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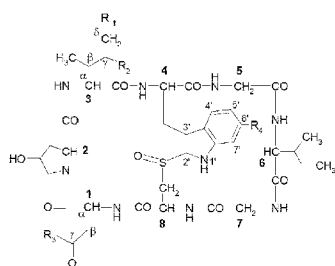
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(54) Title: SYNTHESIS OF (S)-6-HYDROXYTRYPTOPHAN AND DERIVATIVES THEREOF



	R ₁	R ₂	R ₃	R ₄
α-amanitin	OH	OH	NH ₂	OH
β-amanitin	OH	OH	OH	OH
γ-amanitin	H	OH	NH ₂	OH
ε-amanitin	H	OH	OH	OH
amanin	OH	OH	OH	H
amaninamide	OH	OH	NH ₂	H
amanullin	H	H	NH ₂	OH
amanullinic acid	H	H	OH	OH
γ-amanin	H	OH	OH	H
γ-amaninamide	H	OH	NH ₂	H

Fig. 1

(57) Abstract: The present invention relates to novel methods and compounds for synthesizing amanitin derivatives. The invention in particular relates to methods for synthesizing (S)-6-hydroxy-tryptophan derivatives which can be used as building blocks for synthesizing amanitin derivatives or amatoxin drug conjugates. The invention further relates to intermediate compounds of said synthesis pathways for use in amanitin derivative and amatoxin drug conjugate synthesis, and to the use of particular catalysts suited for mediating said synthesis pathways.

Synthesis of (S)-6-Hydroxytryptophan and Derivatives Thereof

Field of the invention

The present invention relates to novel methods and compounds for synthesizing amanitin derivatives. The invention in particular relates to methods for synthesizing (S)-6-hydroxytryptophan derivatives which can be used as building blocks for synthesizing amanitin derivatives or amatoxin drug conjugates. The invention further relates to intermediate compounds of said synthesis pathways for use in amanitin derivative and amatoxin drug conjugate synthesis, and to the use of particular catalysts suited for mediating said synthesis pathways.

Background

Amatoxins are cyclic peptides composed of 8 amino acids that are found in *Amanita phalloides* mushrooms (see Fig. 1). Amatoxins specifically inhibit the DNA-dependent RNA polymerase II of mammalian cells, and thereby also the transcription and protein biosynthesis of the affected cells. Inhibition of transcription in a cell causes stop of growth and proliferation. Though not covalently bound, the complex between amanitin and RNA-polymerase II is very tight ($K_D = 3 \text{ nM}$). Dissociation of amanitin from the enzyme is a very slow process, thus making recovery of an affected cell unlikely. When the inhibition of transcription lasts sufficiently long, the cell will undergo programmed cell death (apoptosis).

Amatoxins can be isolated from collected *Amanita phalloides* mushroom fruit bodies, or from pure cultures (Zhang P, *et al.*, FEMS Microbiol Lett. 2005 Nov 15;252(2):223-8. Epub 2005 Sep 15). However, the amounts of amatoxins that can be obtained are rather low (in the range of about 0.3 - 3 mg/g dry matter from natural fruit bodies, and about 10% thereof from pure culture) and the flexibility for further modifying the naturally occurring amatoxin variants is limited. Alternatively, amatoxins can be obtained from fermentation using a basidiomycete (Muraoka S, and Shinozawa T., J Biosci Bioeng. 2000;89(1):73-6) or *A. fissa* (Guo XW, *et al.*, 2006 Jun;46(3):373-8). Again, yields are low, and flexibility for further modifying the naturally occurring amatoxin variants is limited as well. Finally, amatoxins have been prepared by partial or total synthesis (e.g. Zanotti G, Mähringer C, and Wieland T., Int J Pept Protein Res. 1987 Oct;30(4):450-9; Zanotti G, Wieland T, Benedetti E, Di Blasio S, Pavone V, and Pedone C., Int J Pept Protein Res. 1989 Sep;34(3):222-8). Alternatively, the use of fully-synthetic routes to amatoxins may offer the supply of larger quantities of amatoxins required for therapeutic uses, and may offer the construction of a variety of novel amatoxin variants by using appropriate starting materials as building blocks.

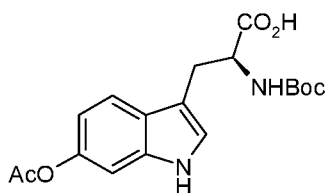
Naturally occurring amanitins such as α -amanitin, β -amanitin or γ -amanitin are comprising a phenolic hydroxy (-OH) function at the 6'-position of tryptophan, that represents amino acid 4 in the cyclic amanitin octapeptide, which allows for coupling of a linker to the amatoxin (see Fig. 1). Target-binding macromolecules, such as antibodies or aptamers, can then be coupled via said linker in order to generate conjugates, e.g., antibody-drug conjugates. The use of amatoxins as cytotoxic moieties for tumor therapy has already been explored in 1981 by coupling an anti-Thy 1.2 antibody to α -amanitin using a linker attached to the indole ring of tryptophan (Trp, amino acid 4; see Fig. 1) via diazotation (Davis & Preston, 1981, Science 213: 1385-1388).

With the currently explored fully synthetic amanitin compounds, incorporation of a functionalized tryptophan for coupling of linker elements has only been achieved very recently. Prior to that, therefore, linkers were coupled to fully synthetic amanitins predominantly at positions amino acid 1 (aspartic acid) or at the indol-nitrogen (N1) of amino acid 4 (tryptophan). As biologic activity profiles of amanitins coupled via different anchorage positions of the amanitin framework have been shown to be very different, it is of great interest to be able to provide synthetic tryptophan hydroxylated at its 6'-position, *i.e.*

corresponding to the native α -amanitin, β -amanitin or γ -amanitin, as a building block that can be incorporated in the course of amanitin synthesis.

The synthetic introduction of tryptophan into the amanitin framework can be performed by way of the Savige-Fontana reaction (Savige & Fontana, 1980, Int J Pept Protein Res. 15(3): 285-97). According to this reaction, tryptophan is converted into a mixture of Boc-protected cis-2-carboxy-3a-hydroxy-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-b]indol and trans-2-carboxy-3a-hydroxy-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-b]indol and then incorporated into the amino acid sequence of the linear amanitin precursor.

For the fully synthetic production of amanitin, (S)-6-hydroxytryptophan synthesis and synthesis of respective building blocks are of essential importance. In particular, one of these building blocks of essential importance is (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan (HDP 30.2550).



HDP 30.2550

So far, a satisfying and efficient synthesis pathway for (S)-6-hydroxytryptophan and its building blocks has not been described in the prior art. In particular, a synthesis pathway yielding the necessary purity of enantiomers (L or S) of this amino acid derivative has not been available. Different basic options of synthesis could principally be considered. A first option could be the crystallization of the racemic form with chiral auxiliary bases or acids. However, only a maximal yield of 50% could be achieved with this method. A second option is an enzymatic production; employing this method, however, is time-consuming, results are uncertain and reproducibility is weak.

It was hence one object of the present invention to provide an efficient, simple and reproducible method for synthesizing (S)-6-hydroxytryptophan, its derivatives and its building blocks. Preferably, it was one object of the present invention to provide an efficient

method for synthesizing (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan (HDP 30.2550) with a sufficiently high enantiomeric purity.

As one further object of the present invention, the use of said (S)-6-hydroxytryptophan and its derivatives, preferably (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan, as building blocks for the fully synthetic production of amatoxins should be provided.

These and further objects are met with methods and means according to the independent claims of the present invention. The dependent claims are related to specific embodiments.

Summary of the Invention

The present invention provides methods for synthesis of (S)-6-hydroxytryptophan, (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan, and their derivatives and building blocks for their synthesis. The present invention further provides compounds and building blocks for use in synthesizing amanitin or amanitin derivatives or amatoxin-drug conjugates.

The invention and general advantages of its features will be discussed in detail below.

Description of the Figures

Fig. 1 shows the structural formulae of different amatoxins. The numbers in bold type (1 to 8) designate the standard numbering of the eight amino acids forming the amatoxin. The standard designations of the atoms in amino acids 1, 3 and 4 are also shown (Greek letters α to γ , Greek letters α to δ , and numbers from 1' to 7', respectively).

Fig. 2 illustrates the synthesis of compound HDP 30.2822.

Fig. 3 illustrates the synthesis of compound HDP 30.2758.

Fig. 4 illustrates the synthesis of compounds HDP 30.2550 and HDP 30.2555.

Fig. 5 shows structural compositions of catalysts used for the conversion of HDP 30.2824 to HDP 30.2826.

Fig. 6 shows ¹H NMR spectroscopy results from compound HDP 30.2826 (400MHz, CDCl₃, δ=ppm).

Fig. 7 shows ¹³C NMR spectroscopy results from compound HDP 30.2826 (100MHz, CDCl₃, δ=ppm).

Detailed Description of the Invention

Before the invention is described in detail, it is to be understood that this invention is not limited to the particular component parts of the means described or process steps of the methods described as such means and methods may vary. It is also to be understood that the terminology used herein is for purposes of describing particular embodiments only, and is not intended to be limiting. It must be noted that, as used in the specification and the appended claims, the singular forms "a", "an", and "the" include singular and/or plural referents unless the context clearly dictates otherwise. It is moreover to be understood that, in case parameter ranges are given which are delimited by numeric values, the ranges are deemed to include these limitation values.

It is further to be understood that embodiments disclosed herein are not meant to be understood as individual embodiments which would not relate to one another. Features discussed with one embodiment are meant to be disclosed also in connection with other embodiments shown herein. If, in one case, a specific feature is not disclosed with one embodiment, but with another, the skilled person would understand that this does not necessarily mean that said feature is not meant to be disclosed with said other embodiment. The skilled person would understand that it is the gist of this application to disclose said feature also for the other embodiment, but that just for purposes of clarity and to keep the specification in a manageable volume this has not been done.

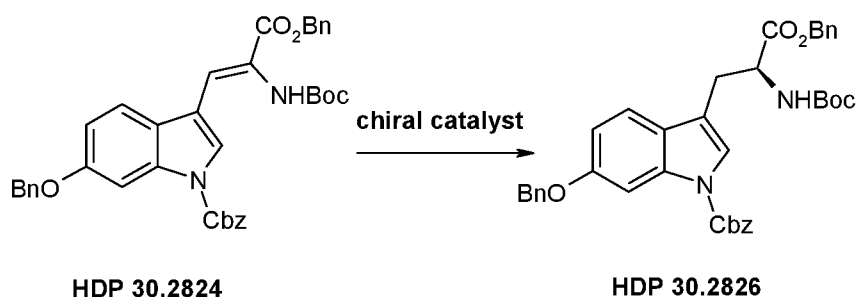
Furthermore, the content of the prior art documents referred to herein is incorporated by reference. This refers, particularly, for prior art documents that disclose standard or routine methods. In that case, the incorporation by reference has mainly the purpose to provide sufficient enabling disclosure, and avoid lengthy repetitions.

A large-scale, industrially feasible production of (S)-6-hydroxytryptophan as comprised in naturally occurring α -, β - and γ -amanitin, and/or chemically protected forms thereof, requires an efficient, simple and reproducible synthesis method. For the first time, the inventors were able to provide such a simple and efficient production method by employing an enantiomer-selective hydrogenation of olefinic amino acid precursors. Using specific chiral catalysts, they surprisingly found that such type of reaction can yield very high enantiomeric purity.

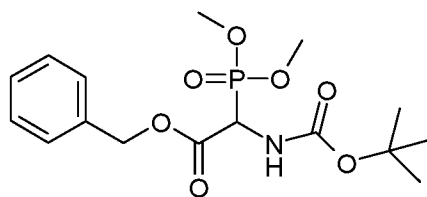
The simple synthetic pathway disclosed yields (S)-6-hydroxytryptophan that is fully identical to the respective native structure. The capability of using said compound and protected forms thereof for industrial large-scale production of amanitins will considerably reduce costs and increase efficiency of, for example, manufacturing synthetic α -, β - or γ -amanitin as well as of amatoxin-based antibody-drug conjugates for therapeutic applications.

According to a first aspect, the present invention relates to a method for synthesis of (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan (HDP 30.2550) or (S)-6-hydroxytryptophan, wherein said method comprises at least one step of enantiomer-selective hydrogenation of an olefinic precursor compound by use of at least one chiral catalyst.

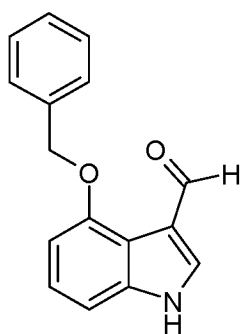
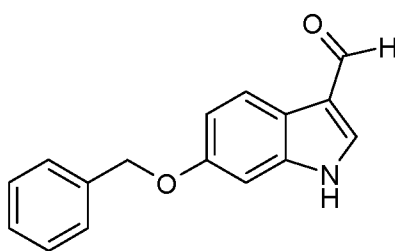
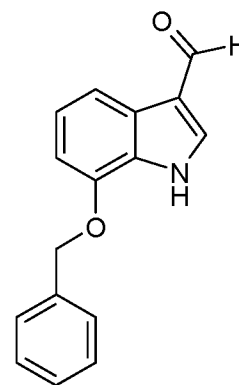
In one embodiment of the method claimed, said olefinic precursor used for asymmetric hydrogenation is an olefinic unsaturated amino acid precursor. Preferably it is compound HDP 30.2824.



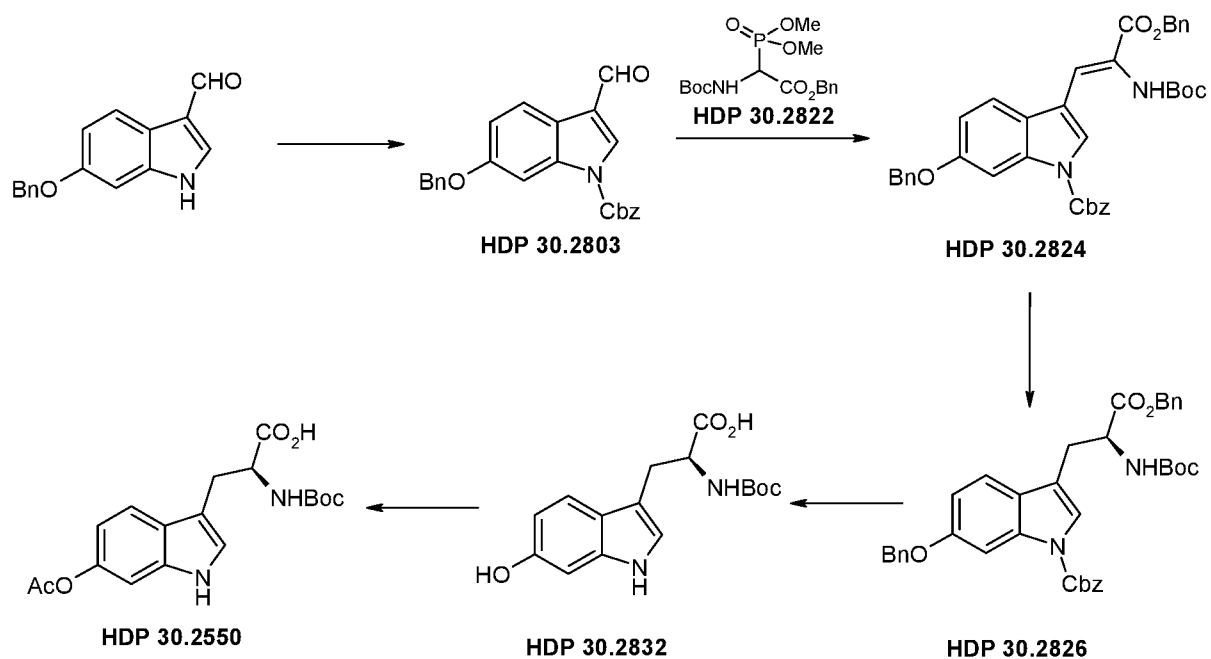
In one embodiment of the method claimed, said olefinic precursor is synthesized by use of the following compound:

**HDP 30.2822**

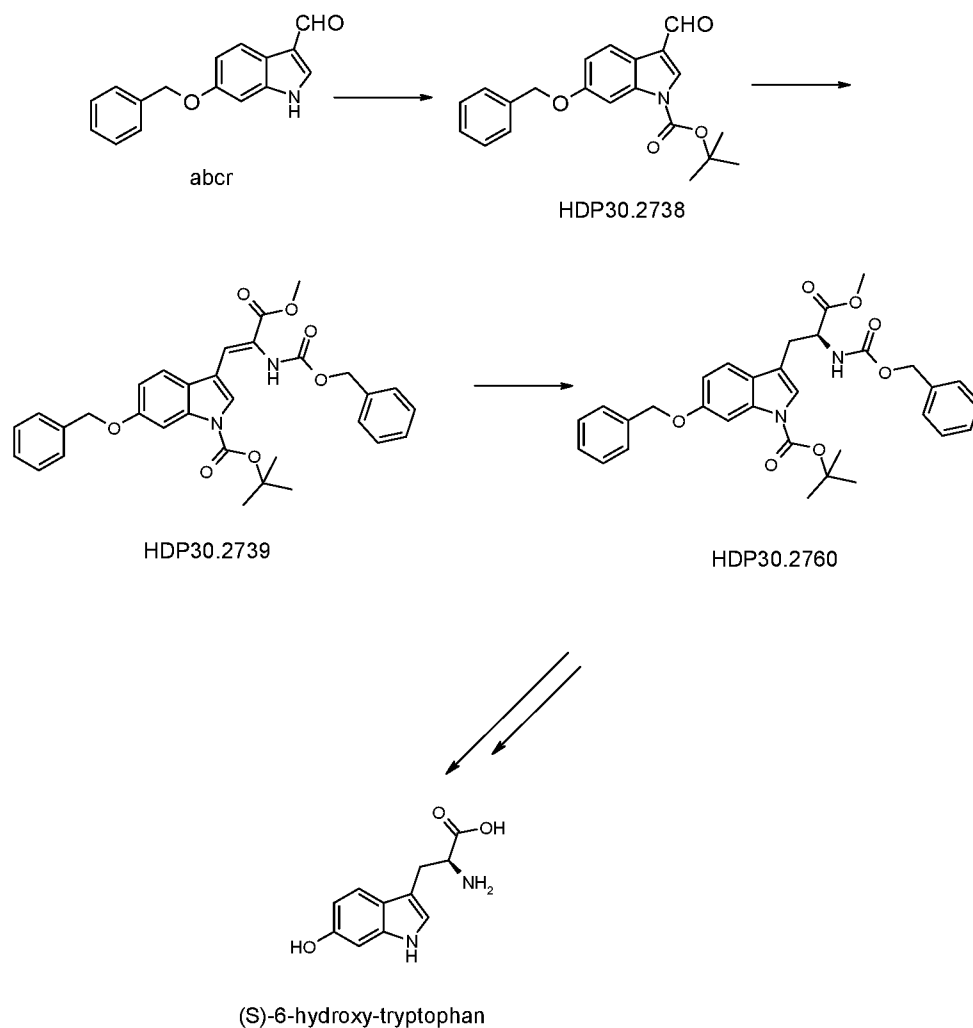
Said olefinic precursor can further be synthesized by use of any of compounds A, B, or C.

**A****B****C**

In a further embodiment of the method claimed, said method comprises at least the following steps:



In a still further embodiment of the method claimed, said method comprises at least the following steps:



In the context of the present invention the term “amatoxin” includes all cyclic peptides composed of 8 amino acids as isolated from the genus *Amanita* and described in Wieland, T. and Faulstich H. (Wieland T, Faulstich H., *CRC Crit Rev Biochem.* 5 (1978) 185-260), further all chemical derivatives thereof; further all semisynthetic analogs thereof; further all synthetic analogs thereof built from building blocks according to the master structure of the natural compounds (cyclic, 8 amino acids), further all synthetic or semisynthetic analogs containing non-hydroxylated amino acids instead of the hydroxylated amino acids, further all synthetic or semisynthetic analogs, in which the thioether sulfoxide moiety is replaced by a sulfide, sulfone, thioether, or by atoms different from sulfur, e.g. a carbon atom as in a carbanalog of amanitin. Functionally, amatoxins are defined as peptides or depsipeptides that inhibit mammalian RNA polymerase II. Preferred amatoxins are those with a functional

group (e.g. a carboxylic group, an amino group, a hydroxy group, a thiol or a thiol-capturing group) that can be reacted with linker molecules or target-binding moieties as defined below.

In the context of the present invention, the term “amanitins” particularly refers to bicyclic structure that are based on an aspartic acid or asparagine residue in position 1, a proline residue, particularly a hydroxyproline residue in position 2, an isoleucine, hydroxyisoleucine or dihydroxyisoleucine in position 3, a tryptophan or hydroxytryptophan residue in position 4, glycine residues in positions 5 and 7, an isoleucine residue in position 6, and a cysteine residue in position 8, particularly a derivative of cysteine that is oxidized to a sulfoxide or sulfone derivative (for the numbering and representative examples of amanitins, see Figure 1), and furthermore includes all chemical derivatives thereof; further all semisynthetic analogues thereof; further all synthetic analogues thereof built from building blocks according to the master structure of the natural compounds (cyclic, 8 amino acids), further all synthetic or semisynthetic analogues containing non-hydroxylated amino acids instead of the hydroxylated amino acids, further all synthetic or semisynthetic analogues, in each case wherein any such derivative or analogue is functionally active by inhibiting mammalian RNA polymerase II.

The term “target-binding moiety”, as used herein, refers to any molecule or part of a molecule that can specifically bind to a target molecule or target epitope. Preferred target-binding moieties in the context of the present application are (i) antibodies or antigen-binding fragments thereof; (ii) antibody-like proteins; and (iii) nucleic acid aptamers. “Target-binding moieties” suitable for use in the present invention typically have a molecular mass of 40 000 Da (40 kDa) or more.

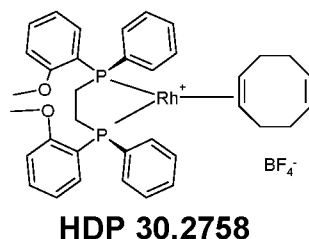
A "linker" in the context of the present application refers to a molecule that increases the distance between two components, e.g. to alleviate steric interference between the target binding moiety and the amatoxin, which may otherwise decrease the ability of the amatoxin to interact with RNA polymerase II. The linker may serve another purpose as it may facilitate the release of the amatoxin specifically in the cell being targeted by the target binding moiety. It is preferred that the linker and preferably the bond between the linker and the amatoxin on one side and the bond between the linker and the antibody on the other side is stable under the physiological conditions outside the cell, e.g. the blood, while it can be cleaved inside the cell, in particular inside the target cell, e.g. cancer cell or immune cell. To

provide this selective stability the linker may comprise functionalities that are preferably pH-sensitive or protease sensitive. Alternatively, the bond linking the linker to the target binding moiety may provide the selective stability. Preferably a linker has a length of at least 1, preferably of 1-30 atoms length (e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30 atoms) wherein one side of the linker has been reacted with the amatoxin and, the other side with a target-binding moiety. In the context of the present invention, a linker preferably is a C₁₋₃₀-alkyl, C₁₋₃₀-heteroalkyl, C₂₋₃₀-alkenyl, C₂₋₃₀-heteroalkenyl, C₂₋₃₀-alkynyl, C₂₋₃₀-heteroalkynyl, cycloalkyl, heterocycloalkyl, aryl, heteroaryl, aralkyl, or a heteroaralkyl group, optionally substituted. The linker may contain one or more structural elements such as amide, ester, ether, thioether, disulfide, hydrocarbon moieties and the like. The linker may also contain combinations of two or more of these structural elements. Each one of these structural elements may be present in the linker more than once, e.g. twice, three times, four times, five times, or six times. In some embodiments the linker may comprise a disulfide bond. It is understood that the linker has to be attached either in a single step or in two or more subsequent steps to the amatoxin and the target binding moiety. To that end the linker to be will carry two groups, preferably at a proximal and distal end, which can (i) form a covalent bond to a group, preferably an activated group on an amatoxin or the target binding-peptide or (ii) which is or can be activated to form a covalent bond with a group on an amatoxin. Accordingly, if the linker is present, it is preferred that chemical groups are at the distal and proximal end of the linker, which are the result of such a coupling reaction, e.g. an ester, an ether, a urethane, a peptide bond etc. The presence of a "linker" is optional, i.e. the toxin may be directly linked to a residue of the target-binding moiety in some embodiments of the target-binding moiety toxin conjugate.

According to a second aspect, the present invention relates to (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan (HDP 30.2550), (S)-6-hydroxy-tryptophan or any of the precursor compounds according to the present invention for use in synthesis of amanitin or amanitin derivatives or amatoxin-drug conjugates.

In one embodiment, the present invention relates to a dehydro-amino acid compound selected from the group consisting of compounds I, II, III, IV and V,

[(R,R)-DIPAMP] rhodium tetrafluoroborate, HDP 30.2758, yielded the highest enantiomeric purity which was > 98%).



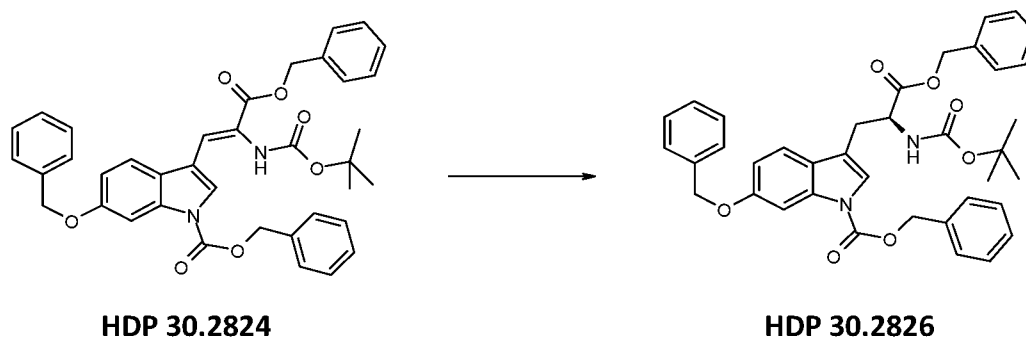
It was found that only the catalyst HDP 30.2758 yielded a very high purity of more than 98% of (S)-enantiomers. All other catalysts except (R,R)-Et-DUPHOS (BF₄⁻) yielded considerably lower (S)-enantiomer purities of 50-70%. In addition, the overall absolute compound turnover and yield rates were much poorer than with HDP 30.2758.

The catalysts tested for asymmetric hydrogenation and respective (S)-enantiomer purity levels are compiled in Table 1.

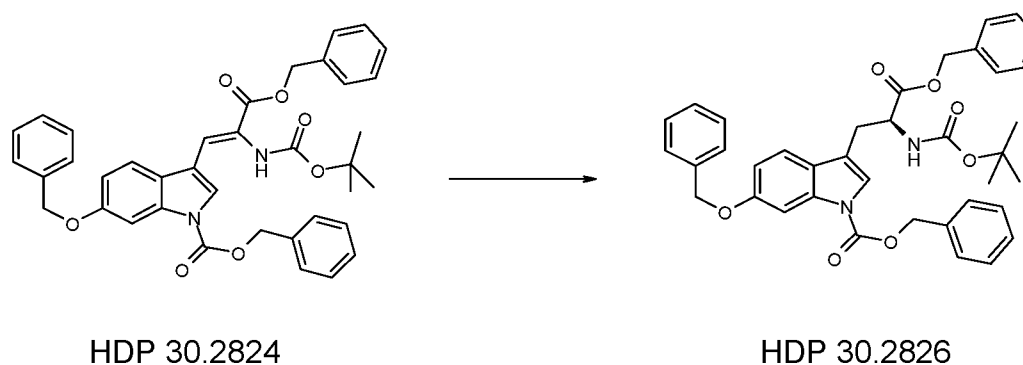
Table 1: Comparative Assessment of various catalysts used for enantiomer-selective hydrogenation

Catalyst #	Identity	Purity Level
1	HDP 30.2758	>98% chiral purity
2	(R,R)-Et-DUPHOS (CF ₃ SO ₃ ⁻)	no conversion
3	(R,R)-Et-DUPHOS (BF ₄ ⁻)	90-95% chiral purity
4	(R,R)-DuPhos-Ferrocene (BF ₄ ⁻)	65% chiral purity
5	(R,R)-DuPhos-Ferrocene-Et ₂ (BF ₄ ⁻)	55% chiral purity
6	(R,R)-DuPhos-Alkyl (CF ₃ SO ₃ ⁻)	73% chiral purity
7	(R,R)-Phenyl-DuPhos- Alkyl (BF ₄ ⁻)	<60% chiral purity

Thus according to a third aspect, the present invention relates to the use of a compound selected from the group consisting of compound HDP 30.2758, (R,R)-Et-DUPHOS (BF₄⁻), (R,R)-DuPhos-Ferrocene (BF₄⁻), (R,R)-DuPhos-Ferrocene-Et₂ (BF₄⁻), (R,R)-DuPhos-Alkyl (CF₃SO₃⁻), and (R,R)-Phenyl-DuPhos- Alkyl (BF₄⁻) as catalyst for hydrogenation in the following reaction:



According to this aspect, the present invention preferably relates to the use of the chiral catalysts cyclooctadiene-1,5-[(R,R)-DIPAMP] rhodium tetrafluoro-borate (HDP 30.2758) or (R,R)-Et-DUPHOS (BF₄-), most preferably of the chiral catalyst cyclooctadiene-1,5-[(R,R)-DIPAMP] rhodium tetrafluoro-borate (HDP 30.2758), as catalyst for hydrogenation in the following reaction:



The incorporation of tryptophan derivatives as building blocks into amanitin precursors has been described before in PCT/EP2018/071268, the content of which is incorporated herein by reference. Such as HDP 30.2115 can be synthesized by incorporation of unsubstituted Hpi building block into an amanitin peptidic precursor molecule in order to yield synthetic amanitin, the 6-hydroxy-substituted building block HDP 30.2555 can be used for synthesis of 6-hydroxy-substituted amanitins according to the present invention.

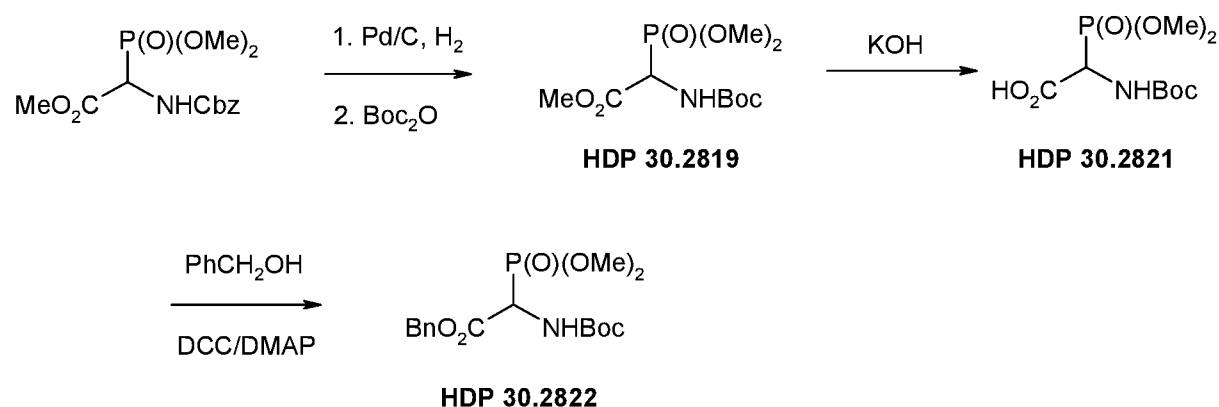
Examples

While the invention has been illustrated and described in detail in the drawings and foregoing description, such illustration and description are to be considered illustrative or exemplary

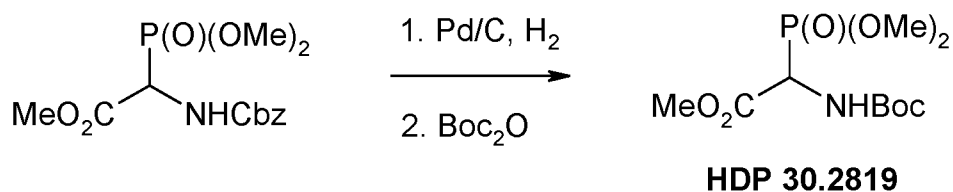
and not restrictive; the invention is not limited to the disclosed embodiments. Other variations to the disclosed embodiments can be understood and effected by those skilled in the art in practicing the claimed invention, from a study of the drawings, the disclosure, and the appended claims. In the claims, the word “comprising” does not exclude other elements or steps, and the indefinite article “a” or “an” does not exclude a plurality. The mere fact that certain measures are recited in mutually different dependent claims does not indicate that a combination of these measures cannot be used to advantage. Any reference signs in the claims should not be construed as limiting the scope.

Example 1: Synthesis of the phosphonium precursor (building block) HDP 30.2822

The synthesis of the phosphonium precursor (building block) HDP 30.2822 was performed as described (CHEMISTRY A European Journal, 2018, Vol. 24, Issue 7, pp 1544-1553):



Example 1.1: Preparation of (R,S)-Boc- α -phosphonoglycine trimethyl ester HDP 30.2819

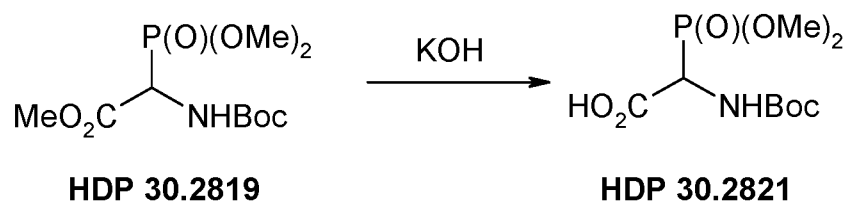


5.0g (15.1 mmol) (R,S)-N-Cbz-phosphonoglycine trimethyl ester (CAS: 88568-95-0) was hydrogenated with 1.4g 10% Pd.C in 100ml methanol at 1 atm until the reaction was

complete by TLC (chloroform / methanol 15:1). The reaction was complete in 3 hours. The catalyst was filtered off over a plug of Celite® (diatomaceous earth) and the methanolic solution of the free amine was concentrated *in vacuo* to a colorless oil (2.9g). The crude oil was used for the next step without purification.

2.9g crude hydrogenation product was dissolved in 20ml dichloromethane and treated with 3.23ml (15.1mmol) di-tert-butyl dicarbonate (Boc₂O). After 17 hours stirring at ambient temperature under argon, the reaction mixture was concentrated to dryness. The remaining colorless oil crystallizes to a white solid (4.3g). The crude HDP 30.2819 was used for the next step without purification.

Example 1.2: Preparation of (R,S)-N-Boc- α -phosphonoglycine trimethyl ester HDP 30.2821



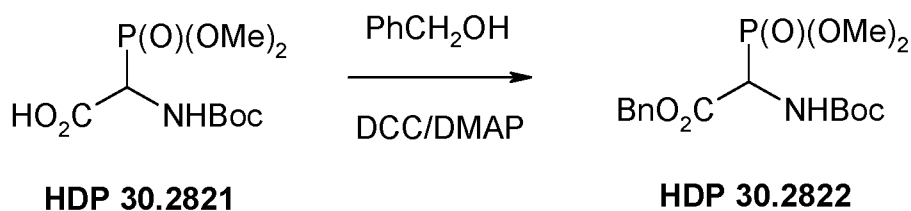
4.3g (assumed 14.3mmol) crude HDP 30.2819 was dissolved in 10ml 1,4-dioxane and rapidly treated under argon and room temperature with 14.5ml 1 N KOH. After 85 minutes, the reaction mixture was diluted with 36ml water and extracted with 35ml ethyl acetate. The ethyl acetate extract was discarded and the aqueous solution acidified to pH 3 by dropwise addition of 1 N HCl. The reaction mixture was extracted with 60ml ethyl acetate (2x), and dried over MgSO₄. The resulting white solid, 2.1g of HDP 30.2821, was dried *in vacuo* and used directly without purification for the next reaction step.

Mp: 148-150°C (Lit. JACS 111, 6244, 1989 mp: 154-155°C)

MS (ESI⁺) found: 282.00 [M-H]⁺; calc.: 283.08 (C₉H₁₈NO₇P)

MS (ESI⁻) found: 238.17 [M-CO₂]⁻

Example 1.3: Preparation of (R,S)-N-Boc- α -dimethylphosphono)-glycine benzyl ester HDP 30.2822



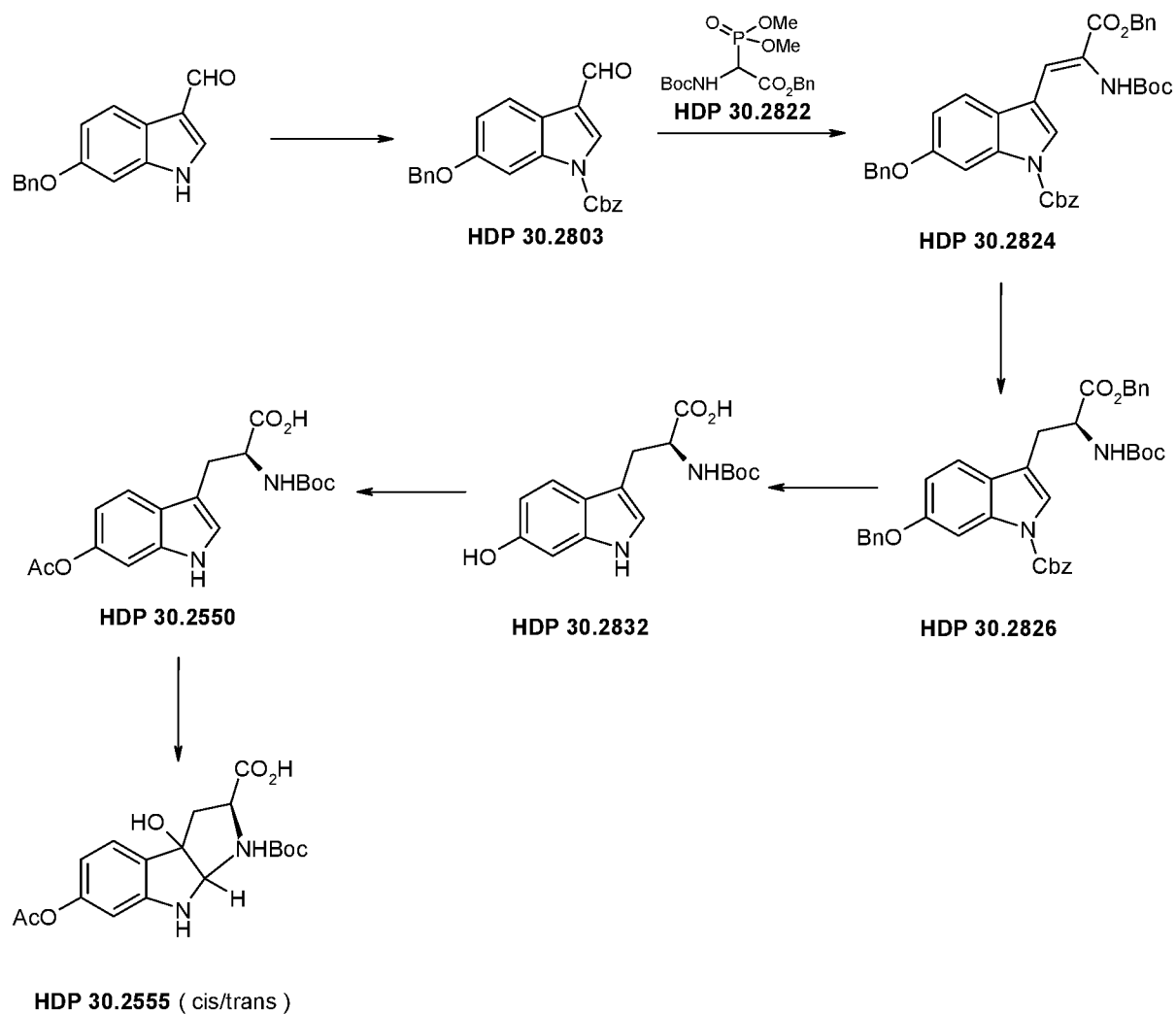
2.0g (7.1mmol) HDP 30.2821 in 90ml dry dichloromethane was treated with 4.6ml (44.1mmol) benzylic alcohol 230mg DMAP and 2.2g (10.6mmol) DCC dissolved in 7ml dichloromethane. The reaction mixture was stirred under argon at ambient temperature for 24 hours. Then the urea was filtered off and the organic phase washed with 5% citric acid and dried over MgSO_4 . After the evaporation of the dichloromethane, the remaining semi-solid was taken off in ethyl acetate and filtered again to remove additional urea. The crude product was purified by flash chromatography on a 330 g silica gel column (detection wave length 254 nm) with a gradient of n-hexane to n-hexane / ethyl acetate (1:2) and gave after evaporation 1.94g (73 %) HDP 30.2822 as a white solid.

MS (ESI⁺) found: 373.92 [MH]⁺; calc.: 373.13 (C₁₆H₂₄O₇P)

MS (ESI⁺) found: 396.17 [M+Na]⁺

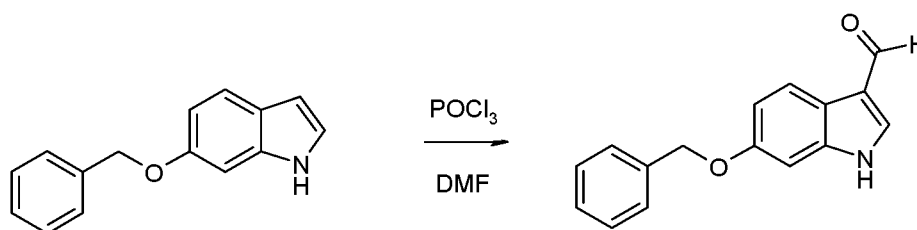
Example 2: Synthesis of (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan HDP 30.2550 as precursor of HDP 30.2555 (Hydroxy-Hpi)

The synthesis pathway is summarized in the following synthesis scheme.

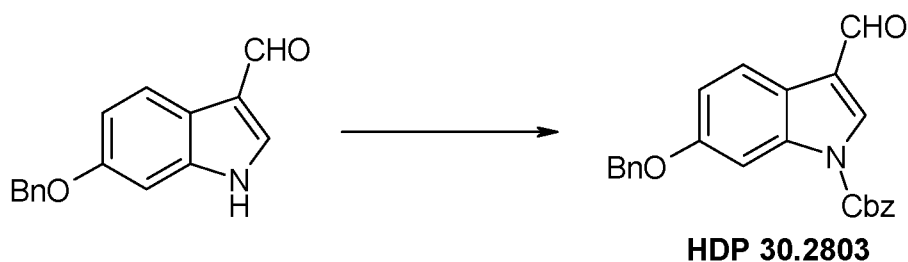


Example 2.1: Preparation of N-Cbz-6-benzyloxy-indole-3-aldehyde HDP 30.2803

The starting material 6-Benzyloxyindole-3-aldehyde for the synthesis is commercially available or can be produced by Vilsmeier reaction in high yields, starting from 6-Benzyloxyindole.



No chromatography is required for purification.

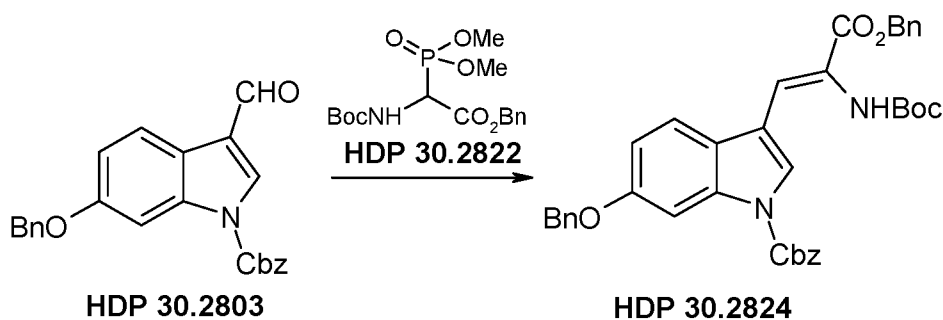


Triethylamine (1.66 mL, 11.94 mmol, 1.50 eq) was added via syringe to a solution of 6-benzyloxy-3-formyl indole (2.00 g, 7.96 mmol, 1 eq) and (DMAP) 4-dimethylaminopyridine (97.23 mg, 796 μ mol) in dichloromethane (20 mL) at 23 °C. Benzyl chloroformate (1.45 mL, 10.35 mmol, 1.30 eq) was added dropwise to the solution via syringe. After 1 h, another portion of benzyl chloroformate (223 μ L, 1.59 mmol, 0.20 eq) was added via syringe. After 95 min, the reaction mixture was diluted with dichloromethane (85 mL) and washed with saturated aqueous sodium bicarbonate solution (85 mL). The aqueous layer was further extracted with dichloromethane (2 $\times$ 20 mL). The combined organic layers were washed with aqueous hydrogen chloride (1 N, 85 mL) and the resulting aqueous layer was extracted with dichloromethane (2 $\times$ 20 mL). The combined organic layers were dried over anhydrous MgSO_4 , were filtered, and were concentrated under reduced pressure. The crude product was purified by flash chromatography on a 330 g silica gel column (detection wave length 254 nm) with a gradient of n-hexane/ ethyl acetate 4:1 to n-hexane / ethyl acetate (1:1) and gave after evaporation 2.33g (76 %) HDP 30.2803 as a white solid.

$^1\text{H-NMR}$ (400MHz, CDCl_3 , δ =ppm)

δ =5.05 (s, 2H, OCH_2); 5.47 (s, 2H, COOCH_2); 7.06 – 8.14 (m, Ar-H, 14H); 10.01 (s, 1H, CHO)

Example 2.2: Preparation of [6-Benzyloxy-1H-(benzyloxycarbonyl)-3-indole]-2-(tert-butyloxycarbonylamino)-acrylic acid benzyl ester HDP 30.2824

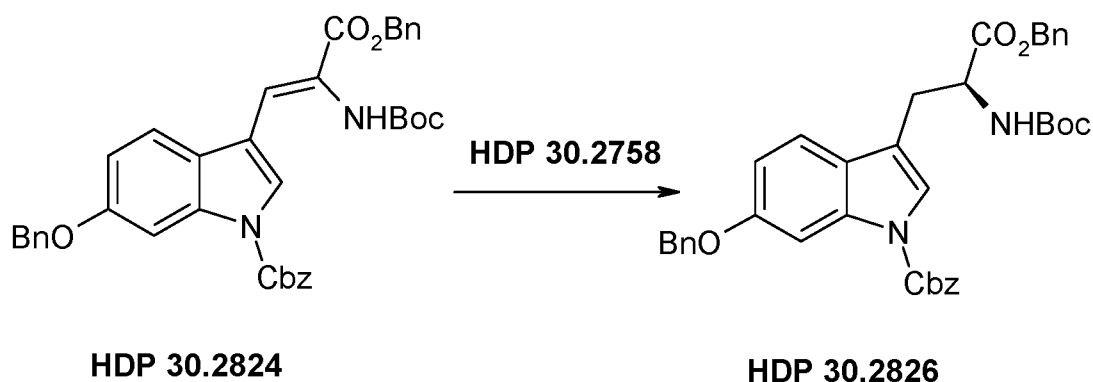


1.90g (5.09 mmol) (R,S)-N-Boc- α -dimethylphosphono)-glycine benzyl ester HDP 30.2822 was suspended under argon in 8ml dichloromethane. 0.705ml (4.73 mmol) DBU was added. After 10 minutes stirring, 1.66g (4.31mmol) N-Cbz-6-benzyloxy-indole-3-aldehyde HDP 30.2803 in 4.7ml dichloromethane was slowly added. The reaction mixture was stirred for 5 hours and the solvent was evaporated under reduced pressure. The residue was dissolved in 120ml ethyl acetate, and the organic solution was washed 2 times with 50ml 1N HCl and 50ml brine, dried over MgSO_4 and concentrated under reduced pressure to give 2.70g of crude material. The crude product was purified by flash chromatography on a 330 g silica gel column (detection wave length 254 nm) with a gradient of n-hexane to n-hexane / ethyl acetate (1:1) and gave after evaporation 2.00g (73 %) HDP 30.2824 as a white solid.

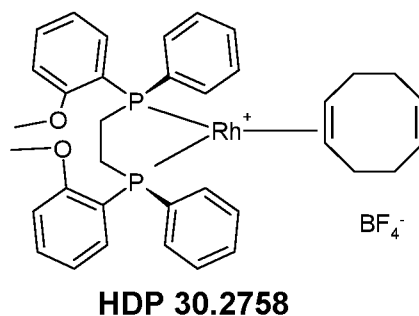
MS (ESI^+) found: 632.92 $[\text{MH}]^+$; calc.: 632.25 ($\text{C}_{38}\text{H}_{36}\text{N}_2\text{O}_7$)

MS (ESI^+) found: 655.25 $[\text{M}+\text{Na}]^+$

Example 2.3: Preparation of (S)- 6-Benzyloxy-N-tert-butoxycarbonyl-1-Cbz-L-tryptophan benzyl ester HDP 30.2826



Example 2.3.1: Synthesis of cyclooctadiene-1,5-[(R,R)-DIPAMP] rhodium tetrafluoroborate catalyst HDP 30.2758



97.0mg (0.20mmol) Bis(cyclooctadiene-1,5)-dichloro dirhodium $[\text{Rh}(\text{COD})\text{Cl}]_2$ (CAS:12092-47-6, Alfa Aesar) was added to a suspension of 180.0mg (0.39mmol) (R,R)-DIPAMP (CAS:55739-58-7, Alfa Aesar) in 2.0ml methanol/water (1.5ml/0.5ml). The orange colored slurry, stirred for 1 hour under argon, gave an orange solution. The complex was precipitated by adding slowly (over 30 minutes) a solution of 65.0mg (0.6mmol) sodium tetrafluoroborate in 0.5ml water. After 2.5 hours stirring at room temperature the orange crystals were filtered off, washed twice with small portions of water, and dried at high vacuum. 240mg (81%) of the catalyst cyclooctadiene-1,5-[(R,R)-DIPAMP]rhodium tetrafluoroborate HDP 30.2758 was obtained as a bright yellow powder. The catalyst was used without further purification.

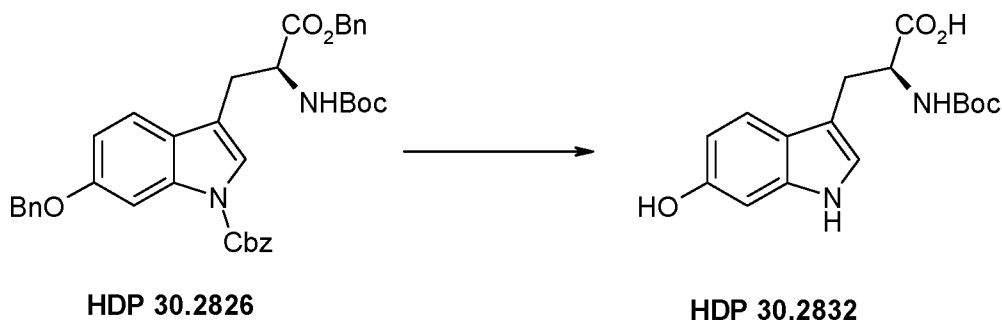
Example 2.3.2: Synthesis of (S)- 6-Benzyloxy-N-tert-butoxycarbonyl-1-Cbz-L-tryptophan benzyl ester HDP 30.2826

A 250ml stainless steel autoclave was charged with 35.0mg (0.08mmol) cyclooctadiene-1,5-[(R,R)-DIPAMP]rhodium tetrafluoroborate HDP 30.2758, 1000mg (1.8mmol) [6-Benzyloxy-1H-(benzyloxycarbonyl)-3-indole]-2-(tert-butyloxycarbonylamino)-acrylic acid benzyl ester HDP 30.2824 in 40ml dry methanol / 15ml dichloromethane. After four vacuum/Ar and H_2 cycles, the reaction was pressurized to an initial pressure of 12 bar. The reaction was allowed to proceed for 4 days at ambient temperature. After the evaporation of the solvent, the crude product was purified by flash chromatography on a 220 g silica gel column (detection wave length 254 nm) with n-hexane / ethyl acetate (3:1) and gave after evaporation 0.79g (79 %) HDP 30.2826 as a white powder.

MS (ESI^+) calc.: 634.26 ($\text{C}_{38}\text{H}_{38}\text{N}_2\text{O}_7$)

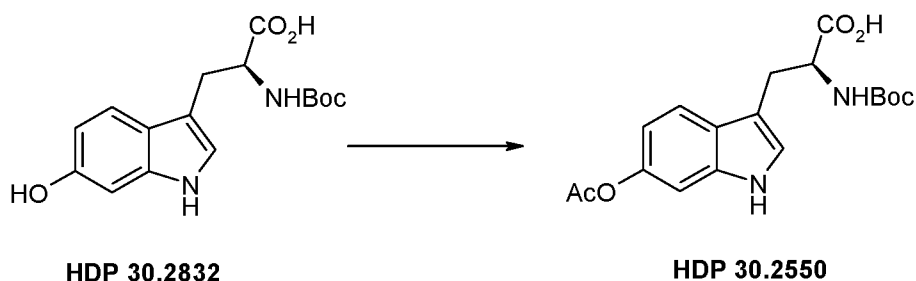
MS (ESI^+) found: 657.33 $[\text{M}+\text{Na}]^+$;

Example 2.4: Preparation of N-tert-butoxycarbonyl-(S)-tryptophan HDP 30.2832



700mg (1.10mmol) (S)-6-Benzyloxy-N-tert-butoxycarbonyl-1-Cbz-tryptophan benzyl ester HDP 30.2826 was hydrogenated with 100mg Pd-C 10% in a mixture of 7ml ethyl acetate and 4ml methanol. After 3 hours hydrogenation (TLC control with chloroform/methanol 19:1 + 1% AcOH) at room temperature and 1 atm., the catalyst was removed by filtering over a plug of Celite®. The solvent was removed, and the remaining crude residue, 378mg of HDP 30.2832, was used for the next step without purification.

Example 2.5: Preparation of (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan HDP 30.2550



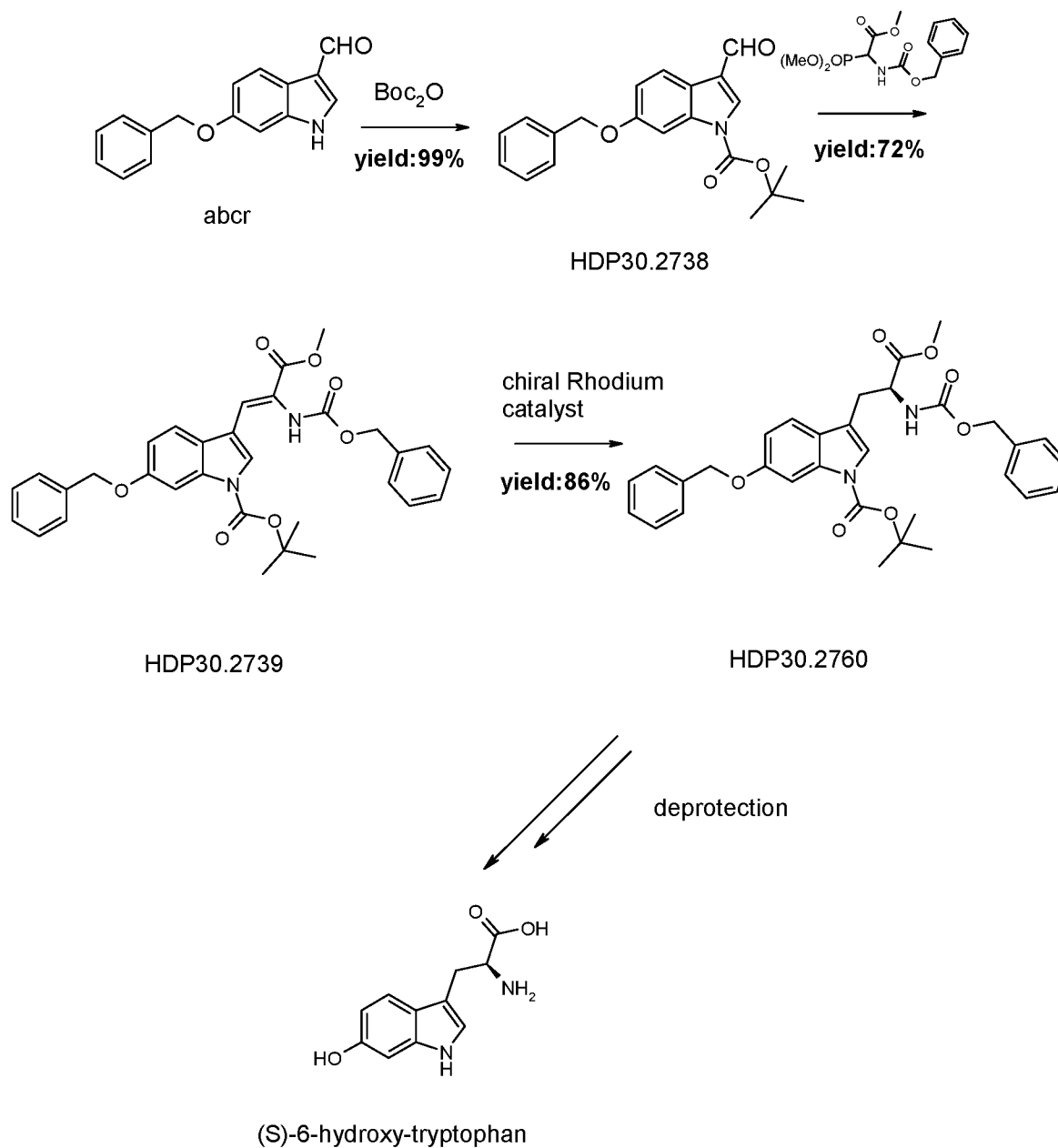
378mg crude HDP 30.2832 (assumed 1.10mmol) was dissolved in 2.21ml 1 N NaOH. Under argon and ambient temperature, 208.5µl (2.20mmol) acetic anhydride was added at once. The mixture was stirred for 3.5 hours and acidified with 5% citric acid. The reaction mixture was extracted 3x with ethyl acetate, and the combined organic phases washed with 5% sodium chloride and dried over MgSO₄. Filtration and evaporation to dryness gave 380mg crude material. The crude product was purified by flash chromatography on a 120 g silica gel column (detection wave length 254 nm) with a gradient of dichloromethane + 2% AcOH / dichloromethane / methanol (15:1) + 2% AcOH and gave after evaporation 270mg (68 %) HDP 30.2550 as a white solid.

MS (ESI⁺) found: 361.17 [M-H]⁺; calc.: 362.15 (C₁₈H₂₂N₂O₆)

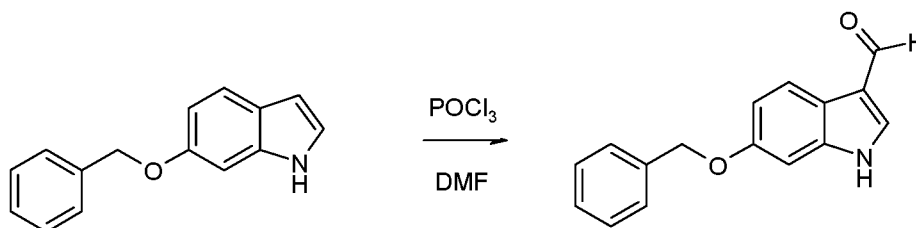
MS (ESI⁺) found: 723.08 [2M-H]⁺

Example 3: Preparation of (S)-6-Hydroxy-Tryptophan by asymmetric hydrogenation of dehydro amino acid

The synthesis pathway is summarized in the following synthesis scheme.

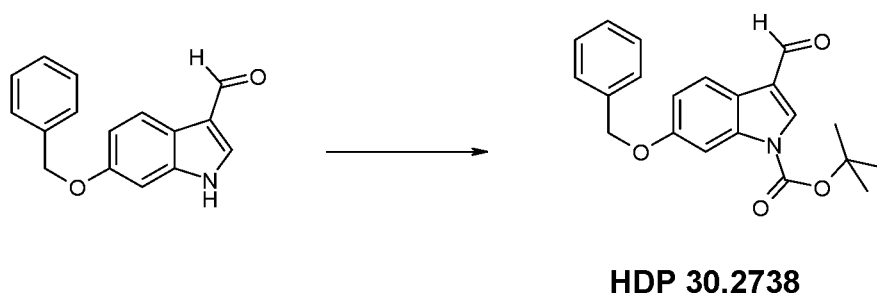


Example 3.1: Preparation of 6-Benzyloxy-1H-indole-3-carbaldehyde



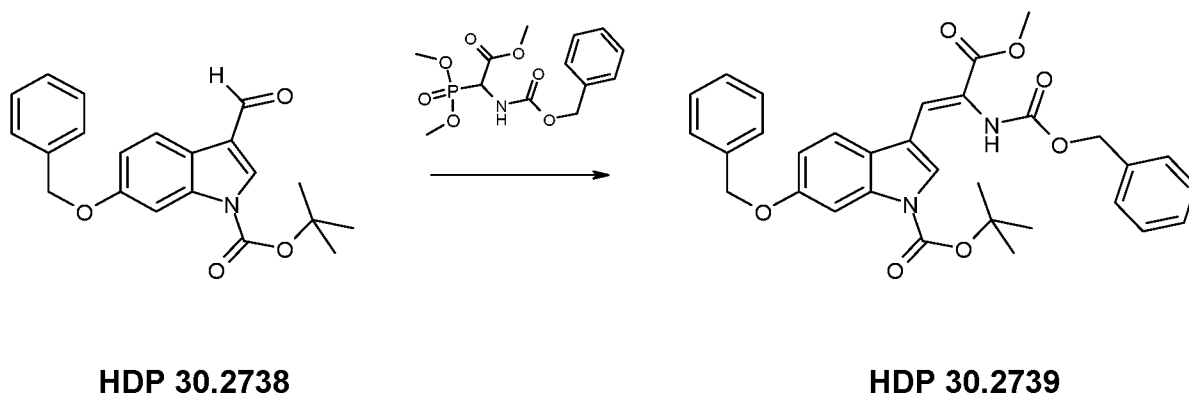
To a stirred solution of phosphorus oxychloride (10.0 mL, 107.0mmol) in DMF (35 mL) a solution of 6-benzyloxyindole (22.3 g, 100.0mmol) in DMF (25 mL) was added at room temperature. After 45 min, the reaction mixture was poured into ice-water (200 mL). To this mixture solid NaOH (19.0 g, 475.0mmol) and water (100 mL) was added. After 30 minutes additional water (200ml) was added and the whole mixture was refluxed for 3 minutes. The precipitate was collected, washed with 5 portions of 50ml cold water and dried to give 24.8 g (98.8%) 6-Benzyloxy-1H-indole-3-carbaldehyde as a white powder. The compound was identical with reference material and sufficiently pure for the next reaction.

Example 3.2: Preparation of 6-Benzyloxy-1H-1-tert-butoxycarbonyl-indole-carbaldehyde HDP 30.2738



10.0g (39.8mmol) 6-Benzyloxy-1H-indole-3-carbaldehyde was suspended in 100ml dichloromethane and treated with 0.56g (4.5mmol) 4-dimethylaminopyridine DMAP and 10.5g (47.3mmol) di-tert-butyl dicarbonate Boc_2O dissolved in 10ml dichloromethane. After stirring for 2 hours, 100ml 1N KHSO_4 was added and dichloromethane was evaporated. The aqueous layer was extracted with several portions of diethyl ether (2 x 200ml) and the combined organic extracts were washed with 250ml 1N KHSO_4 , 250ml 1N NaHCO_3 and 250ml brine. The organic layer was dried over MgSO_4 and concentrated under reduced pressure to give 12.0g (86%) red-brownish powder. The compound was sufficiently pure for the next reaction step.

Example 3.3: Preparation of 3-[6-Benzyloxy-1H-(1-tert-butoxycarbonyl)-3-indole]-2-(benzyloxycarbonylamino)-acrylic acid methyl ester HDP 30.2739

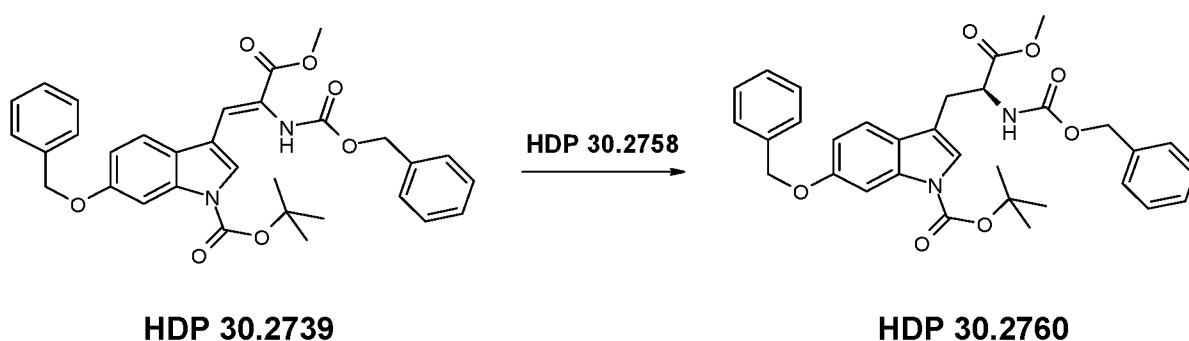


5.12g (15.44mmol) (R,S)-Benzyloxycarbonyl- δ -phosphono-glycine trimethylester (CAS: 88568-95-0, Alfa Aesar) was dissolved under argon in 18ml dichloromethane. 2.14ml (14.31mmol) DBU was added. After 10 minutes stirring, 4.60g (13.07mmol) 6-Benzyloxy-1H-1-tert-butyloxycarbonyl-indole-3-carbaldehyde HDP 30.2738 in 14ml dichloromethane was added slowly. The reaction mixture was stirred for 6 hours and the solvent was evaporated under reduced pressure. The residue was dissolved in 300ml ethyl acetate, then the organic solution was washed 2 times with 120ml 1N HCl and 120ml brine, dried over MgSO_4 and concentrated under reduced pressure to give 7.43g of crude material. The crude product was purified by flash chromatography on a 330 g silica gel column (detection wave length 254 nm) with a gradient of n-hexane to n-hexane / ethyl acetate (2:1) and gave after evaporation 5.23g (72 %) HDP 30.2739 as a white solid.

MS (ESI⁺) found: 557.17 [MH]⁺; calc.: 557.22 (C₃₂H₃₂N₂O₇)

MS (ESI⁺) found: 579.25 [M+Na]⁺

Example 3.4: Preparation of 6-Benzyloxy-N-Carbobenzyloxy-1-tert-butyloxycarbonyl-L-tryptophan methyl ester HDP 30.2760



Example 3.4.1: Synthesis of cyclooctadiene-1,5-[(R,R)-DIPAMP] rhodium tetrafluoroborate HDP 30.2758

The catalyst cyclooctadiene-1,5-[(R,R)-DIPAMP]rhodium tetrafluoroborate HDP 30.2758 was synthesized as described in Example 2.3.1.

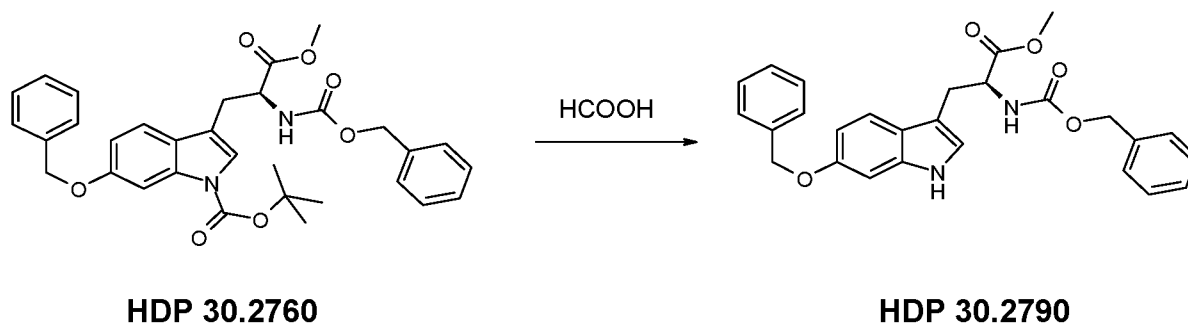
Example 3.4.2: Synthesis of (S)-6-Benzyloxy-N-carbobenzyloxy-1-tert-butoxycarbonyl-tryptophan methyl ester HDP 30.2760

A 250ml stainless steel autoclave was charged with 60.0mg (0.08mmol) cyclooctadiene-1,5-[(R,R)-DIPAMP]rhodium tetrafluoroborate HDP 30.2758 and 1000mg (1.8mmol) [6-Benzyloxy-1H-(1-tert-butoxycarbonyl)-3-indole]-2-(benzyloxycarbonyl-amino)acrylic acid methyl ester HDP 30.2739 in 40ml dry methanol. After four vacuum/Ar and H₂ cycles, the reaction was pressurized to an initial pressure of 30 bar. The reaction was allowed to proceed for 4 days at ambient temperature. After the evaporation of the solvent, the crude product was purified by flash chromatography on a 120 g silica gel column (detection wave length 254 nm) with a gradient of n-hexane to n-hexane / ethyl acetate (2:1) and gave after evaporation 0.85g (86 %) HDP 30.2760 as a white solid.

MS (ESI⁺) calc.: 558.23 (C₃₃H₃₄N₂O₇)

MS (ESI⁺) found: 581.17 [M+Na]⁺; 1138.83 [2M+Na]⁺

Example 3.5: Preparation of 6-Benzyloxy-N-carbobenzyloxy-L-tryptophan methylester HDP 30.2790

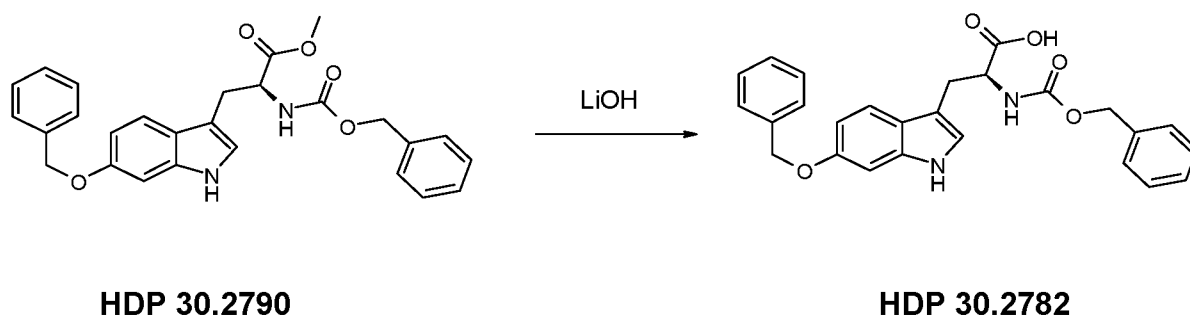


100.0mg (0.18mmol) (S)-6-Benzyloxy-N-carbobenzyloxy-1-tert-butoxycarbonyl-tryptophan methyl ester HDP 30.2760 was dissolved in 5.0ml formic acid and stirred 1 hour at 40°C. The reaction mixture was evaporated to dryness and the residue dissolved in ethyl acetate. The ethyl acetate solution was washed with water, saturated NaHCO₃ and brine and dried over MgSO₄. After the evaporation of the solvent, the crude product was purified by flash chromatography on a 24 g silica gel column (detection wave length 254 nm) with a gradient of n-hexane to n-hexane / ethyl acetate (1:1) and gave after evaporation 29mg (35 %) HDP 30.2790 as a white solid.

MS (ESI⁺) calc.: 458.52 (C₂₇H₂₆N₂O₅)

MS (ESI⁺) found: 459.25 [M+H]⁺

**Example 3.6: Preparation of (S)-6-Benzyloxy-N-Carbobenzyloxy-tryptophan
HDP 30.2782**

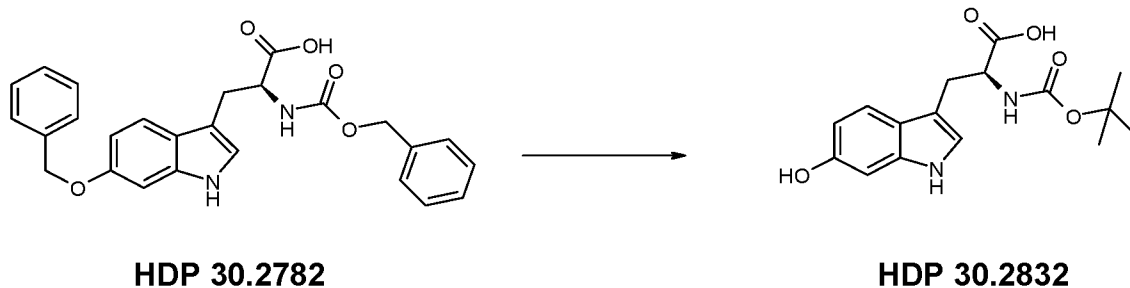


A 2N aqueous solution of LiOH (84.7μl) was added to a solution of HDP 30.2790 25.9mg (0.056mmol) in 1000μl tetrahydrofuran/water (10:1) at ambient temperature. The reaction mixture was stirred for 2.5 hours and partitioned between ethyl acetate and 5% citric acid. The aqueous layer was extracted with ethyl acetate and the organic layers were combined, dried (MgSO₄) and concentrated. The resulting carboxylic acid HDP 30.2782 was purified on silica gel using dichloromethane / methanol (+1% acetic acid) as mobile phase. 13.7mg (55%) white solid.

MS (ESI⁺) calc.: 444.17.23 (C₂₆H₂₄N₂O₅)

MS (ESI⁺) found: 445.25 [M+H]⁺; 467.17 [M+Na]⁺

Example 3.7: Preparation of (S)-6-Hydroxy-N-(tert-butoxycarbonyl)-tryptophan

HDP 30.2832

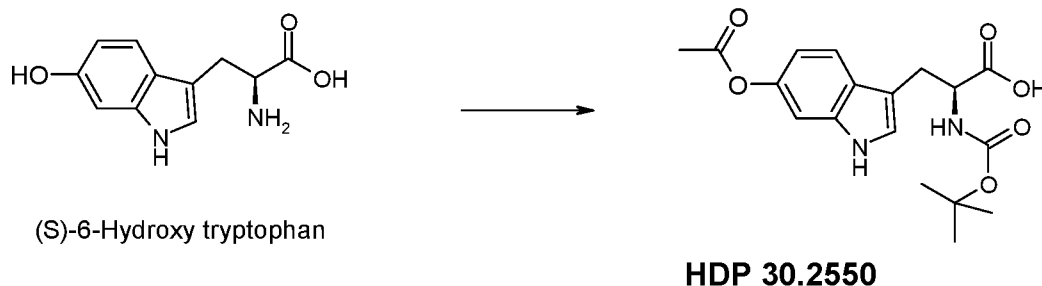
Palladium on charcoal 10mg (10%weight) was added to a solution of 50mg (0.11mmol) HDP 30.2782 in 800μl methanol. The reaction mixture was purged three times with hydrogen and stirred for 2.5 h at room temperature. The suspension was filtered through a plug of Celite[®] washed with methanol and concentrated to dryness. The solid residue (22.2mg) (S)-6-hydroxytryptophan was dissolved in 1000μl 1,4-dioxane/water (1:1) and treated with 101μl (0.101mmol) 1N NaOH and 21.57μl (0.10mmol) Di-tert-butyl dicarbonate. The reaction mixture was stirred for 16 hours and adjusted to pH 2 with 1N hydrochloric acid. The aqueous solution was extracted three times with ethyl acetate and the combined organic phases washed with brine, dried and evaporated to dryness. The crude HDP 30.2832 was purified on silica gel using dichloromethane / methanol (+1% acetic acid) as mobile phase. 9.9mg (31%) of a white solid. The material was identical with a reference sample.

MS (ES⁺) calc.: 320.14 (C₁₆H₂₀N₂O₅)

MS (ESI⁻) found: 319.08 [M-H]⁻

Example 4: Preparation of cis,trans-1-(tert-butoxycarbonyl)-2-carboxy-3a-hydroxy-6-acetoxy-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-b]indole cis-HDP 30.2555 and trans HDP 30.2555 (cis,trans-6-Acetoxy-Hpi)

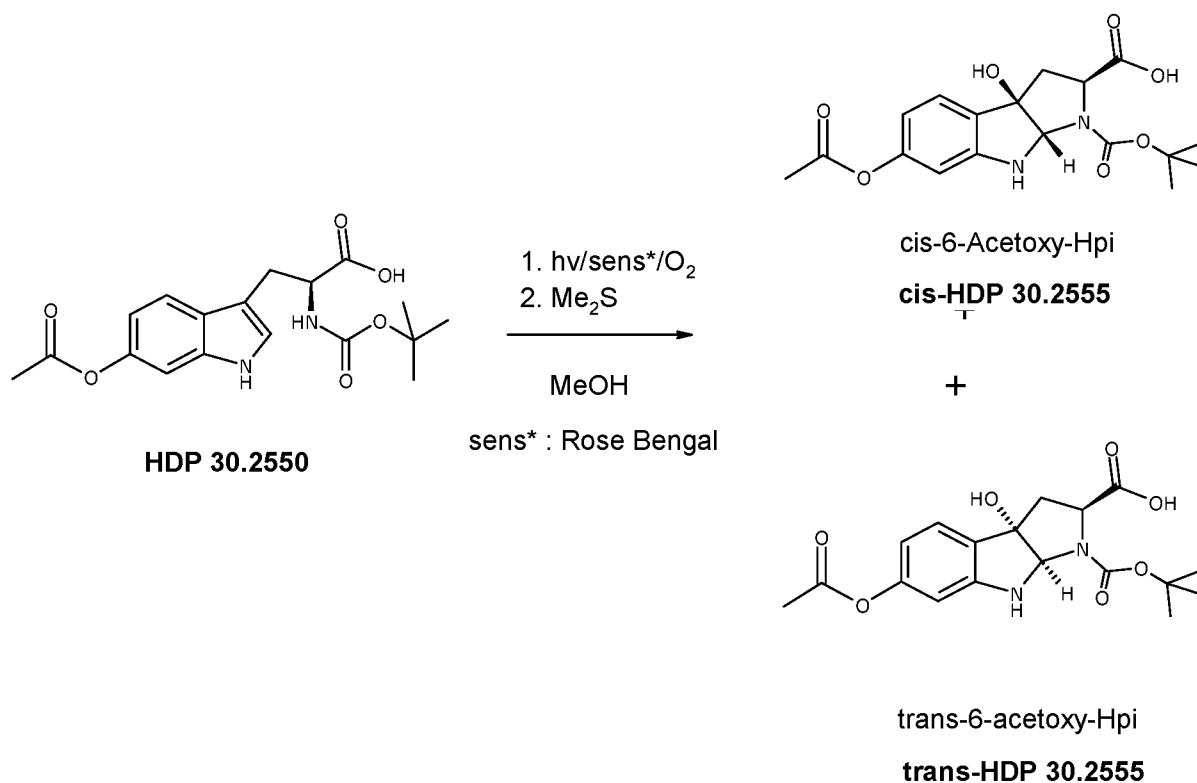
Example 4.1: Preparation of (S)-N-(tert-butoxycarbonyl)-6-acetoxy-tryptophan HDP 30.2550



590,0mg (2.68mmol) (S)-6-Hydroxytryptophan from the hydrogenation step in Example 3.7 was suspended in a mixture of 30ml 1,4-dioxane/water 1:1 (v,v). Under argon 2,68 ml (2,68mmol) 1N NaOH was added at once at ambient temperature. The resulting yellow solution was then treated with 574,6ml (2,68mmol) Boc anhydride (Boc_2O) and stirred for 24 hours at room temperature. The solution was acidified with 1N hydrochloric acid to pH 2,4 and extracted 3 times with 25ml ethyl acetate. The combined ethyl acetate extracts were washed with saturated NaCl solution and dried over MgSO_4 . Filtration and evaporation to dryness gave 785,0mg crude material. The crude N-Boc-6-hydroxy-L-tryptophan was dissolved in 4.91 ml (4.91mmol) 1N NaOH and treated with 463,2ml (500,3mg, 4,90mmol) acetanhydride. The reaction mixture was stirred for 3 hours under argon and acidified with 5% citric acid. The aqueous phase was extracted three times with 25ml ethyl acetate, washed with saturated NaCl and dried over MgSO_4 . Filtration and evaporation gave 635mg of a crude solid. The crude product was purified by flash chromatography on a 330 g silica gel column (detection wave length 254 nm) with a gradient of CH_2Cl_2 +1% acetic acid to $\text{CH}_2\text{Cl}_2/\text{MeOH}$ (15:1) +1% acetic acid and gave after co evaporation with toluene 564,4mg (56 % yield) of a white powder.

MS (ESI⁺)found: 361.08[M-H]⁺; calc.: 362.15 ($\text{C}_{18}\text{H}_{22}\text{N}_2\text{O}_6$)

Example 4.2: Preparation of cis,trans-6-Acetoxy-Hpi



The photo-oxygenation was carried out with a 400W high-pressure sodium vapor lamp (Sirius X400 lamp 230V, 400W; 55000 lumen at a distance of 1.3m). Rose Bengal is used as a dye sensitizer. The reaction was carried out in a 500ml cylindrical reaction vessel with heat exchange jacket made of borosilicate glass, flat bottom and flat laboratory flange (DN) with two connectors with GL 18 thread. The distance from lamp to reaction vessel, was 15cm and the reaction temperature was in a range of 3-4°C.

The final product was purified on a Teledyne ISCO Flash chromatography system with a 330g Silica Redi Sept Flash column (Teledyne ISCO cat. 69-2203-330). Solvents CH_2Cl_2 , CH_3OH , CH_3COOH were standard HPLC or BP grade. Dry oxygen (99.5% purity) was bubbled through the reaction mixture with a rate of 2-4 l per minute.

943,0mg (2,60mmol) N-(tert-butoxycarbonyl)-L-6-acetoxy-tryptophan HDP 30.2550 and 100mg Rose Bengal were dissolved in 500ml methanol and cooled to 3°C by using a Huber cryostat with glycol/water as cooling media. The reaction solution was irradiated with the 400W high-pressure sodium vapor lamp. During the irradiation a slow stream of oxygen was bubbled through the reaction solution. After 5 hours irradiation, oxygenation and cooling was stopped and the reaction media was treated with 10ml of dimethyl sulfide. The mixture was

stirred for 2 hours and evaporated to dryness by using a rotary evaporator with a water bath temperature of 35°C. The dark red residue was dried further in high vacuum to a crystalline solid of 1,20g. The crude product was purified on a 330g silica gel column (detection wave length 254nm) with a gradient of CH₂Cl₂ +5% acetic acid to CH₂Cl₂/MeOH (30:1) +5% acetic acid. 380mg cis-HDP 30.2555 and 290mg trans-HDP 30.2555 were eluted and co-evaporated with toluene. After lyophilisation in tert-butanol both isomers were obtained as off-white powders.

cis-1-(tert-butoxycarbonyl)-2-carboxy-3a-hydroxy-6-acetoxy-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-b]indole (cis-HDP 30.2555)

380mg cis-HDP 30.2555 yield: 39%

¹H-NMR (400MHz, CD₃OD, δ=ppm)

δ=1.22, 1.44, 1.54 [s, 9H, C(CH₃)₃]; 2.23 (s, 3H, OCOCH₃); 2.46-2.63 (m, 2H, CH₂); 4.14-4.29 (m, 1H, 2-H); 5.35 (s, 1H, 8a-H); 6.39-6.46 (m, 2H, 7-H, 5-H); 7.20- 7.24 (m, 1H, 4-H)

¹³C-NMR (100MHz, CD₃OD, δ=ppm)

δ=20.93, 28.45, 31.12, 42.80, 61.12, 69.44, 82.21, 85.82, 87.93, 104.97, 112.98, 124.84, 129.42, 151.51, 154.04, 155.97, 171.34, 175.79

MS (ESI⁺) found: 378.92 [MH]⁺; calc.: 378.14 (C₁₈H₂₂N₂O₇)

MS (ESI⁺) found: 401.17 [M+Na]⁺; calc.: 401.14 (C₁₈H₂₂N₂NaO₇)

UV/VIS (CH₃OH): λ_{max} = 296nm, 239nm, 215nm

λ_{min} = 266nm, 227nm

trans-1-(tert-butoxycarbonyl)-2-carboxy-3a-hydroxy-6-acetoxy-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-b]indole (trans-HDP 30.2555)

290mg trans-HDP 30.2555 yield: 30%

¹H-NMR (400MHz, CD₃OD, δ=ppm)

δ=1.22,1.45, 1.54 [s, 9H, C(CH₃)₃]; 2.22 (s, 3H, OCOCH₃); 2.55-2.73 (m, 2H, CH₂); 4.51-4.57 (m, 1H, 2-H); 5.21-5.24 (s, 1H, 8a-H); 6.36-6.41 (m, 2H, 7-H, 5-H); 7.17- 7.18 (m, 1H, 4-H)

¹³C-NMR (100MHz, CD₃OD, δ=ppm)

δ =20.95, 28.50, 31.12, 42.47, 60.97, 69.44, 82.06, 84.84, 87.54, 104.74, 112.67, 125.03, 128.70, 152.31, 154.22, 156.00, 171.23, 174.67

MS (ESI⁺) found: 379.00 [MH]⁺; calc.: 378.14 (C₁₈H₂₂N₂O₇)

MS (ESI⁺) found: 401.17 [M+Na]⁺; calc.: 401.14 (C₁₈H₂₂N₂NaO₇)

MS (ESI⁺) found: 779.00 [2M+Na]⁺; calc.: 779.28 (C₃₆H₄₄N₄Na₂O₁₄)

UV/VIS (CH₃OH): $\lambda_{\max}$ = 299nm, 241nm, 215nm

$\lambda_{\min}$ = 268nm, 228nm

Example 4.3: Introduction of cis,trans-6-Acetoxy-Hpi into amanitin precursor

Synthesis of amanitin using cis,trans-6-Acetoxy-Hpi was performed as described in PCT/EP2018/071268, the content of which is incorporated by reference hereby, mainly for purposes of enablement.

References:

Muraoka S, and Shinozawa T., J Biosci Bioeng. 2000;89(1):73-6

Wieland T., Faulstich H. 1978. CRC Crit Rev Biochem. Vol. 5: 185-260.

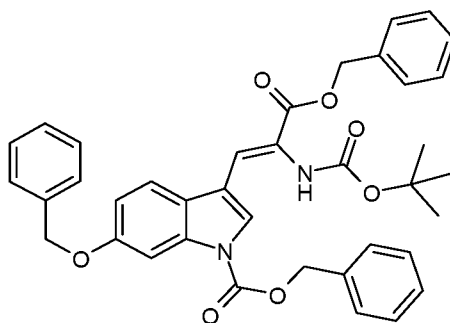
Zanotti G, Mähringer C, and Wieland T., Int J Pept Protein Res. 1987 Oct;30(4):450-9;

Zanotti G, Wieland T, Benedetti E, Di Blasio S, Pavone V, and Pedone C., Int J Pept Protein Res. 1989 Sep;34(3):222-8

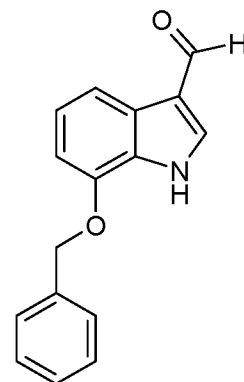
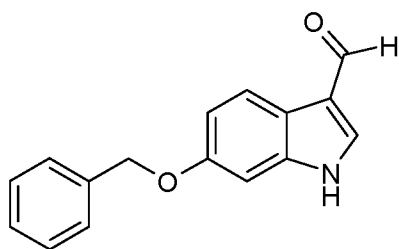
Zhang P, *et al.*, FEMS Microbiol Lett. 2005 Nov 15;252(2):223-8. Epub 2005 Sep 15

What is claimed is:

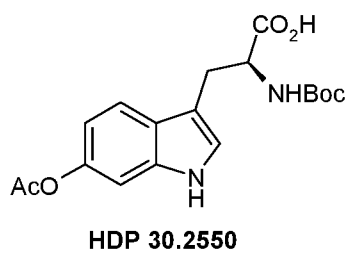
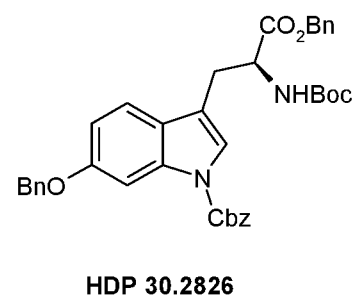
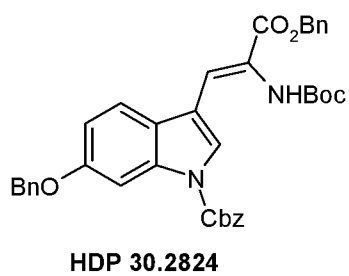
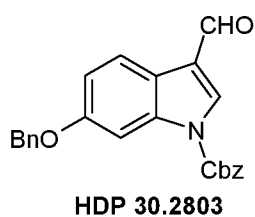
1. Method for synthesis of (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan (HDP 30.2550) or (S)-6-hydroxytryptophan, wherein said method comprises at least one step of enantiomer-selective hydrogenation of an olefinic precursor compound by use of at least one chiral catalyst.
2. The method according to claim 1, wherein said olefinic precursor is an olefinic amino acid precursor, preferably wherein said olefinic precursor is compound HDP 30.2824.

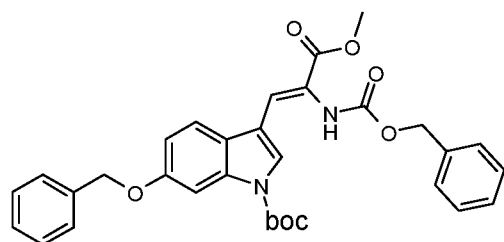
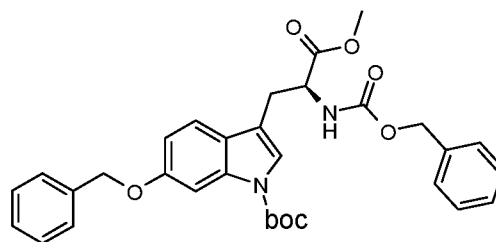
**HDP 30.2824**

3. The method according to any of claims 1 or 2, wherein said chiral catalyst is a compound selected from the group consisting of compound HDP 30.2758, (R,R)-Et-DUPHOS (BF₄-), (R,R)-DuPhos-Ferrocene (BF₄-), (R,R)-DuPhos-Ferrocene-Et₂ (BF₄-), (R,R)-DuPhos-Alkyl (CF₃SO₃-), and (R,R)-Phenyl-DuPhos- Alkyl (BF₄-).
4. The method according to claim 3, wherein said chiral catalyst is compound HDP 30.2758 or (R,R)-Et-DUPHOS (BF₄-).
5. The method according to any of claims 1 - 4, wherein said olefinic precursor is synthesized by use of compound HDP 30.2822.
6. The method according to claim 5, wherein said olefinic precursor is further synthesized by use of compound B.

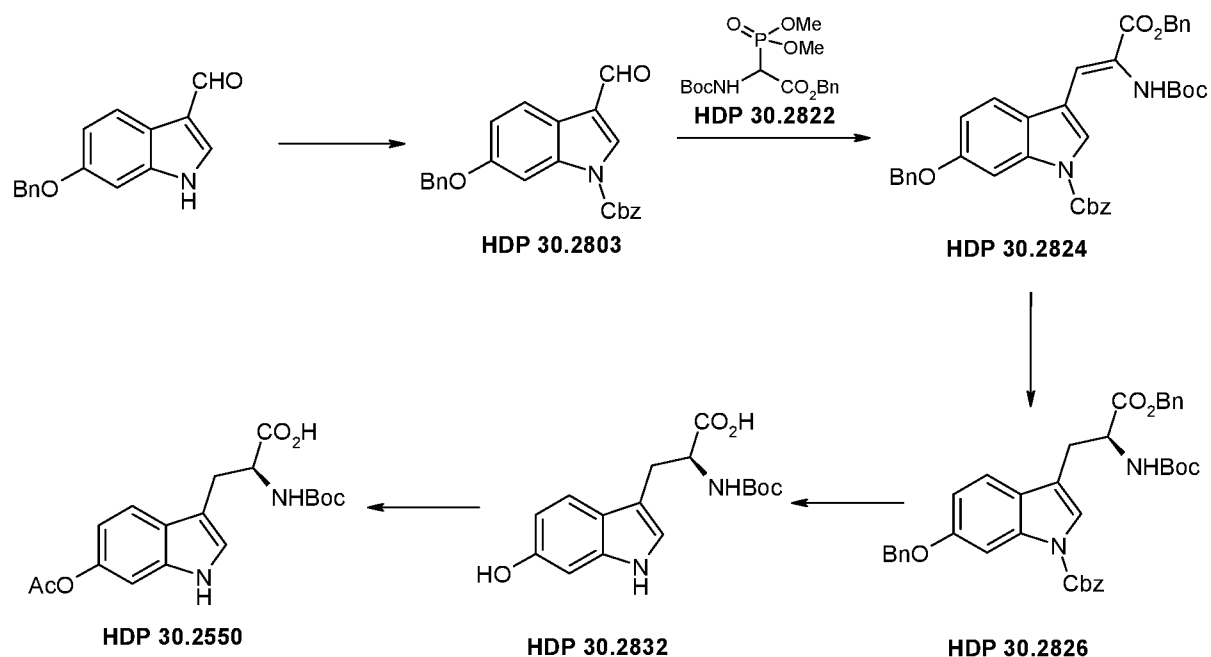
**Compound B**

7. The method according to any of claims 1 – 6, wherein said method comprises the use of at least one starting or intermediate compound selected from the group consisting of

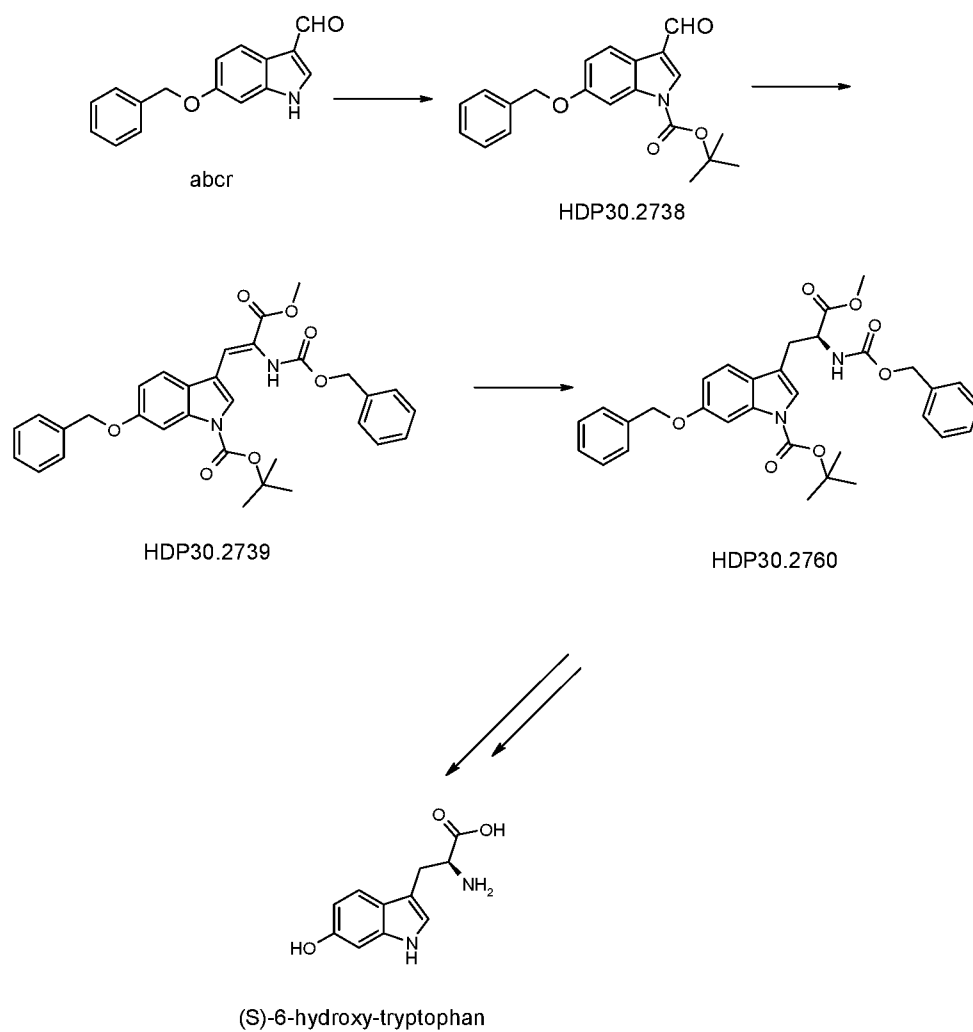


**HDP30.2739****HDP30.2760**

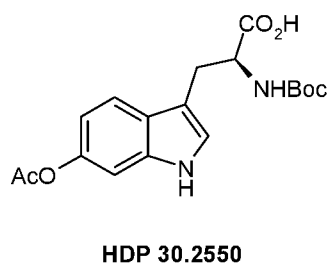
8. The method for synthesis of (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan (HDP 30.2550) according to any of claims 1 – 7, wherein said method comprises at least the following steps:



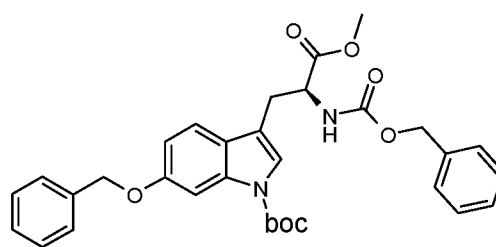
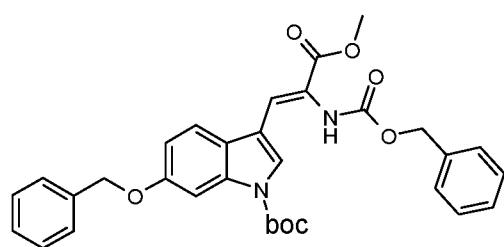
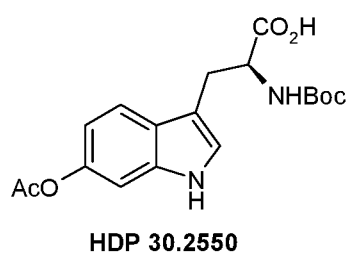
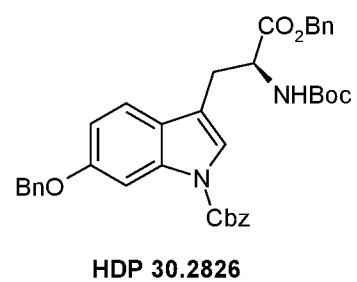
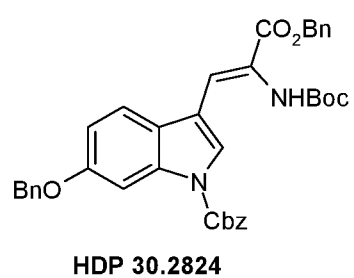
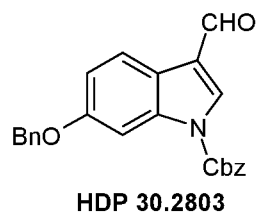
9. The method for synthesis of (S)-6-hydroxytryptophan according to any of claims 1 – 7, wherein said method comprises at least the following steps:

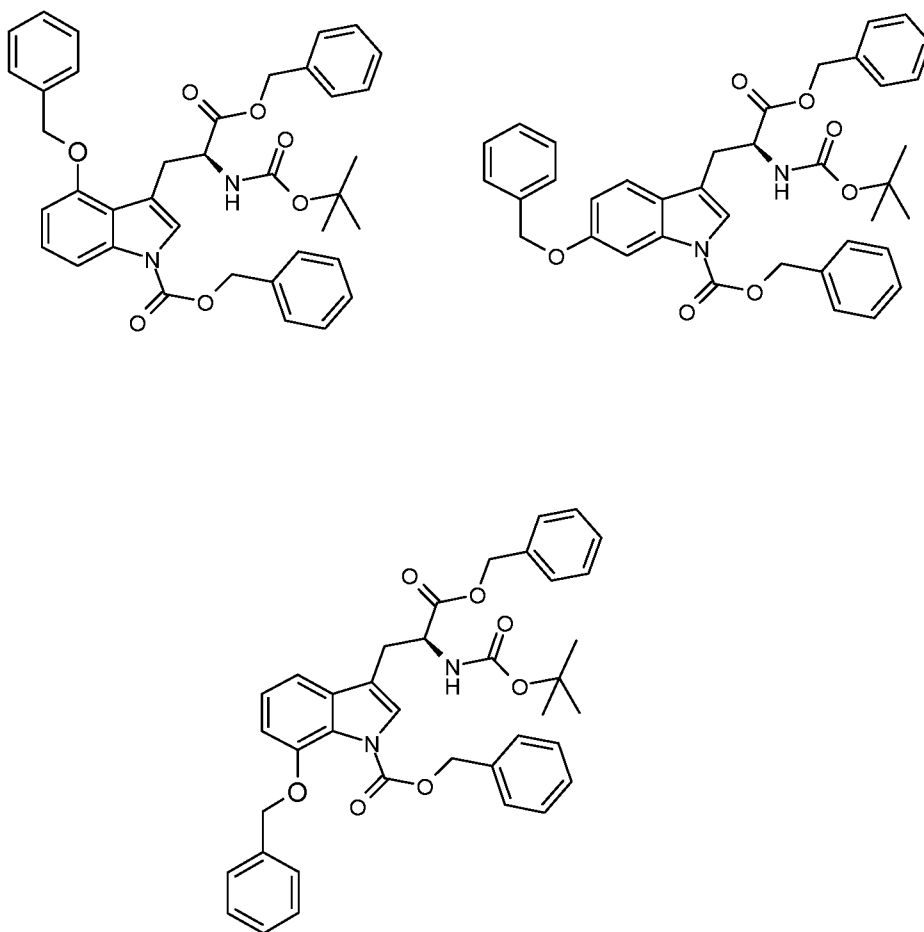


10. (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan (**HDP 30.2550**),

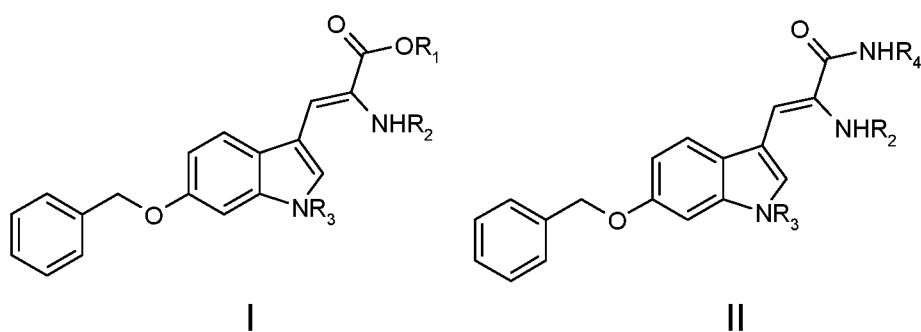


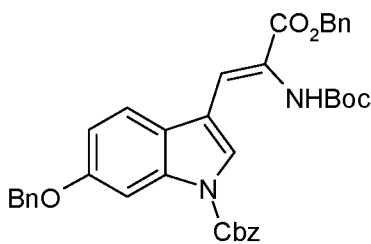
or any of the precursor or derivative compounds selected from the group consisting of:



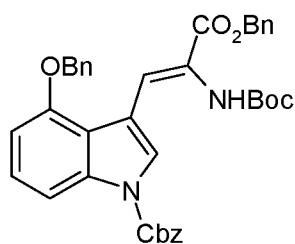


11. (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan (HDP 30.2550), (S)-6-hydroxy-tryptophan or any of the precursor compounds according to claims 7, 8 and 9, respectively, for use in synthesis of amanitin or amanitin derivatives or amatoxin-drug conjugates, wherein said amatoxin-drug conjugate optionally comprises a linker.
12. A dehydroamino acid compound selected from the group consisting of compounds I, II, III, IV and V

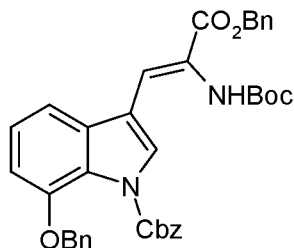




III



IV



V

wherein

R1 is selected from: H, alkyl, alkenyl, arylalkyl optional substituted,

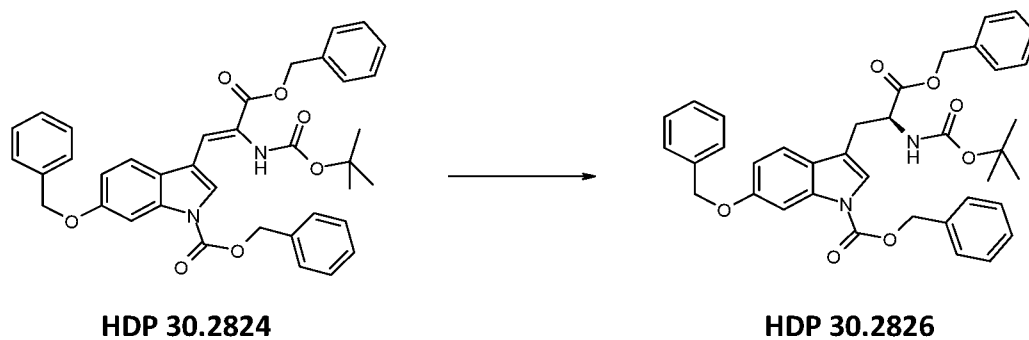
R2 is selected from: Boc, Cbz, N protecting groups,

R3 is selected from: Boc, Cbz, N protecting groups,

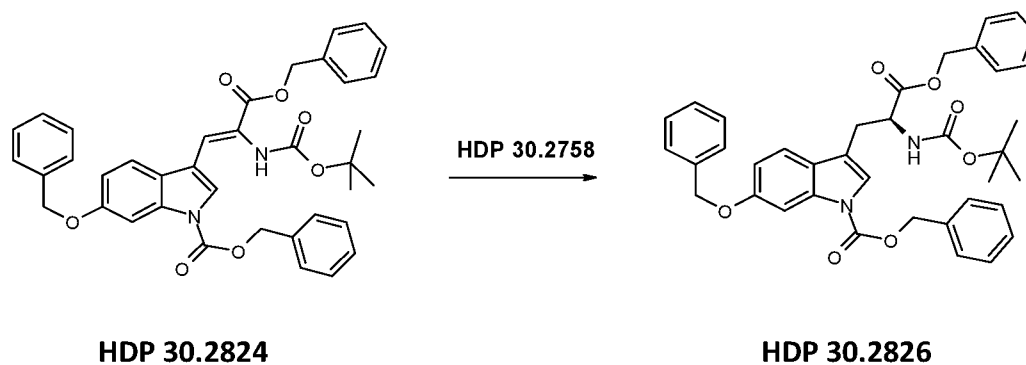
R4 is an amino acid residue,

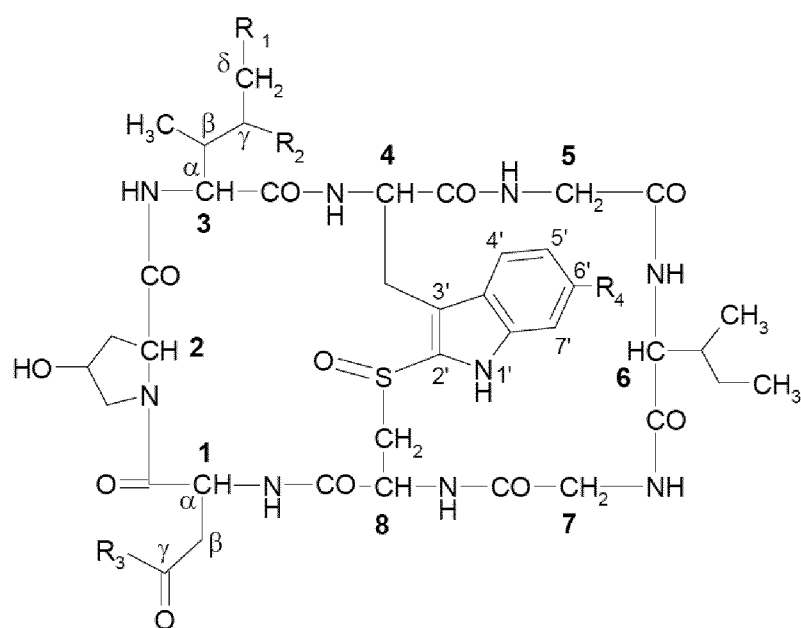
for use in synthesis of amanitin or amanitin derivatives or amatoxin-drug conjugates.

13. Use of (S)-6-Acetyloxy-N-tert-butoxycarbonyl-tryptophan (HDP 30.2550) or (S)-6-hydroxy-tryptophan for synthesis of amanitin or amanitin derivatives or amatoxin-drug conjugates, preferably wherein said amatoxin-drug conjugate comprises a linker.
14. Use of a compound selected from the group consisting of compound HDP 30.2758, (R,R)-Et-DUPHOS (BF₄⁻), (R,R)-DuPhos-Ferrocene (BF₄⁻), (R,R)-DuPhos-Ferrocene-Et₂ (BF₄⁻), (R,R)-DuPhos-Alkyl (CF₃SO₃⁻), and (R,R)-Phenyl-DuPhos-Alkyl (BF₄⁻) as catalyst for hydrogenation in the following reaction:



15. Use of cyclooctadiene-1,5-[(R,R)-DIPAMP] rhodium tetrafluoro-borate (HDP 30.2758) as catalyst for hydrogenation in the following reaction:





	R ₁	R ₂	R ₃	R ₄
α-amanitin	OH	OH	NH ₂	OH
β-amanitin	OH	OH	OH	OH
γ-amanitin	H	OH	NH ₂	OH
ε-amanitin	H	OH	OH	OH
amanin	OH	OH	OH	H
amaninamide	OH	OH	NH ₂	H
amanullin	H	H	NH ₂	OH
amanullinic acid	H	H	OH	OH
γ-amanin	H	OH	OH	H
γ-amaninamide	H	OH	NH ₂	H

Fig. 1

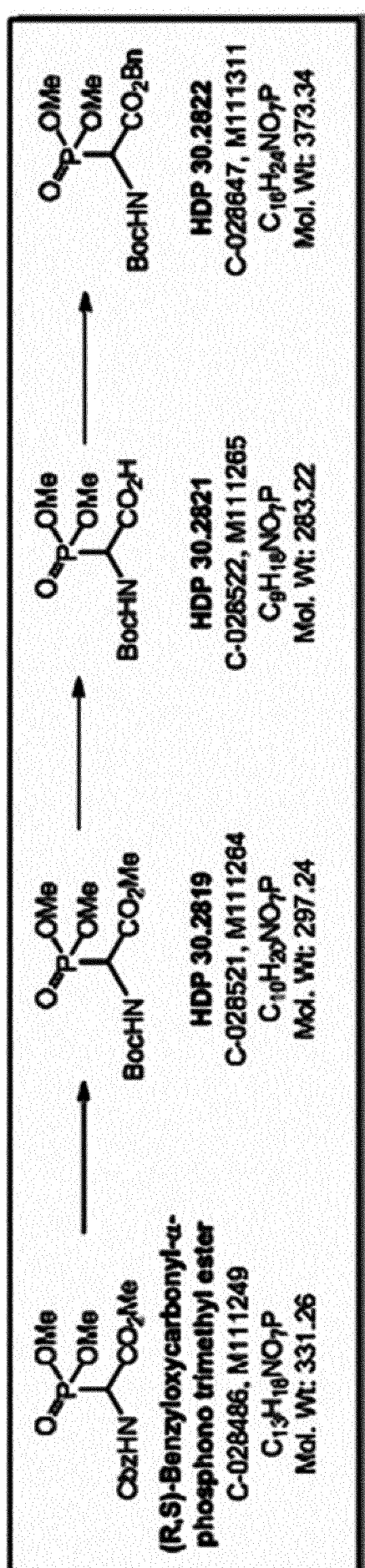


Fig. 2

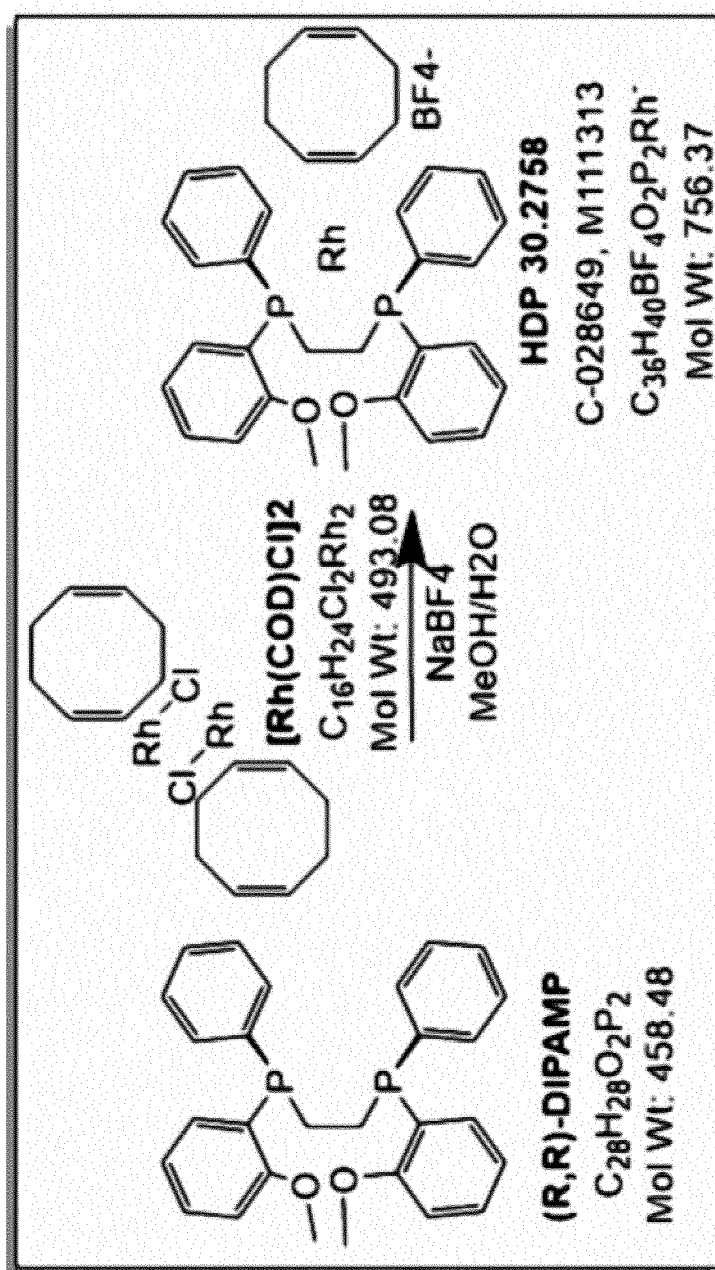


Fig. 3

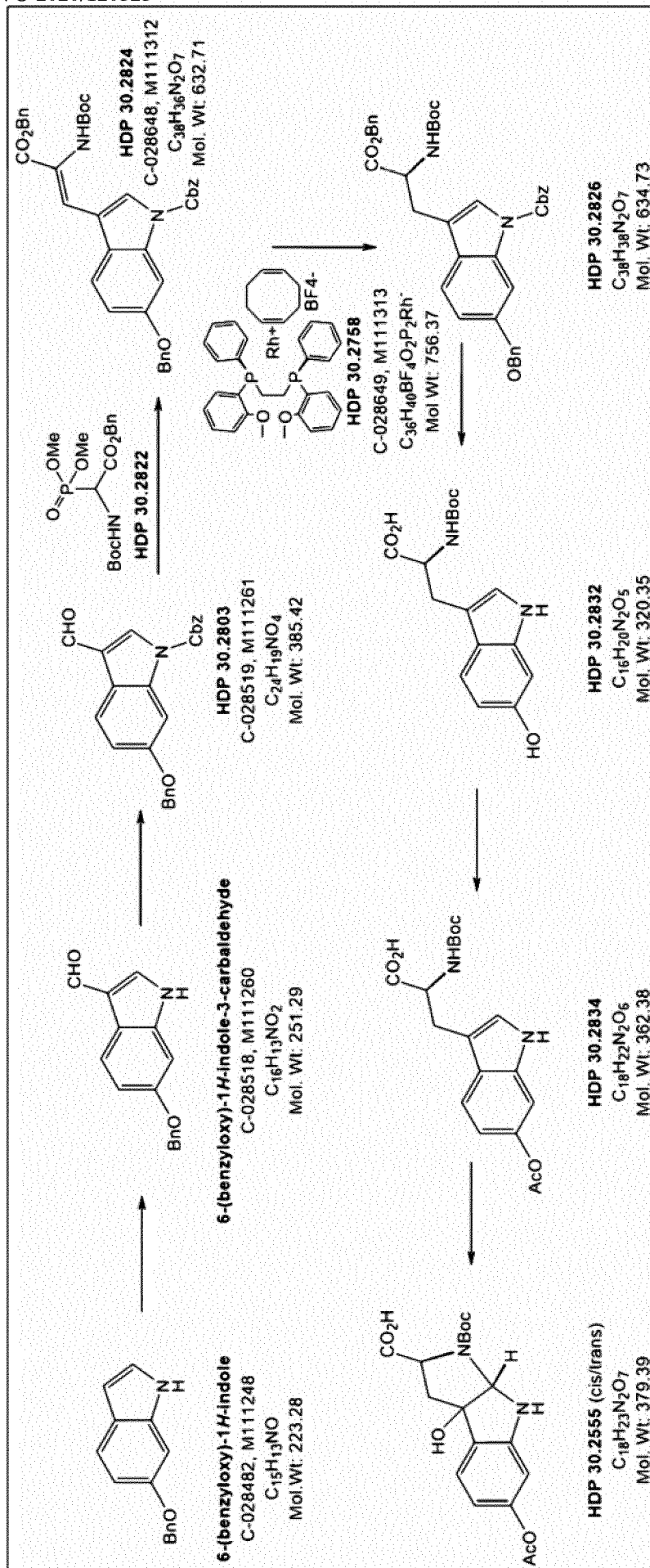


Fig. 4

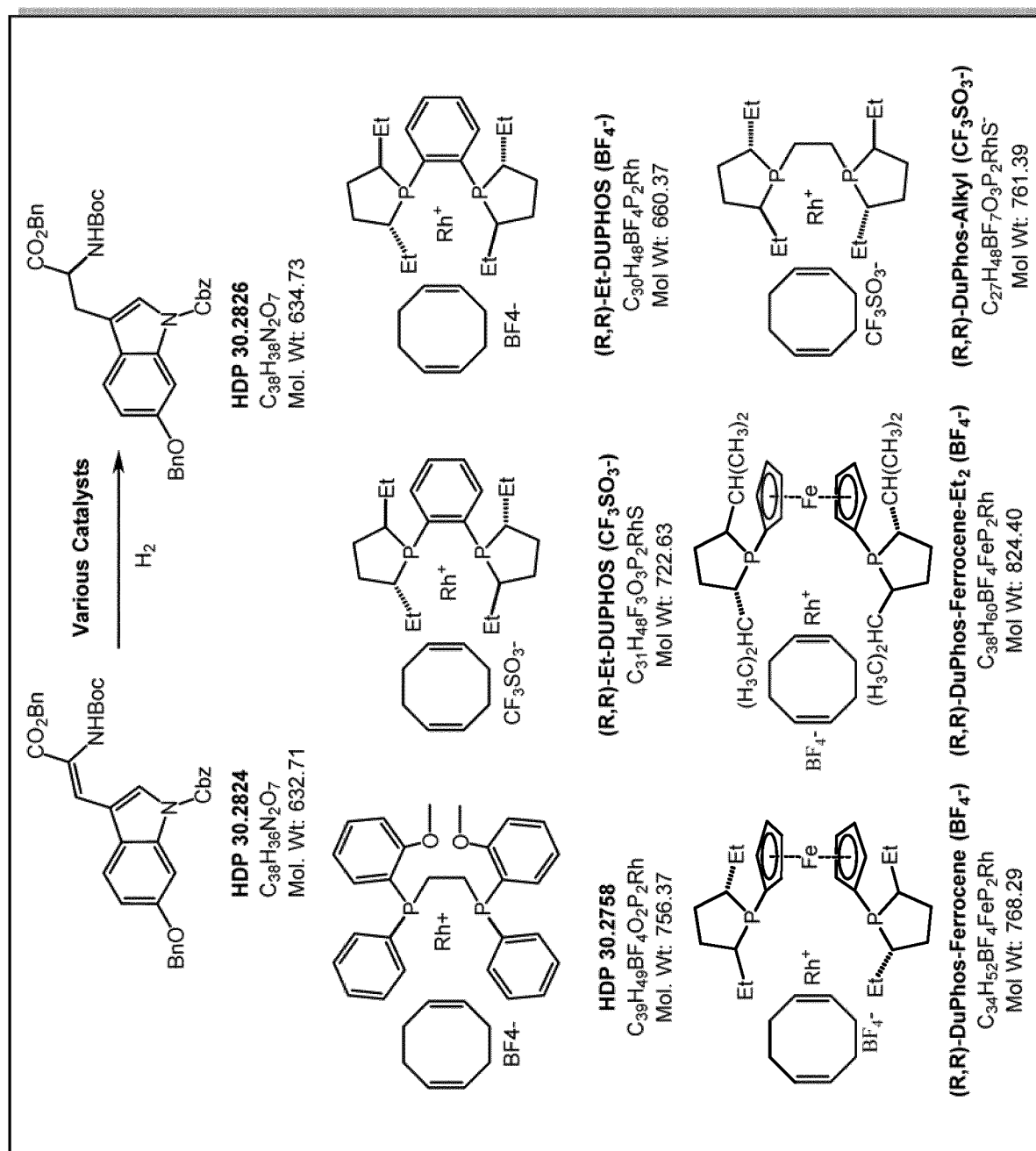


Fig. 5

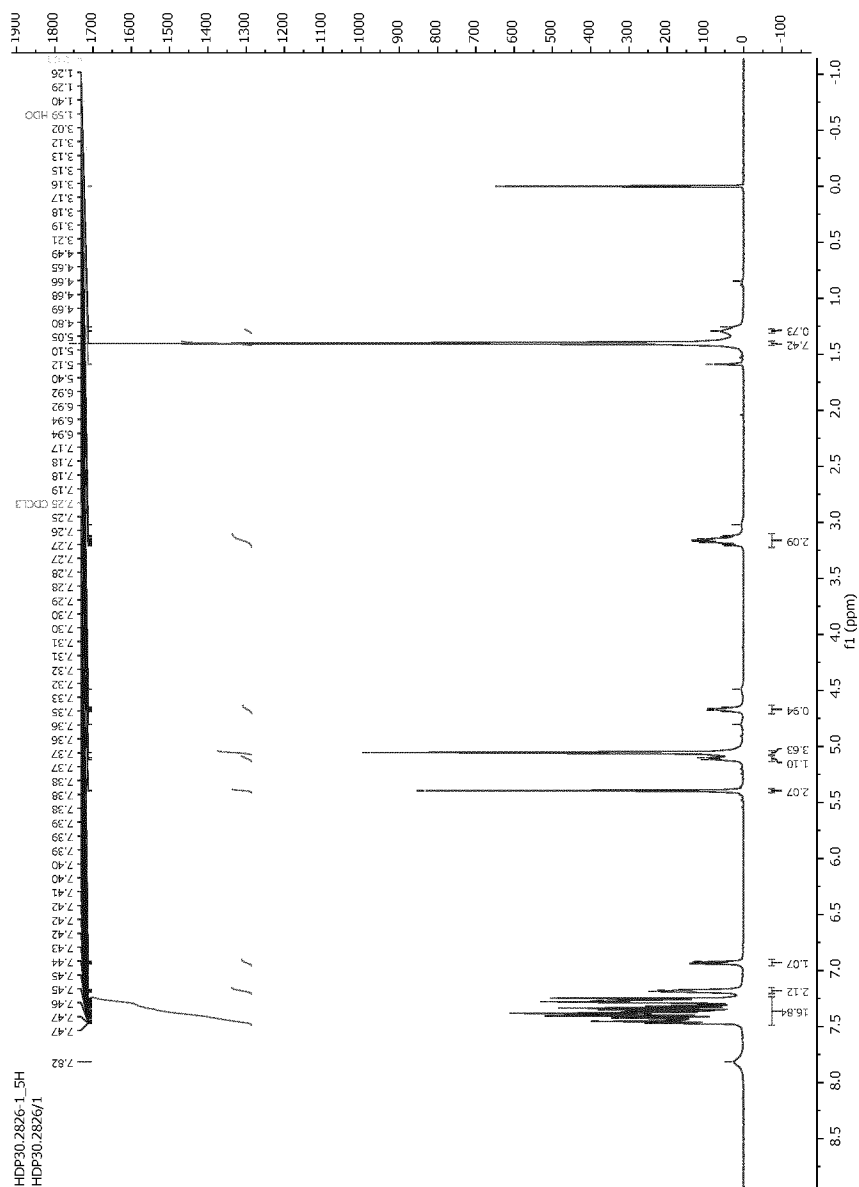


Fig. 6

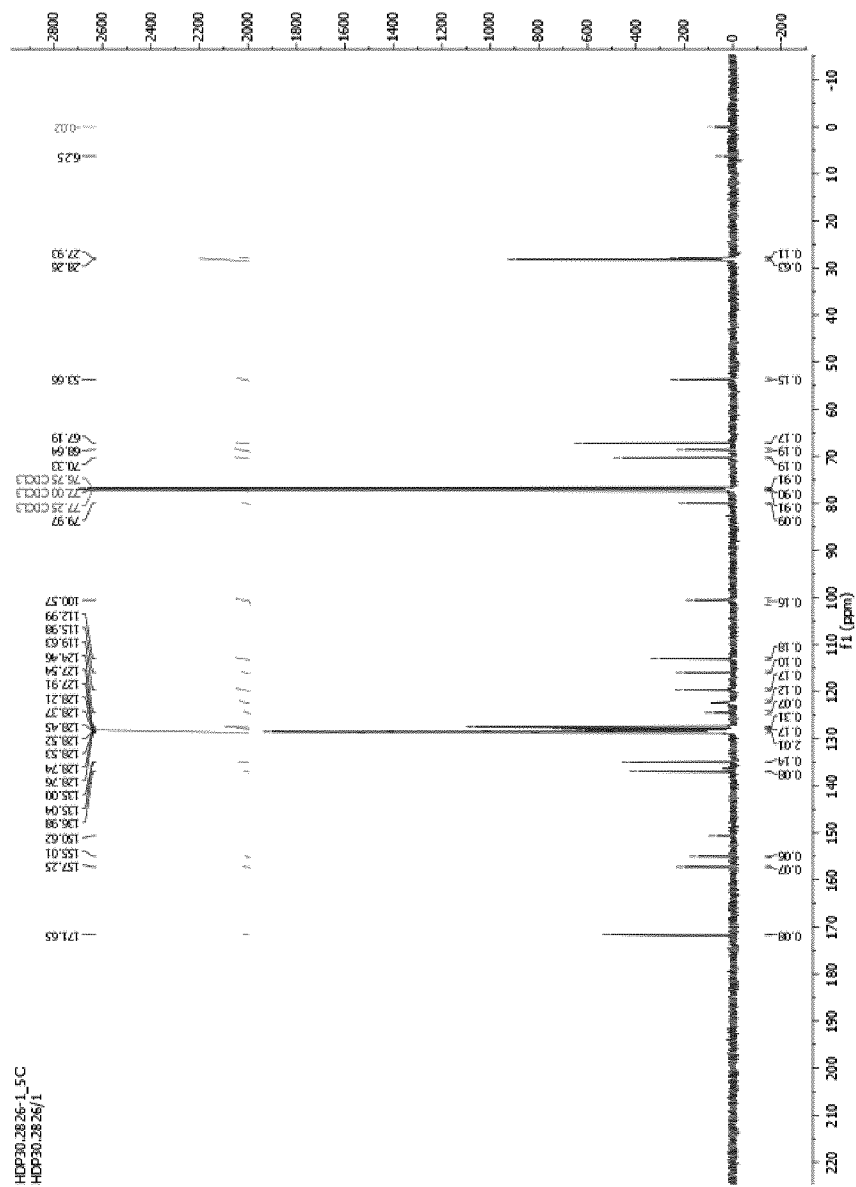


Fig. 7

INTERNATIONAL SEARCH REPORT

International application No

PCT/EP2019/084544

A. CLASSIFICATION OF SUBJECT MATTER

INV. C07D209/20

ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

C07D

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal, CHEM ABS Data, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X,P	WO 2019/030173 A1 (HEIDELBERG PHARMA GMBH [DE]) 14 February 2019 (2019-02-14) preparation of HDP30.2550 -----	10,11,13
X,P	WO 2018/237153 A1 (SQUIBB BRISTOL MYERS CO [US]) 27 December 2018 (2018-12-27) steps 1 to 4 in the preparation on p.75 -----	1-10,14, 15
Y	WO 2011/073173 A1 (HELMHOLTZ INFEKTIONSFORSCHUNG [DE] ET AL.) 23 June 2011 (2011-06-23) claims -----	1-10,14, 15
Y	EP 2 138 507 A1 (HELMHOLTZ INFEKTIONSFORSCHUNG [DE] ET AL.) 30 December 2009 (2009-12-30) examplesp.11-13; scheme 2, 3, 4; page 6 -----	1-10,14, 15
	-/--	



Further documents are listed in the continuation of Box C.



See patent family annex.

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"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

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"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

16 January 2020

Date of mailing of the international search report

28/01/2020

Name and mailing address of the ISA/

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040,
Fax: (+31-70) 340-3016

Authorized officer

Schuemacher, Anne

INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2019/084544

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	ARISTEIDIS CHIOTELLIS ET AL: "Synthesis and Biological Evaluation of 18 F-Labeled Fluoroethoxy Tryptophan Analogues as Potential PET Tumor Imaging Agents", MOLECULAR PHARMACEUTICS, vol. 11, no. 11, 14 July 2014 (2014-07-14), pages 3839-3851, XP055578046, US ISSN: 1543-8384, DOI: 10.1021/mp500312t scheme 2 -----	10
A	LI ET AL: "Synthesis of potent BCRP inhibitor-Kol43", TETRAHEDRON LETTERS, ELSEVIER, AMSTERDAM, NL, vol. 49, no. 9, 8 January 2008 (2008-01-08), pages 1480-1483, XP022439753, ISSN: 0040-4039, DOI: 10.1016/J.TETLET.2007.12.130 scheme 2; p.1480, right-hand column, second paragraph. -----	1-10,14, 15

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No

PCT/EP2019/084544

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 2019030173	A1	14-02-2019	NONE
WO 2018237153	A1	27-12-2018	NONE
WO 2011073173	A1	23-06-2011	AU 2010332988 A1 21-06-2012 CA 2783938 A1 23-06-2011 EP 2513016 A1 24-10-2012 ES 2564930 T3 30-03-2016 JP 5735977 B2 17-06-2015 JP 2013513643 A 22-04-2013 US 2012295941 A1 22-11-2012 WO 2011073173 A1 23-06-2011
EP 2138507	A1	30-12-2009	AT 557032 T 15-05-2012 AU 2009270571 A1 21-01-2010 CA 2729188 A1 21-01-2010 EP 2138507 A1 30-12-2009 EP 2303916 A1 06-04-2011 ES 2391220 T3 22-11-2012 JP 5634397 B2 03-12-2014 JP 2011525512 A 22-09-2011 US 2011311564 A1 22-12-2011 WO 2010006682 A1 21-01-2010