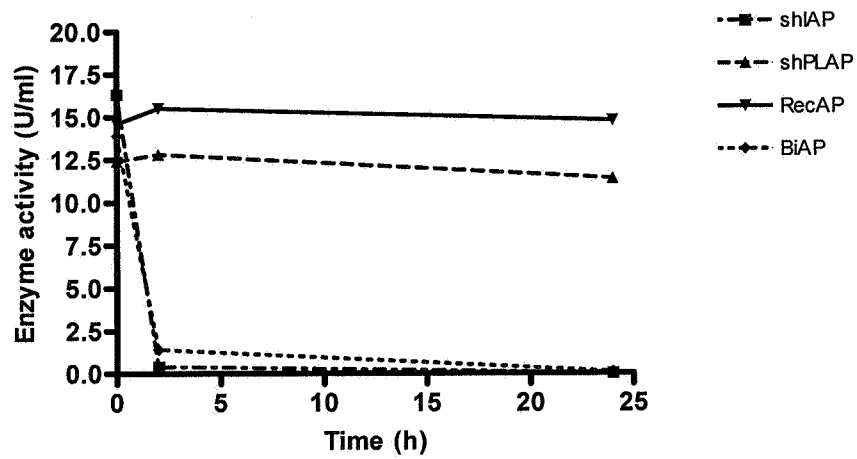
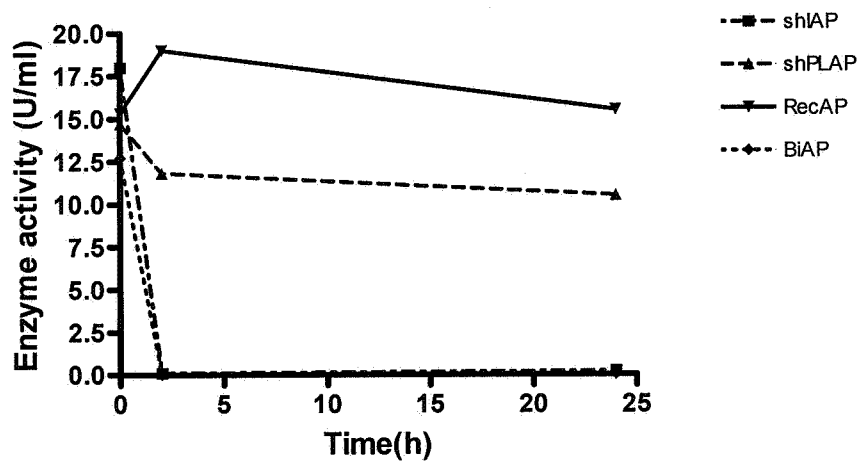
D 37°C without Zn²⁺E 56°C with Zn²⁺F 56° without Zn²⁺

Figure 10

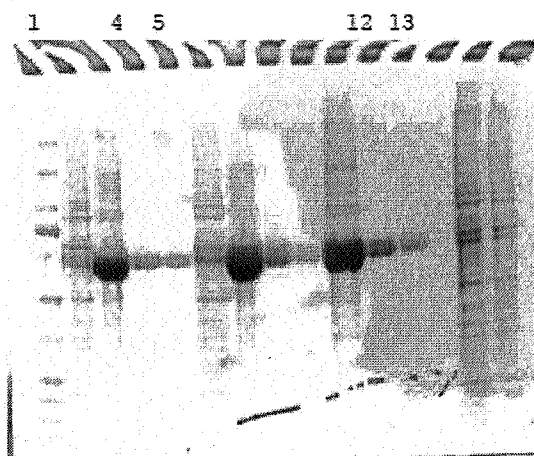


Figure 11

