

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property

Organization

International Bureau

(43) International Publication Date

11 June 2020 (11.06.2020)



(10) International Publication Number

WO 2020/113330 A1

(51) International Patent Classification:

C02F 1/52 (2006.01) C02F 1/26 (2006.01)

B01J 20/22 (2006.01) C02F 1/28 (2006.01)

C02F 1/00 (2006.01)

MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

(21) International Application Number:

PCT/CA2019/051739

Published:

- with international search report (Art. 21(3))
- in black and white; the international application as filed contained color or greyscale and is available for download from PATENTSCOPE

(22) International Filing Date:

03 December 2019 (03.12.2019)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

62/774,822 03 December 2018 (03.12.2018) US

62/775,682 05 December 2018 (05.12.2018) US

62/775,696 05 December 2018 (05.12.2018) US

62/775,708 05 December 2018 (05.12.2018) US

(71) Applicant: CARBONET NANOTECHONOLOGIES

INC. [CA/CA]; 2366 Main Mall, Room 289, Vancouver, British Columbia V6T 1Z4 (CA).

(72) Inventors: CARLSON, Michael; c/o Carbonet Nanote-

chologies Inc., 2366 Main Mall, Room 289, Vancouver, British Columbia V6T 1Z4 (CA). CATALDO-HERNAN-DEZ, Macarena; 406-2336 York Ave., Vancouver, British Columbia V6K 1C7 (CA). CULOTTA, Anne Marie; 205 Shirleen Drive, Seabrook, Texas 77586 (US).

(74) Agent: C6 PATENT GROUP INCORPORATED (OP-ERATING AS CARBON PATENT GROUP); Carbon

Patent Group, 1-64 Holland St. W., P.O. Box 173, Bradford, Ontario L3Z 2A8 (CA).

(81) Designated States (unless otherwise indicated, for every

kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DJ, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IR, IS, JO, JP, KE, KG, KH, KN, KP, KR, KW, KZ, LA, LC, LK, LR, LS, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every

kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV,

(54) Title: NANONETS FOR REMOVAL OF CONTAMINANTS FROM AQUEOUS SOLUTIONS, KITS THEREFOR AND METHODS OF THEIR USE

(57) Abstract: Provided are nanonets comprising a) a surfactant aggregate having an average aggregate diameter; and b) a polymer having an average particle diameter which average particle diameter is the same or smaller than the average aggregate diameter, wherein the nanonet has a diameter larger than the average particle diameter. Also provided are kits therefor and methods for sequestering non-water moieties from aqueous solutions using nanonets.



WO 2020/113330 A1

NANONETS FOR REMOVAL OF CONTAMINANTS FROM AQUEOUS SOLUTIONS, KITS THEREFOR AND METHODS OF THEIR USE

TECHNICAL FIELD

This invention relates to the field of sequestering and more particularly in the field of sequestering material from aqueous environments.

BACKGROUND

The decontamination and disposal of organic waste materials such as sewage sludge, animal manure, food processing waste, fracking fluid, oil and gas, and other industrial wastewater presents both environmental and public health concerns. The handling and disposal of contaminated water has significant social, environmental, and economic implications. Currently, residual wastewater sludge is commonly digested, incinerated, deposited in landfills, returned to the same environment, or used as fertilizer through agricultural land application of the residual biosolids.

The encapsulation of inorganic particles with polymers has been demonstrated (E. Bourgeat-Lami and E. Duguet: Polymer encapsulation of inorganic particles; in Functional coatings, S.K. Ghosh (ed.); 2006, Wiley-VCH, Weinheim; Chapter 4, pp. 85-152). In biomedical applications, the coating of inorganic nanoparticles with a polymer, or their encapsulation in a polymer matrix or layer of detergent is important for properties such as enhanced solubility (R. Ladj *et al.* Polymer encapsulation of inorganic nanoparticles for biomedical applications; Internat. J. Pharm. 2013 (458) 230-241). In the fields of physics and engineering, magnetic fluids comprised of magnetic nanoparticles coated with a polymer and/or surfactant to stabilize the particle in the host liquid are studied. Various synthetic methods for the preparation of such systems have been published (e.g. J. Sommertune *et al.* Polymer/Iron oxide nanoparticles composites – a straight forward and scalable synthesis approach; Int. J. Mol. Sci. 2015 (16) 19752-19768).

SUMMARY

This invention is based, at least in part, on the elucidation that nanonet formation depends on surfactant average aggregate diameter, and the length of the surrounding polymer is determinative of the minimum surfactant aggregate diameter size for nanonet formation. Contrary to solubilization studies of lipid bilayers with styrene-maleic acid polymers, association of a surfactant with a polymer does not exclusively lead to a decrease in surfactant aggregate size. Rather, the combination of a polymer and a surfactant aggregate may result in an associated combination that is either larger than, or smaller than, a surfactant aggregate alone in solution. In the latter case, the polymer may cause relatively larger surfactant aggregates to dissociate into the smaller associated combination of surfactant aggregate and polymer. Further, utilizing a longer polymer length for nanonet formation may result in accelerated or delayed flocculation, which may be beneficial for removal of precipitated nanonets by physical separation or enhanced contact in a treated fluid. Consequently, a nanonet composed of a different molecular weight polymer may have advantages when applied to various wastewater treatment processes.

In illustrative embodiments, there is provided a kit for sequestering non-water moieties from an aqueous solution, the kit comprising: a) a surfactant aggregate having an average aggregate diameter; and b) a polymer having an average particle diameter which average particle diameter is the same or smaller than the average aggregate diameter; and c) a precipitation agent, provided that when the surfactant is selected from the group consisting of: Triton X-100TM, Triton X-305TM, N-dodecyl-beta-D-maltoside (DDM), sodium oleate, sodium cholate, sodium deoxycholate (DOC), beta-octyl glucoside (Beta-OG), lauryl dimethylamine n-oxide (LDAO), then the polymer is not selected from the group consisting of: SMA-2000TM SMA-2021TM, and Di-isobutylene-co-maleic acid (DIBMA).

In illustrative embodiments, there is provided a kit described herein wherein the precipitation agent is selected from the group consisting of: a cation

generator, a cation, an acid, a divalent cation, a trivalent cation and mixtures thereof.

In illustrative embodiments, there is provided a kit described herein wherein the precipitation agent is selected from the group consisting of: acetic acid, malic acid, citric acid, tartaric acid, fumaric acid, lactic acid, hydrochloric acid, sulfuric acid, and phosphoric acid.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant is selected from the group consisting of: anionic surfactants, nonionic surfactants, sugar-derived surfactants, glycidyl-derived surfactants, fatty acid alcohol-derived surfactants, nonionic surfactants, saccharide polyethyleneoxide combination surfactants, saccharide ester surfactants, sulfonated sugar based surfactants, aldonamide based surfactants, amide sugar based surfactants, amino alcohol surfactants, amino acid based surfactants, polyol surfactants, 1,2 glycol surfactants, and zwitterionic surfactants.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant is a nonionic surfactant.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant is an ionic surfactant.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant is an anionic surfactant.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant has a neutral charge.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant is an uncharged surfactant.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant is soluble in water.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant is insoluble in water.

In illustrative embodiments, there is provided a kit described herein wherein the polymer is a long polymer.

In illustrative embodiments, there is provided a kit described herein wherein the polymer has a molecular weight of at least 1,500 Da.

In illustrative embodiments, there is provided a kit described herein wherein the polymer has a molecular weight of at least 26,000 Da.

In illustrative embodiments, there is provided a kit described herein wherein the polymer has a molecular weight of at least 100,000 Da.

In illustrative embodiments, there is provided a kit described herein wherein the polymer has a molecular weight of at least 125,000 Da.

In illustrative embodiments, there is provided a kit described herein wherein the polymer is an amphipathic polymer.

In illustrative embodiments, there is provided a kit described herein wherein the polymer comprises an alkyl chain having at least 8 carbon atoms in length.

In illustrative embodiments, there is provided a kit described herein wherein the polymer is a block co-polymer.

In illustrative embodiments, there is provided a kit described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 4:1.

In illustrative embodiments, there is provided a kit described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 3:1.

In illustrative embodiments, there is provided a kit described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 2:1 to about 3:1.

In illustrative embodiments, there is provided a kit described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 2:1.

In illustrative embodiments, there is provided a kit described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 3:1.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant is selected from the group consisting of: octylphenol ethoxylates, nonylphenol ethoxylates, PGL1, PGL2, PGL3, PGL4, PGL5, Triton X-45TM, Triton X-100TM, Triton X-305TM, sodium oleate, LDAO, and mixtures thereof.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant is a mixture of PGL5 and sodium oleate.

In illustrative embodiments, there is provided a kit described herein wherein the surfactant is a mixture of PGL5 and potassium oleate.

In illustrative embodiments, there is provided a kit described herein wherein the polymer is selected from the group consisting of: SMA-725TM, SMA-2000TM, SMA-3000TM, SMA-2021TM, polymethacrylate, DIBMA, styrene carbamate block co-polymers, and styrene maleimide block co-polymers.

In illustrative embodiments, there is provided a kit described herein wherein the polymer is SMA-725TM.

In illustrative embodiments, there is provided a kit described herein wherein at least a portion of the polymer is hydrolyzed.

In illustrative embodiments, there is provided a nanonet comprising: a) a surfactant aggregate having an average aggregate diameter; and b) a polymer having an average particle diameter which average particle diameter is the same or smaller than the average aggregate diameter, provided that when the surfactant is selected from the group consisting of: Triton X-100TM, Triton X-305TM, N-dodecyl-beta-D-maltoside (DDM), sodium oleate, sodium cholate, sodium deoxycholate (DOC), beta-octyl glucoside (Beta-OG), lauryl dimethylamine n-oxide (LDAO), then the polymer is not selected from the group consisting of: SMA-2000TM, SMA-2021TM, and Di-isobutylene-co-maleic acid (DIBMA).

In illustrative embodiments, there is provided a nanonet described herein wherein the nanonet has a diameter larger than the average polymer particle diameter.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant is selected from the group consisting of: anionic surfactants, nonionic surfactants, sugar-derived surfactants, glycidyl-derived surfactants, fatty acid alcohol-derived surfactants, nonionic surfactants, saccharide polyethyleneoxide combination surfactants, saccharide ester surfactants, sulfonated sugar based surfactants, aldonamide based surfactants, amide sugar based surfactants, amino alcohol surfactants, amino acid based surfactants, polyol surfactants, 1,2 Glycol surfactants, and zwitterionic surfactants.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant is a nonionic surfactant.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant is an ionic surfactant.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant is an anionic surfactant.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant has a neutral charge.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant is an uncharged surfactant.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant is soluble in water.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant is insoluble in water.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer is a long polymer.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer has a molecular weight of at least 1,500 Da.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer has a molecular weight of at least 26,000 Da.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer has a molecular weight of at least 100,000 Da.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer has a molecular weight of at least 125,000 Da.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer is an amphipathic polymer.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer comprises an alkyl chain having at least 8 carbon atoms in length.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer is a block co-polymer.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 4:1.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 3:1.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 2:1 to about 3:1.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 2:1.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 3:1.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant is selected from the group consisting of: octylphenol ethoxylates, nonylphenol ethoxylates, PGL1, PGL2, PGL3, PGL4, PGL5, Triton X-45TM, Triton X-100TM, Triton X-305TM, sodium oleate, LDAO, and mixtures thereof.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant is a mixture of PGL5 and sodium oleate.

In illustrative embodiments, there is provided a nanonet described herein wherein the surfactant is a mixture of PGL5 and potassium oleate.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer is selected from the group consisting of: SMA-725TM, SMA-2000TM, SMA-3000TM, SMA-2021TM, polymethacrylate, DIBMA, styrene carbamate block co-polymers, and styrene maleimide block co-polymers.

In illustrative embodiments, there is provided a nanonet described herein wherein the polymer is SMA-725TM.

In illustrative embodiments, there is provided a nanonet described herein wherein at least a portion of the polymer is hydrolyzed.

In illustrative embodiments, there is provided a nanonet described herein further comprising a sequestered non-water moiety.

In illustrative embodiments, there is provided a nanonet described herein wherein the sequestered non-water moiety is an iron moiety.

In illustrative embodiments, there is provided a method of sequestering a non-water moiety from an aqueous solution, the method comprising: a) adding a surfactant aggregate having an average aggregate diameter to the aqueous solution comprising the non-water moiety; and b) adding a polymer having an average particle diameter which average particle diameter is the same or smaller than the average aggregate diameter to the aqueous solution comprising the non-water moiety, thereby forming a treated aqueous solution, provided that when the surfactant is selected from the group consisting of: Triton X-100TM, Triton X-305TM, N-dodecyl-beta-D-maltoside (DDM), sodium oleate, sodium cholate, sodium deoxycholate (DOC), beta-octyl glucoside (Beta-OG), lauryl dimethylamine n-oxide (LDAO), then the polymer is not selected from the group consisting of: SMA-2000TM, SMA-2021TM, and Di-isobutylene-co-maleic acid (DIBMA)

In illustrative embodiments, there is provided a method described herein further comprising adding a precipitation agent to the treated aqueous solution.

In illustrative embodiments, there is provided a method described herein further comprising mixing.

In illustrative embodiments, there is provided a method described herein further comprising removal of solid particles from the treated aqueous solution.

In illustrative embodiments, there is provided a method described herein wherein the removal of solid particles comprises at least one selected from the group consisting of: filtration, centrifugation, gravity separation, flotation, skimming, and electromagnetic attraction.

In illustrative embodiments, there is provided a method described herein wherein the surfactant is selected from the group consisting of: anionic surfactants, nonionic surfactants, sugar-derived surfactants, glycidyl-derived surfactants, fatty acid alcohol-derived surfactants, nonionic surfactants, saccharide polyethyleneoxide combination surfactant, saccharide ester surfactants, sulfonated sugar based surfactants, aldonamide based surfactants, amide sugar based surfactants, amino alcohol surfactants, amino acid based surfactants, polyol surfactants, 1,2 glycol surfactants, and zwitterionic surfactants.

In illustrative embodiments, there is provided a method described herein wherein the surfactant is a nonionic surfactant.

In illustrative embodiments, there is provided a method described herein wherein the surfactant is a anionic surfactant.

In illustrative embodiments, there is provided a method described herein wherein the surfactant has a neutral charge.

In illustrative embodiments, there is provided a method described herein wherein the surfactant is an uncharged surfactant.

In illustrative embodiments, there is provided a method described herein wherein the surfactant is soluble in water.

In illustrative embodiments, there is provided a method described herein wherein the surfactant is insoluble in water.

In illustrative embodiments, there is provided a method described herein wherein the polymer is a long polymer.

In illustrative embodiments, there is provided a method described herein wherein the polymer has a molecular weight of at least 1,500 Da.

In illustrative embodiments, there is provided a method described herein wherein the polymer has a molecular weight of at least 26,000 Da.

In illustrative embodiments, there is provided a method described herein wherein the polymer has a molecular weight of at least 100,000 Da.

In illustrative embodiments, there is provided a method described herein wherein the polymer has a molecular weight of at least 125,000 Da.

In illustrative embodiments, there is provided a method described herein wherein the polymer is an amphipathic polymer.

In illustrative embodiments, there is provided a method described herein wherein the polymer comprises an alkyl chain having at least 8 carbon atoms.

In illustrative embodiments, there is provided a method described herein wherein the polymer is a block co-polymer.

In illustrative embodiments, there is provided a method described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 4:1.

In illustrative embodiments, there is provided a method described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 3:1.

In illustrative embodiments, there is provided a method described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 2:1 to about 3:1.

In illustrative embodiments, there is provided a method described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 2:1.

In illustrative embodiments, there is provided a method described herein wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 3:1.

In illustrative embodiments, there is provided a method described herein wherein the surfactant is selected from the group consisting of: octylphenol

ethoxylates, nonylphenol ethoxylates, PGL1, PGL2, PGL3, PGL4, PGL5, Triton X-45TM, Triton X-100TM, Triton X-305TM, sodium oleate, LDAO, and mixtures thereof.

In illustrative embodiments, there is provided a method described herein wherein the surfactant is a mixture of PGL5 and sodium oleate.

In illustrative embodiments, there is provided a method described herein wherein the surfactant is a mixture of PGL5 and potassium oleate.

In illustrative embodiments, there is provided a method described herein wherein the polymer is selected from the group consisting of: SMA-725TM, SMA-2000TM, SMA-3000TM, SMA-2021TM, polymethacrylate, DIBMA, styrene carbamate block co-polymers, and styrene maleimide block co-polymers.

In illustrative embodiments, there is provided a method described herein wherein the polymer is SMA-725TM.

In illustrative embodiments, there is provided a method described herein wherein at least a portion of the polymer is hydrolyzed.

In illustrative embodiments, there is provided a method of sequestering a non-water moiety from an aqueous solution, the method comprising adding a nanonet described herein to the aqueous solution comprising a non-water moiety, thereby forming a treated aqueous solution.

In illustrative embodiments, there is provided a method described herein further comprising adding a precipitation agent to the treated aqueous solution.

In illustrative embodiments, there is provided a method described herein further comprising mixing.

In illustrative embodiments, there is provided a method described herein further comprising removal of solid particles from the treated aqueous solution.

In illustrative embodiments, there is provided a method described herein wherein the removal of solid particles comprises at least one selected from the group consisting of: filtration, centrifugation, gravity separation, flotation, skimming, and electromagnetic attraction.

Other aspects and features of the present invention will become apparent to those ordinarily skilled in the art upon review of the following description of specific embodiments of the invention in conjunction with the accompanying figures.

BRIEF DESCRIPTION OF THE DRAWINGS

In drawings which illustrate aspects of the invention,

Figure 1: Structure function relationship between polymer length and surfactant aggregate diameter in nanonet formation. A) Nanonet formation when polymer length and surfactant aggregate size are well matched. B) Nanonet formation does not occur when surfactant aggregate diameter is less than compact polymer particle diameter. C) Nanonet formation when a large surfactant aggregate diameter is broken down into multiple, smaller nanonets. D) Nanonet formation when a large surfactant aggregate is stabilized by multiple polymers, leading to an overall increase in particle size.

Figure 2: Co-precipitation of complexed polymer and surfactant due to addition of acid. A) Absorbance of acid precipitated nanonets formed from varying ratios of Triton X-305TM surfactant. Absorbance was measured at 540nm as a proxy for floc scattered light. Nanonets were formed with long chain polymer (SMA-725TM, triangles), short chain polymer (SMA-2000TM, squares), and compared to a surfactant only control (control, circles). B) As in A, with Triton X-100TM as surfactant. C) As in A, with Triton X-45TM as surfactant.

Figure 3: SMA-725TM enhances flocculation speed of nanonets. A) Distribution assay schematic. Precipitated nanonets flocculate and float with time, the total floc mass consolidates to the indicated volume of the tube (Top, top-middle, bottom-middle, and bottom: 100-75%, 75-50%, 50-25%, and 0-25%, respectively) B) Distribution of precipitated sodium oleate over time. C) Distribution of precipitated sodium oleate/ SMA-725TM nanonets over time. D) Distribution of precipitated sodium oleate/ SMA-2000TM nanonets over time. E) Solution clearing of precipitated sodium oleate (triangles pointing to the top of the page), sodium oleate and SMA-725TM added sequentially (triangles pointing to

the bottom of the page), sodium oleate/ SMA-725TM nanonets (circles), and sodium oleate SMA-2000TM nanonets (squares). Solution clearing was measured by absorbance at 540nm.

Figure 4: Dissolved Iron removal for nanonets in varying concentrations of divalent cations. SMA-2000TM nanonets formed with the indicated surfactant at a 5:1 ratio (surfactant:polymer) were incubated with iron solutions overnight, then filtered and iron contents quantified. As a control, iron solutions were filtered after shifting solution pH or without treatment.

Figure 5: Dissolved Iron removal for nanonets in varying concentrations of divalent cations. SMA-2000TM nanonets formed with the indicated surfactant at a 5:1 ratio (surfactant:polymer) were incubated with iron solutions for 10 minutes, then filtered and iron contents quantified. As a control, iron solutions were filtered after shifting solution pH or without treatment.

Figure 6: Iron removal efficiencies with SMA-2000TM and SMA-725TM nanonets in varying buffer concentrations.

Figure 7: Removal of iron from 100ppm iron solutions (75:25 Fe³⁺:Fe²⁺) in SMA-725TM nanonets

Figure 8: Graph showing the effect polymer and surfactant:polymer ratio on iron removal in sodium oleate nanonets.

Figure 9: Effect of various nanonets treatment on produced water samples (500ppm oil, >100,000 TDS, >1,000 ppm divalent cations).

Figure 10: Post filtration quantification of treated produced water. Oil and gas (O and G), total suspended solids (TSS), and total iron were quantified after filtration of the samples in Figure 9.

Figure 11: Contaminant removal from produced water with nanonets OPGW (SMA-725TM, 75% sodium oleate, 25% polyglyceryl-10 monostearate, 1:1 wt ratio polymer:surfactant; 10 min residence time, after filtration). A) Iron removal B) Colour removal and C) Turbidity removal.

Figure 12: Contaminant removal of produced water with nanonet OPGW after dissolved air flotation, followed by filtration. A) Total iron

concentration pre- and post-treatment. Total iron concentration was determined by the Ferrozine method. B) Water colour pre- and post-treatment. Yellow colour indicative of dissolved hydrocarbons and colloidal iron was measured by absorbance at 350nm. C) Turbidity pre- and post-treatment. Turbidity was measured by absorbance at 540nm.

Figure 13: A) Removal of extractable petroleum hydrocarbons with chain length 10-32 carbons wherein “Start” is before treatment, “Filtered” is treatment without nanonets and filtration, and “Filtered+Nanonets” is treatment with both nanonets and filtration. B) Removal of Zn and Fe from produced water. C) Removal of various metals from produced water.

Figure 14: Illustration of monomeric units of a polymer in hydrated and un-hydrated forms.

Figure 15: Nanonet formation with model surfactants a) Triton X-100™ and b) Triton X-45™ and styrene maleic acid block co-polymers.

Figure 16: Evolution of quantitative scoring method for nanonet formation.

Figure 17: Nanonet score for each SMA polymer combination.

Figure 18: Nanonet scores for sulfonates, lactylates and trideceth sulfonate with differing block-copolymers.

Figure 19: Nanonet formation scores with SMA polymers with DIBMA.

Figure 20: Nanonet scores with differing block-copolymers over differing HLB values.

Figure 21: Effect of alkyl chain and hydrophilic headgroup length on Nanonet formation with varying surfactant head groups. Effect of alkyl chain length on nanonet scores for surfactants with A) amino acid head groups (N-dodecanoyl methionine, N-dodecanoyl threonine, N-decanoyl methionine, N-decanoyl threonine, N-octanoyl methionine, and N-octanoyl threonine in order from top to bottom), B) Sodium Sulfonate head groups (Na-dodecylbenzene sulfonate, Na-dodecane sulfonate, 1-octane sulfonate, p-toluene-1-sulfonate, and Na-pentane sulfonates in order from top to bottom) C) ethylene glycol ether head groups (hexaethylene glycol monododecyl ether, heptaethylene glycol

monododecyl ether, hexaethylene glycol monodecyl ether, and hexaethylene glycol mono-octyl in order from top to bottom). D) Thioglucosides (n-dodecyl- β -D-thioglucopyranoside, n-dodecyl- β -D-thiomaltopyranoside, n-decyl- β -D-thioglucopyranoside, n-decyl- β -D-thiomaltopyranoside, n-nonyl- β -D-thioglucopyranoside, n-octyl-maltoside, fluorinated, n-octyl- β -D-thiomaltopyranoside, and n-heptyl- β -D-thioglucopyranoside, in order from top to bottom). Effect of hydrophilic head group size (represented by HLB) on nanonet scores for surfactants with E) polyethoxylated sorbitan headgroups (SPAN 20TM, TWEEN 65TM, TWEEN 85TM, TWEENTM 80, TWEEN 60TM, TWEEN 40TM, and TWEEN 20TM in order of top to bottom). F) Phenyl polyethylene glycol headgroups (TRITON X-45TM, TRITON X-100TM, and TRITON X-305TM).

Figure 22: Nanonet library with anionic surfactants employed to screen Cu²⁺ metal ion removal from seawater.

Figure 23: Nanonet library with non-ionic surfactants used to screen Cu²⁺ metal ion removal from seawater.

Figure 24: Nanonet library with anionic surfactants utilized to screen Zn²⁺ metal ion removal from seawater.

Figure 25: Nanonet library with non-ionic surfactants applied to screen Zn²⁺ metal ion removal from seawater.

Figure 26: Example batch settling curve of a SMA-725TM and BIOSOFT D-40 mixture added to a calcium solution. Floc sedimentation is quantified over time and a sigmoid function is used to fit the data.

Figure 27: Bar graph quantifying apparent settling speed (t_{50}) for three different polymers. Error bars are shown in black.

Figure 28: Selected water parameters from treated and untreated produced water samples. A). Iron levels pre- and post-treatment. B) Turbidity levels pre- and post-treatment C). Oxidative Reductive Potential pre- and post-treatment D). Total dissolved solids pre- and post-treatment E) Bicarbonate levels pre- and post-treatment.

Figure 29: Bar graphs illustrating (A) iron removal from produced water using various concentration of nanonets and (B) residual ClO₂.

DETAILED DESCRIPTION

Nanonets described herein provide for efficient removal of a targeted contaminant by selecting a suitable polymer length and polymer amount based on the surfactant that is most suitable for removal of the targeted contaminant. Virtually any surfactant aggregate may be used in a nanonet provided that a suitable polymer is selected. Neutralization of the polymer provides for precipitation of the nanonets and associated sequestered contaminants. This is advantageous as a solid precipitate comprised of nanonets and associated sequestered contaminants can be easily removed by physical methods of separation. Non-limiting examples of physical separation methods include dissolved air flotation (DAF), settling, filtration, centrifugation, gravity separation, flotation, skimming, electromagnetic attraction or repulsion, microfiltration and nanofiltration. The relationship between polymer size and surfactant aggregate structure allows nanonet formulations to be designed that target select contaminants that fit into common physical separation systems.

Surfactants are typically amphipathic molecules with a hydrophilic head group and a lipophilic/hydrophobic functional group. Depending on the size and charge of the hydrophobic to hydrophilic headgroups, surfactants can be soluble or insoluble in water. This balance is typically called the hydrophilic lipophilic balance (HLB) (Griffin, William C. (1949), "Classification of Surface-Active Agents by 'HLB'", Journal of the Society of Cosmetic Chemists, 1 (5): 311–26), as the head group typically has an affinity for water and the tail group has an affinity for lipids and oils. Surfactants can be insoluble in water, where they are typically utilized to prepare water in oil emulsions and have an HLB value <10 , or soluble in water, where they are typically utilized to prepare oil in water emulsions and have an HLB value >10 . Soluble surfactants typically form Gibbs monolayers in aqueous solutions, where free surfactant is able to exchange from the bulk water phase to the air water interface. In contrast, insoluble surfactants form Langmuir monolayers which are compressible, and do not exchange, or only sparingly, with the bulk water phase (Rogalska, E., *et al.*, (2000). *Formation and properties of*

Langmuir and Gibbs monolayers: a comparative study using hydrogenated and partially fluorinated amphiphilic derivatives of mannitol. Chemistry and Physics of Lipids, 105(1), 71–91. doi:10.1016/s0009-3084(99)00130-, 9). Examples of lipids include phosphatidylcholine, phosphatidylethanolamine, phosphatidylglycerol, and lecithins. Examples of soluble surfactants include Triton X-100™, Sodium dodecyl sulfate, and polyglycerol-10-laurate, which all have an HLB >10. Examples of insoluble surfactants include SPAN 80™ or TWEEN 85™.

As used herein the term “nanonet” refers to a particle that is formed by an association between a polymer and surfactant aggregate. The nanonet self-assembles in an aqueous environment, is stable in aqueous solution, and is comprised of i) a polymer and ii) a surfactant aggregate. Nanonets remain associated at lower concentrations relative to surfactant aggregates in the absence of the polymer. The solution stability of nanonets may be disrupted by the addition of a suitable destabilization material. For example, and without limitation, neutralization of charged carboxylic acid groups by an acid may cause destabilization of a nanonet which comprises a maleic acid component in the polymer. Without being limited by theory, it is believed that nanonets are the result of interactions between the alkyl chains of the surfactants and the alkyl chains of the hydrophobic portions of the polymer. Often, nanonets are colloidal particles comprising amphipathic block co-polymers and surfactants. The amphipathic block co-polymers often comprise a hydrophilic functional group and a hydrophobic functional group. Examples of the hydrophilic functional group include, but are not limited to, a maleic acid, a carboxylic acid and an amide linked ethanol group. Examples of the hydrophobic functional group include, but are not limited to, a styrene, di-isobutyl.

As used herein “nanonet diameter” refers to the size of a nanonet in an aqueous environment as measured by dynamic light scattering. An “average nanonet diameter” refers to the average of the size of a plurality nanonets in an aqueous environment as measured by dynamic light scattering. The term “average nanonet diameter” may be used interchangeably herein with the term “nanonet diameter” or “nanonet size”. Examples, without limitation, of how to

measure an average nanonet diameter are provided in the Examples section herein.

As used herein, the term “aqueous solution” refers to a liquid environment in which water is a major component. Examples of aqueous solutions include, but are not limited to, waste water, aqueous material recovered from a process, (such as sewage sludge, animal manure, food processing waste), oil and gas wastewater, used fracking fluid, industrial effluent, ground water and the like.

As used herein, the term "precipitation agent" refers to any substance which is cationic and/or generates a cation upon exposure to an aqueous environment. A substance which is cationic is a cation and a substance that generates a cation upon exposure to an aqueous environment is termed herein "a cation generator". Examples, without limitation, of precipitation agents include, an acid, a divalent cation, a trivalent cation, acetic acid, malic acid, citric acid, tartaric acid, fumaric acid, lactic acid, hydrochloric acid, sulfuric acid, phosphoric acid, and mixtures thereof.

As used herein, the term “surfactant” refers to any surface active agent containing a polar portion that thermodynamically prefers to be solvated by a polar solvent, and a non-polar portion that thermodynamically prefers to be solvated by a non-polar solvent. The term “surfactant” is also meant to encompass anionic, cationic, non-ionic and zwitterionic surfactants. Surfactants suitable for use in the present invention form aggregates in aqueous solution. Surfactants that do not form aggregates are not encompassed by the term “surfactant” as used herein.

An “anionic surfactant” is a surfactant with a polar portion that ionizes to form an anion in aqueous solution (the anion residing on the polar portion connected to the non-polar portion). A “cationic surfactant” refers to a surfactant having a cationic polar portion that ionizes to form a cation in aqueous solution (the cation residing on the polar portion connected to the non-polar portion). A “non-ionic” surfactant refers to a surfactant having a polar portion that does not ionize in aqueous solution. A “zwitterionic surfactant” refers to a surfactant that has separate cationic and anionic sites and is uncharged (for example, non-

limiting examples of zwitterions include dialkylammonio acetate as the zwitterion form of N,N-dialkylammonium acetic acid, or betaine dialkylsulfonio acetate). (from pg. 224-25, *The Vocabulary of Organic Chemistry*, Milton Orchin *et al.*, Eds., John Wiley & Sons, 1980). A soap is a salt of an anionic surfactant, typically a fatty acid.

A person of skill in the art familiar with surfactants will appreciate the vast array of suitable surfactants for use in the present invention, see for example and without limitation the following references: Drew Myers, *Surfactant Science and Technology*, VCH Publishers, Inc. 1988.; Bordes, Romain, and Krister Holmberg. 2015. "Amino Acid-Based Surfactants – Do They Deserve More Attention?" *Advances in Colloid and Interface Science* 222 (August): 79–91.; *Essentials of Glycobiology*. 3rd edition Varki A, Cummings RD, Esko JD, et al., editors. Cold Spring Harbor (NY): Cold Spring Harbor Laboratory Press; 2015-2017; Reference for definition: Zhang, X., Song, F., Taxipalati, M., Wei, W., & Feng, F. (2014). Comparative Study of Surface-Active Properties and Antimicrobial Activities of Disaccharide Monoesters. *PLoS ONE*, 9(12), e114845. doi:10.1371/journal.pone.0114845; R. H. Beck, N. J. Kalff, and H. W. W. Roper, European Patent Application, EP 536939, 1993; V. Au, B. Harichian, A. Hung, and R. Vermeer, US 5872111 A, 1999; J. J. Scheibel, D. S. Connor, R. E. Shumate, and L. J. Charles, PCT Int Appl. WO 92 06984, 1992; C. L. Mehlretter, R. L. Mellies, and J. C. Rankin, US patent 2670345, 1954; Z. Wang. United States patent US 7655611 B2, 2010; and Sari-Chmayssem N., Pessel F., Guégan J. P., S. Taha, Mawlawi H., and Benvegnu T. Direct and one-pot conversion of polyguluronates and alginates into alkyl-L-gulonamide-based surfactant compositions, *Green Chemistry*; Novel surfactants and novel emulsifiers made from disaccharides. WO 2006/100091; US 6,184,196 B1. SUCROSE BASED SURFACTANTS AND METHODS THEREOF; WO 94/12511; and Safety Assessment of Alkyl Sultaines as Used in Cosmetics. Christine Burnett. *Cosmetic Ingredient Review*, June 30, 2017.

Nanonet formation begins with surfactants operable to form an aggregate that have an average aggregate diameter that exceeds a threshold average

particle size that may be determined based on the average particle size of the polymer for use in the nanonet composition. The surfactants set forth below may be used in embodiments of nanonets described herein. In some cases, surfactants may be described according to their polar group, and their non-polar group. Common linkages between a polar component and a non-polar component of a surfactant include, but are not limited to, ether, amine, ester, phosphate, thioester and amide linkers.

Anionic Surfactants

Sulfonate polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R-(SO_3M)_z$.

Sulfate polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R-(OSO_3M)_z$.

Carboxylate polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R-(COOM)_z$.

Phosphate polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R-(OPO_3M)_z$.

Phosphonate polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R-(PO_3M)_z$.

Alpha-sulfonated carboxylic acid polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R-(CH(SO_3M)COOM)_z$. R may be any one or more of lauric, palmitic and stearic acids.

Alpha-sulfonated carboxylic acid ester polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R-(CH(SO_3M)COOR^1)_z$. R may be any one or more of lauric, palmitic and stearic acids. R^1 is C_1 - C_5 alkyl linear or branched. Z may be 1, 2, 3 or any combination thereof and the non-R group may reside on the same carbon in R or on different carbons.

Sulfonate ester polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R(COX(CH_2)_nSO_3M)_z$ (n may be in the range of 2 and 8 and X is O, NH or $N-R^1$).

Arylamide sulfonate ester polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R(COXArSO_3M)_z$.

Xanthate polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R(OCS_2M)_z$.

In any of the above formulae, M may be any monovalent cation including, for example, Na^+ , K^+ , Li^+ .

Nonionic Surfactants

Polyoxyethylene polar groups may be attached to a non-polar group to make a surfactant of the general formula: $R(X(CH_2CH_2O)_nCH_2CH_2OH)_z$ (X is O, S or NH).

Polyoxypropylene polar groups may be attached to a non-polar group to make a surfactant of the general formula:

$R(X(CH_2CH(CH_3)O)_nCH_2CH(CH_3)OH)_z$ (X is O, S or NH).

Bis – Polyoxyethylene amine polar groups may be attached to a non-polar group to make a surfactant of the general formula:

$(RX((CH_2CH_2O)_nCH_2CH_2OH)_2)$ (X is N)

Bis – Polyoxypropylene amine polar groups may be attached to a non-polar group to make a surfactant of the general formula:

$(RX((CH_2CH(CH_3)O)_nCH_2CH(CH_3)OH)_2)$ (X is N)

In the chemical formulae represented above, R is a linear or branched chain, saturated, mono-unsaturated or poly-unsaturated, aliphatic cyclic, or aromatic cyclic, or polycyclic, and all possible combinations thereof. In some embodiments, the aromatic or non-aromatic cyclic is alkylated. In some embodiments, R may additionally be comprised of O, N, S, and combinations thereof, and additionally be functionalized with any one or more moieties such as carboxylic acid, carboxylic amide, thio-carboxylic acid and combinations thereof. Generally, R has as few as 8 and as many as 40 carbon atoms. In some embodiments R has as few as 7 or more carbon atoms. In some embodiments, R has from 7 to 40 carbon atoms. In some embodiments, R has from 8 to 40 carbon atoms. In some embodiments, R has from 7 to 20 carbon atoms, or R has from 8 to 20 carbon atoms, or R has from 7 to 20 carbon atoms. In some

embodiments, R has from 7 to 18 carbon atoms, or R has from 8 to 18 carbon atoms, or R has from 9 to 18 carbon atoms.

Where a chemical formula above includes the “n” parameter, unless some other value of n is expressed next to a particular chemical formula, n may have the following values: In some embodiments, n may be in the range of 4 and 20, or in the range of 5 and 18, or in the range of 6 and 17, or in the range of 7 and 16, or in the range of 8 and 15.

For a chemical formula above including the “z” parameter, z may be 1, 2, 3 or any combination thereof and the non-R group may reside on the same carbon in R or on different carbons of the R group.

For a chemical formula above including the parameter “Ar” in the chemical formula, Ar may be benzene, *p*-toluene, or naphthalene.

In some embodiments of a surfactant represented by the above chemical formulae, R may be cholic acid or a derivative thereof. In some embodiments, R may be any one or more of lauric, palmitic and stearic acids. In some embodiments, R may include up to five pendant acetic acid functional groups. In other embodiments, R may be cocoyl. In some embodiments, R may be derived from a fatty acid or fatty acid ester.

Embodiments employing surfactants having one of the nonionic polar groups described above employ combinations of alkyl groups that may optimize surfactant average aggregate size in relation to the polymer average particle size to be employed in the nanonet composition and further considering the contribution of a hetero-atom containing portion of the structures.

Sugar, Glycidyl, and Fatty acid alcohol derived Nonionic Surfactants

Non-limiting examples of surfactants under this category include: polyethylene glycol derivatives of fatty acids, such as, PEG derivatives of the amines of fatty acids. Examples of specific surfactants under this category include, but are not limited to: PEG-2 cocamine, PEG-3 cocamine, PEG-4 cocamine, PEG-5 cocamine, PEG-8 cocamine, PEG-10 cocamine, PEG-12 cocamine, PEG-15 cocamine, PEG-20 cocamine, PEG-2 hydrogenated tallow amine, PEG-5 hydrogenated tallow amine, PEG-8 hydrogenated tallow amine,

PEG-10 hydrogenated tallow amine, PEG-15 hydrogenated tallow amine, PEG-20 hydrogenated tallow amine, PEG-30 hydrogenated tallow amine, PEG-40 hydrogenated tallow amine, PEG-50 hydrogenated tallow amine, PEG-2 lauramine, PEG-2 oleamine, PEG-5 oleamine, PEG-6 oleamine, PEG-10 oleamine, PEG-15 oleamine, PEG-20 oleamine, PEG-25 oleamine, PEG-30 oleamine, PEG-12 palmitamine, PEG-2 rapeseedamine, PEG-2 soyamine, PEG-5 soyamine, PEG-8 soyamine, PEG-10 soyamine, and PEG-15 soyamine.

Sugar based functional groups

Acidic Sugars: Sugars that have had one or more hydroxyl groups substituted with a carboxylic acid. Non-limiting examples of acidic sugars include: Gluconic acid, and ascorbic acid

Deoxy Sugars: Sugar that has a hydroxyl group substituted for a hydrogen. Non-limiting examples of deoxy sugars: Rhamnose, Fucose, and Deoxyribose

C5-Monosaccharides (Furanoses): 5 carbon backbone saccharides, they will cyclize to form a 5 carbon ring. Non-limiting examples of C5 monosaccharides include: Fructose, ribose, xylose, arabinose, and lyose.

C6- Monosaccharides (Pyranoses): 6 carbon backbone saccharides, they will cyclize to form a 6 carbon ring. Non-limiting examples of C6 monosaccharides include: Glucose, mannose, allose, altrose, tallose, galactose, glucopyranose

Aminosugars: A sugar molecule in which a hydroxyl group has been replaced with an amine group. The functional group is most likely connected via an amide bond with the fatty acid, but those skilled in the art can also connect the amino sugar through one of the hydroxyl groups. Non-limiting examples of aminosugars include: Galactosamine, glucosamine, sialic acids, N-acetyl-D-glucosamine, mannosamine

Aldonamide: a sugar-based surfactant category. A non-limiting example of an aldonamide based surfactant includes: coco D-gluconamide.

Dissaccharides: Disaccharides are sugars or carbohydrates made by linking two monosaccharides. A glycosidic bond can form between any hydroxyl

group on the monosaccharides, leading to many different combinations of bonds and stereochemistry, producing disaccharides with unique properties. These disaccharides can be subsequently reacted with a free fatty acid by those with skill in the art to form a disaccharide alkyl ester. Non-limiting examples of Disaccharides include: Sucrose, maltose, lactose, cellobiose, mannobiose, trehalose, lactulose, turanose, xylobiose, melibiose, sophorose, and raffinose

Polysaccharides: A chain of more than 2 saccharides, linked together by glycosidic bonds. A non-limiting example of a group of polysaccharides is alkyl polyglucosides (APG).

Sugar Alcohols: Sugars which have had the terminal sugar aldehyde converted into an alcohol. Non-limiting examples of sugar alcohols include: lactitol, sorbitol, isomalt, xylitol, erythritol, and mannitol.

Dehydrated Sugar Alcohols: Sugar alcohols which have been dehydrated to form a cyclized lactone ring. Non limiting examples of dehydrated sugar alcohols include: sorbitan, mannitan, xylitan, anhydroerythritol, furanoside, and furanose.

Methods of addition of alkyl chains and sugars to make sugar based surfactants:

Glycosidic bond: Sugars can be attached to alkyl chains through a glycosidic bond. The glycosidic bond can be N, O or S linked, whereby a hydroxyl group on the sugar is replaced and linked to another carbon chain via an amine, ether, or thioether. All of the sugar functional groups mentioned above can be used to form a glycosidic bond with an alkyl chain. Non-limiting examples of sugar derived surfactants include: Cocoyl glucoside (O-linked), octyl-glucamine (N-linked), alkylpolyglucosides, 3-(butyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(pentyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(hexyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(heptyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(octyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(nonyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(decyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(dodecyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(tetradecyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(hexadecyloxy)-2-hydroxypropyl-beta-D-

galactopyranoside, 3-(octadecyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(eicosyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(docosyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(tetracosyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(hexenyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(decenyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(dodecenyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(tetradecenyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(hexadecenyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(octadecenyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(docosenyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(tetracosenyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(3-oxatridecyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(fluorododecyloxy)-2-hydroxypropyl-beta-D-galactopyranoside, 3-(butyloxy)-2-hydroxypropyl-beta-D-glucopyranoside, 3-(octyloxy)-2-hydroxypropyl-beta-D-mannopyranoside, 3-(tetradecyloxy)-2-hydroxypropyl-beta-D-lactoside, 3-(octadecyloxy)-2-hydroxypropyl-beta-D-maltoside, 3-(octyloxy)-2-hydroxypropyl-beta-D-galactotrioside, and 3-(dodecyloxy)-2-hydroxypropyl-beta-D-cellobioside.

Saccharide polyethyleneoxide combination surfactants:

Non-limiting examples of surfactants in this category include: PEG-20 sorbitan cocoate, PEG-40 sorbitan diisostearate, PEG-2 sorbitan isostearate, PEG-5 sorbitan isostearate, PEG-20 sorbitan isostearate, PEG-40 sorbitan lanolate, PEG-75 sorbitan lanolate, PEG-10 sorbitan laurate, PEG-40 sorbitan laurate, PEG-44 sorbitan laurate, PEG-75 sorbitan laurate, PEG-80 sorbitan laurate, PEG-3 sorbitan oleate, PEG-6 sorbitan oleate, PEG-80 sorbitan palmitate, PEG-40 sorbitan perisostearate, PEG-40 sorbitan peroleate, PEG-3 sorbitan stearate, PEG-6 sorbitan stearate, PEG-40 sorbitan stearate, PEG-60 sorbitan stearate, PEG-20 sorbitan tetraoleate, PEG-30 sorbitan tetraoleate, PEG-40 sorbitan tetraoleate, PEG-60 sorbitan tetraoleate, PEG-60 sorbitan tetrastearate, PEG-20 sorbitan triisostearate, PEG-160 sorbitan triisostearate, PEG-18 sorbitan trioleate, Sorbeth-40 hexaoleate (previously PEG-40 sorbitol hexaoleate), Sorbeth-50 hexaoleate (previously PEG-50 sorbitol hexaoleate),

Sorbeth-30 tetraoleate laurate (previously PEG-30 sorbitol tetraoleate laurate), and Sorbeth-60 tetrastearate (previously PEG-60 sorbitol tetrastearate).

Saccharide Ester Surfactants:

Sugar functional groups, after condensation with a fatty acid, form saccharide esters. All sugars are assumed to be linked through an ester bond formed from a condensation reaction with a carboxylic acid of a hydrolyzed maleic acid or free fatty acid and an alcohol group of the sugar. There are multiple chemical methods to form sugar fatty acid esters. Esterification can also occur enzymatically, most typically with a range of Lipase. Non-limiting examples of saccharide ester surfactants include: Sorbitan monolaurate, mannitan oleate, glucose pentaacetate, maltitol laurate, raffinose isostearate, raffinose myristate, raffinose oleate, sucrose acetate isobutyrate, sucrose acetate/stearate, sucrose benzoate, sucrose cocoate, sucrose dilaurate, sucrose dipalmitate, sucrose distearate, sucrose hexaerucate, sucrose hexaoleate/hexapalmitate/hexastearate, sucrose hexapalmitate, sucrose laurate, sucrose myristate, sucrose octaacetate, sucrose oleate, sucrose palmitate, sucrose palmitate/stearate or sucrose stearate-palmitate ester, sucrose pentaerucate, sucrose pentahydroxystearate, sucrose polybehenate, sucrose polycottonseedate, sucrose polylaurate, sucrose polylinoleate, sucrose polyoleate, sucrose polysoyate, sucrose polystearate, sucrose stearate, sucrose tetrahydroxystearate, sucrose tetraisostearate, sucrose tetrastearate triacetate, sucrose tribehenate, sucrose trilaurate, sucrose tristearate, trehalose isostearate, sorbitan monolaurate, mannitan oleate, erythritol laurate, erythritol myristate, sucrose monolaurate, maltose monolaurate, lactose monolaurate; sucrose monodecanoate, maltose monodecanoate, lactose monodecanoate; and sucrose monooctanoate, maltose monooctanoate, lactose monooctanoate, monostearoyl xyloglucan, and fructose laurate.

Sulfonation of Sugar based surfactants:

Sulfonated sugar esters generally give better solubility than sugar esters alone, allowing for increased solubility of long alkyl chain surfactants (stearate).

Amide sugar based surfactants:

This category of surfactants may be characterized by an amide linking the hydrophilic sugar head groups to a lipophilic tail.

Amino alcohol surfactants:

This class of surfactant is general of the formula: $R-C(=O)-NH-X-OH$, where R is generally, a C₈ to C₄₀ alkyl, and X can be a C₁ to C₁₀ alkyl chain. In some embodiments, R may be a C₄ to C₂₀ alkyl, C₅ to C₁₈ alkyl, C₆ to C₁₇ alkyl, C₇ to C₁₆ alkyl, or a C₈ to C₁₅ alkyl. A non-limiting example of a surfactant: cocamide monoethanolamine, wherein R is a C₁₁ alkyl.

Amino Acid based surfactants:

Non-limiting examples of this class of surfactants includes: Lauroyl Glutamic Acid, Sodium cocoyl Glycinate, - N-dodecanoylaminomalonic acid, and N-dodecanoylaspartic acid.

Polyol Surfactants:

A molecule with more than two hydroxyl group is a polyol. They are relatively easy to attach to a fatty acid, or other non-polar group comprising a carboxylic acid group by condensation of the carboxylic acid and alcohol into a fatty acid ester. Non-limiting examples of polar polyol groups that may be attached to a desired non-polar group include: Polyglycerol, and pentaerythritol. A non-limiting example of a polyol surfactant is polyglyceryl laurate.

Zwitterionic Polar Groups:

Non-limiting examples of zwitterionic polar groups includes: glycinate, betaine, sultaines (sometimes referred to as sulfobetaines), hydroxysultaine, ethanolamide, ammonioalkyl sulfates (general formula: $R-N^+(CH_2)_2XSO_4^-$. X is an alkyl chain can be between 1 to 6 carbons, most typically 3 carbons), and phosphocholines. Non-limiting examples of surfactants in this category include: lysophospholipids, fos-cholines, cocoamidylpropyl betaine, coco-hydroxysultaine, cocamidopropyl hydroxysultaine, capryl sultaine cetyl/lauryl/myristyl hydroxysultaine, coco-sultaine, erucamidopropyl hydroxysultaine, lauramidopropyl hydroxysultaine, lauryl hydroxysultaine, lauryl sultaine myristamidopropyl hydroxysultaine, myristyl sultaine, oleamidopropyl hydroxysultaine, and tallowamidopropyl hydroxysultaine, and lysophospholipid c.

Surfactants suitable for use in the present invention have an aggregate diameter. As used herein the term “aggregate diameter” refers to the size of an aggregate of the surfactant in aqueous conditions as measured by dynamic light scattering. An “average aggregate diameter” refers to the average of the size of a plurality surfactant aggregates in an aqueous environment as measured by dynamic light scattering. The term “average aggregate diameter” may be used interchangeably herein with the term “aggregate diameter”. Examples, without limitation, of how to measure an average aggregate diameter are provided in the Examples section herein.

Surfactant association with non-water moieties is often used to stabilize contaminants in an aqueous solution. Modifying the head and tail regions of a surfactant can substantially alter the morphology of a surfactant aggregate formation, a hydrophobic lipid balance of the molecule, and consequentially the surfactant aggregate’s ability to bind and stabilize various contaminants in a solution. For example, modifying the length of the alkyl chain of a surfactant may modify the hydrophobic lipid balance of the surfactant molecule, which may have an effect on aggregate shape, ability to solubilize oil in water, and/or surfactant aggregation number. Surfactants may also be used to increase flotation efficiency of contaminants by increasing their association with microbubbles. However, often, when using surfactants aggregates alone, surfactant aggregate remains in the water after treatment, making the produced water chemically incompatible with many industrial processes, particularly those which may require high surface tension in the water.

As used herein, the term “polymer” refers to a relatively high molecular weight substance which is at least partially soluble in an aqueous environment. Polymers are composed by linking together small molecules, called monomeric units, in sufficient number such that the addition or removal of one or several units does not substantially change the properties of the polymer. A monomeric unit refers to the group of atoms, derived from a monomer, comprising the unit that is repeated in the polymer. (Milton Orchin, *et al.* in Vocabulary of Organic

Chemistry p. 518 (John Wiley & Sons, 1980)). The monomeric units may be linked in a regular repeating or irregular repeating manner.

In some embodiments, the polymer in a nanonet composition may be comprised of two different monomeric units. In some embodiments, the polymer in a nanonet composition may be comprised of three different monomeric units. In some embodiments, the polymer in a nanonet composition may be comprised of a plurality of different monomeric units. In some embodiments, the polymer in a nanonet composition may be comprised of linear chains or of branched chains. In some embodiments, some monomeric units may be relatively more hydrophilic whereas other monomeric units may be relatively more hydrophobic.

In some embodiments, the different monomeric units comprising the polymer are regularly alternating, for example, --A-B-A-B--. In some embodiments, the different monomeric units comprising the polymer may substantially alternate in a different pattern, for example, --A-A-B-A-A-B--. This type of alternation can be thought of as a block. In some embodiments, a larger number of monomeric units comprising the polymer may repeat in a larger sequence before alternating to repetition of another monomeric unit in the polymer. The larger the number of the same repeating monomeric unit the larger the block in a polymer. In some embodiments of a polymer comprising a nanonet composition, a block comprises two monomeric units of a first type, and one monomeric unit of a second type. In other embodiments, a block may comprise two monomeric units of a first type, a monomeric unit of a second type, and a monomeric unit of third type.

In some embodiments, the polymer comprising the nanonet composition has at least one hydrophilic monomeric unit and at least one hydrophobic monomeric unit. The polymer in a nanonet composition may be made from styrene-maleic acid copolymer. The polymer in a nanonet composition may be hydrolyzed styrene-maleic anhydride copolymer. In some embodiments, the polymer may be hydrolyzed di-isobutylene-maleic anhydride copolymer (DIB-co-maleic anhydride). As with SMA, DIB-co-maleic anhydride is not soluble in aqueous solution, but becomes soluble when at least a portion of the maleic

anhydride monomeric unit are hydrolyzed to the di-carboxylic acid form. What follows holds for each of SMA and DIB-co-maleic anhydride, and a resulting water-soluble di-carboxylic acid containing form therefrom. It is to be appreciated herein that SMA is often used herein as an abbreviation for styrene-maleic acid copolymers and DIBMA is used as an abbreviation for di-isobutylene-maleic anhydride copolymers and that such abbreviations are context specific and these two abbreviations may refer to both the hydrolyzed and non-hydrolyzed forms of these copolymers and a person of skill in the art will be readily able to identify whether or not the copolymer is in a hydrolyzed form or not. The use of the abbreviates hSMA and hDIBMA always refer to the hydrolyzed form of these copolymers.

SMA is generally insoluble in aqueous solutions. To increase the solubility of SMA, the polymer may be heated in a basic pH solution leading to at least some portion of the maleic anhydride units in the polymer becoming hydrolyzed, thereby forming hSMA. This means that some portion of the repeating maleic anhydride units in hSMA are chemically converted to adjacent CH groups each bearing a pendant carboxylic acid group. Chemically, this dicarboxylic acid repeating unit in hSMA may be expressed as follows: $-\text{CH}(\text{COOH})-\text{CH}(\text{COOH})-$, or a salt form thereof wherein one or both of the adjacent carboxylic acid units may be in the protonated acid form, or a salt thereof. In some embodiments, the adjacent carboxylic acid units may be in a salt form with a single divalent or multi-valent cation. For example, divalent and multi-valent cations such as, but not limited to, ammonia ions, calcium ions, magnesium ions, or ferrous or ferric iron ions.

In some embodiments of a nanonet composition, the percentage of hydrolysis of SMA to hSMA (or DIB-co-maleic anhydride to hDIBMA) is an amount effective to substantially solubilize the polymer in aqueous solution under conditions for nanonet formation. In some embodiments, substantially all of the maleic acid units are hydrolyzed to create dicarboxylic acid repeating units in the polymer chain. In some embodiments, not all of the maleic acid units are hydrolyzed.

In some embodiments, at least one hydrophilic monomeric unit is maleic anhydride hydrolyzed to maleic acid or a salt thereof and having the general chemical formula $[-CH(COR^6)CH(COR^7)-]$. In some embodiments, a sufficient number of maleic acid monomeric units (by hydrolysis of some maleic anhydride monomeric units) are present for the polymer to dissolve in aqueous solution in a concentration sufficient to form a nanonet composition (see Figure 14). When all maleic anhydride monomeric units in the polymer are fully hydrolyzed (for example polymer 2000), R^6 and $R^7 = OH$ (carboxylic acid functional group).

In some embodiments, some of the maleic anhydride monomeric units in the polymer may be modified thereby changing any one or more of R^6 , R^7 or both. For example, any one or more of R^6 , R^7 , or both may be OR^1 , OR^2 , NHR^1 , NHR^2 , NR^1_2 , NR^2_2 , and SR^1 . This modification may be accomplished by any one or more substitution, displacement, or condensation reactions known to those skilled in the art, including but not limited to those set out in the Examples section.

R^6 , R^7 or both, R^1 , R^2 or both, may be a pendant group functionalizing the maleic acid monomeric unit in a polymer comprising a nanonet composition. For example, any of the R groups of this paragraph may be any one or more of hydrogen, C_1 - C_{40} , straight or branched chain, saturated, mono-unsaturated or poly-unsaturated, aliphatic cyclic, or aromatic cyclic, polycyclic, a sugar, pyridine, pyrimidine, purine, cyclodextrin, indole, isoindole, glycosides and piperazine. In some embodiments, the aromatic cyclic is alkylated.

In some embodiments the polymer has hydrophobic monomeric units comprising a polymer that include an unsaturated carbon chain having between one and ten additional carbon atoms. For example, styrene, *p*-methyl styrene, chlorostyrene, 2-chlorostyrene, 4-chlorostyrene, fluorostyrene, bromostyrene, propylene, isobutylene, isoprene, and mixtures thereof.

In other embodiments, the hydrophobic monomeric unit, may be a straight or branched chain, saturated, mono-unsaturated or poly-unsaturated, aliphatic cyclic, polycyclic, aromatic cyclic having at least one aromatic ring. In some embodiments, the aromatic cyclic is alkylated. Generally, the hydrophobic monomeric unit has as few as 3 and as many as 10 carbon atoms.

Polymers suitable for use in the present invention have an average particle diameter. As used herein "particle diameter" refers to the size of a polymer in aqueous conditions as measured by dynamic light scattering. "Theoretical polymer diameter" refers to the theoretical minimum diameter of a compact polymer sphere which may be calculated as described below. An "average particle diameter" refers to the average of the size of a plurality polymers in an aqueous environment as measured by dynamic light scattering. The term "average particle diameter" may be used interchangeably herein with the term "particle diameter". Examples, without limitation, of how to measure an average particle diameter are provided in the Examples section herein.

Polymers may be long polymers or short polymers. As used herein the term "long polymer" refers to a polymer with a molecular weight above 40,000 g/mol, and/or a contour length above 100nm. As used herein the term "short polymer" refers to a polymer that with a molecular weight below 40,000 Da, and/or a contour length below 100nm.

In some embodiments, functional groups on polymers suitable for use in nanonets interact with di-valent and tri-valent cations, such as maleic acid. Addition of di-valent and tri-valent cationic metals neutralizes a negative charge of carboxylate group present on the maleic acid, thereby precipitating the polymer. Corresponding association between the maleic acid functionalized block co-polymer with a surfactant aggregate renders the surfactant aggregate sensitive to the cations. Thus, addition of a nanonet into solutions containing high concentrations of di-valent and tri-valent cations will precipitate the entire complex of nanonet and cations. In some instances, suitably high concentrations of monovalent cations may also precipitate the entire complex of nanonets and associated cations. This resulting precipitate is hydrophobic in nature and binds non-water moieties, such as hydrocarbons. As time passes the precipitates begin to flocculate, thereby increasing in size and becoming more amenable to removal from solution by physical means such as settling or floatation. If surfactant and polymer are added sequentially into solutions containing large concentrations of a polymer specific precipitant, the non-water moiety binding

and flocculation effect of the surfactant is lost because the polymer immediately precipitates as individual particles, and the surfactant component remains free to dissociate in solution. In cases where a precipitant which acts on the surfactant is present in solution, sequential addition of polymer and detergent will result in separate polymer and surfactant precipitates and the flocculating effect is lost.

Illustrative embodiments of this invention provide a nanonet comprising:

- a) a surfactant aggregate having an average aggregate diameter; and
- b) a polymer having an average particle diameter which average particle diameter is the same or smaller than the average aggregate diameter. The resulting nanonet often has a diameter larger than the average particle diameter.

Other illustrative embodiments of this invention provide a nanonet comprising:

- a) a surfactant aggregate having an average aggregate diameter; and
- b) a polymer having an average particle diameter which average particle diameter is the same or smaller than the average aggregate diameter. The resulting nanonet often has a diameter larger than the average particle diameter. Specifically excluded from these embodiments are nanonets in which both:

- 1) the surfactant is selected from the group consisting of: Triton X-100TM, Triton X-305TM, N-dodecyl-beta-D-maltoside (DDM), sodium oleate, sodium cholate, sodium deoxycholate (DOC), beta-octyl glucoside (Beta-OG), and lauryl dimethylamine n-oxide (LDAO); and
- 2) the polymer is selected from the group consisting of: SMA-2000TM, SMA-2021TM, and Di-isobutylene-co-maleic acid (DIBMA).

For clarity, embodiments in which the surfactant is selected from Triton X-100TM, Triton X-305TM, DDM, sodium oleate, sodium cholate, DOC, Beta-OG, and LDAO are included in the present invention provided that the polymer is not exclusively and only one of SMA-2000TM, SMA-2021TM or DIBMA. Similarly,

embodiments in which the polymer is selected from the group consisting of: SMA-2000TM, SMA-2021TM and DIBMA are also included in the present invention provided that the surfactant is not exclusively and only one of Triton X-100TM, Triton X-305TM, DDM, sodium oleate, sodium cholate, DOC, Beta-OG, or LDAO.

In some other embodiments still, the surfactant is not a phospholipid. In some other embodiments still, the surfactant is not a lipid.

By selecting for a ratio of surfactant average aggregate diameter to polymer average particle diameter in the range of $1:\leq 1$, nanonet formation can be predicted to occur. Often it is preferable to select a ratio of surfactant average aggregate diameter to polymer average particle diameter in the range of $1:<1$. Often it is preferable to select a ratio of surfactant average aggregate diameter to polymer average particle diameter in the range of $1:<1$, where <1 is close to 1, but still less than 1. This allows for design of nanonets with varying surfactants and varying polymers that are stable and likely to contain a high number of surfactant molecules. By increasing the capture of surfactant molecules into nanonets, physical parameters that may be important for downstream application of the particles may also be selected for.

When selecting materials for preparation of a nanonet, a surfactant aggregate preferably has an average aggregate diameter larger than an average particle diameter of a polymer. This limits the choice of suitable surfactants that may be incorporated into a nanonet for a given size of polymer; the shorter the polymer, the more varieties of nanonets that may be formed. However, with increasing polymer length, nanonets, comprising a surfactant having an average aggregate diameter size that is larger or the same size when compared to the average particle diameter of the polymer may lead to smaller, more stable nanonets that may display enhanced flocculation efficiency and/or flotation characteristics. Therefore, selecting for longer polymers and selecting suitable surfactants based on the ratio of average aggregate diameter to average particle diameter of the selected polymer provides for stable nanonet formation with advantageous properties.

In preferred formation conditions, a polymer, when in a compact polymer particle, has an average particle diameter slightly smaller than the size of the surfactant's average aggregate diameter. In aqueous conditions and using a slight excess of polymer, the surfactant and polymer will self-assemble into a nanonets, leading to an increase in precipitated matter in solution (Fig. 1A). An example of this relationship as shown in the Examples below, where SMA-2000TM (4 nm in diameter, 14nm in length) and Triton X-100TM (5.3 nm in diameter, 16.6nm in circumference) form a nanonet (5.1nm in diameter, 16 nm in circumference) at a 1:1 wt/wt ratio.

A surfactant aggregate may be too large to be encompassed by a single polymer chain. This may lead to splitting of the original surfactant aggregate into smaller nanonets (Fig. 1C), similar to the solubilization action of SMA in lipid bilayers (Dörr, J.M., Koorengevel, M.C., Schäfer, M., Prokofyev, A.V., Scheidelaar, S., van der Crujisen, E.A.w., Dafforn, T.R., Baldus, M., Killian. J.A. Detergent-free isolation of KcsA in nanodiscs. *PNAS*. Dec 2014, 111 (52) 18607-18612). This partitioning of surfactant between polymers is exemplified in the Examples below with PGL1, which was solubilized into 25nm particles in both SMA-2000TM and SMA-725TM (Table 4). However, multiple polymers may also associate around a large surfactant aggregate, which in some cases may lead to an increase in size of the nanonet when compared to the surfactant average aggregate diameter (Fig. 1D).

A surfactant aggregate may also be too small for a given length of polymer (Fig. 2A and 2B, triangles). However, whereas a large surfactant aggregate may form a nanonet with a smaller polymer, a surfactant aggregate that is too small will remain separate and not form a nanonet with a larger polymer. For example, as set out in the Examples below, Triton X-100TM and Triton X-305TM aggregates are both too small to efficiently form nanonets with the large SMA-725TM polymer, but are able to form nanonets with the smaller SMA-2000TM polymer. This may be because of the free-energy landscape which governs amphipathic molecule association. Surfactant aggregates may form due to balancing the

entropic cost of a multimolecular assembly with the free energy conserved by shielding of hydrophobic alkyl chains from aqueous solution. This shielding of the hydrophobic groups from water also occurs in SMA polymers, which will form compact particle structures in buffered aqueous solutions. A compact polymer particle represents the smallest diameter conformation that a polymer can adopt. In the case of a small surfactant aggregate and a large polymer, an increase in size of both the surfactant and the polymer caused by their association would therefore be entropically unfavourable and not occur in the absence of another factor (Fig. 1B). Possible factors may be enthalpic contributions such as hydrogen bonding, salt bridging, or electrostatic interactions between the surfactant and polymer.

In some embodiments, it is possible to determine nanonet assembly using the following formula:

Volume of backbone, non-hydrogen atom (C,O,S,N) $\approx 20 \text{ \AA}^3$

Volume of aromatic ring = $84 \text{ \AA}^3$

Diameter of surfactant aggregate ($D_{\text{aggregate}}$) = $2 \times ((n \times V_{\text{sur}}) / \pi / 4 \times 3)^{1/3}$, where V is the calculated volume of a surfactant monomer, and n is the number of surfactant molecules in the surfactant aggregate.

Diameter of a compact polymer micelle (D_{polymer}) = $2 \times ((n_{\text{block}} \times V_{\text{block}}) / \pi / 4 \times 3)^{1/3}$, where V is the calculated volume of the polymer block, and n is the number of blocks in a single polymer chain.

Calculation of SMA-2000TM compact polymer diameter: $Mw_{\text{block}} = 324 \text{ g/mol}$; (2 styrene, 1 maleic acid), $Mw_{\text{SMA2000}} \approx 7500 \text{ g/mol}$

$n_{\text{block}} = Mw_{\text{polymer}}/Mw_{\text{block}} \approx 23$;

$V_{\text{block}} = 2 \text{ aromatic rings, } 8 \text{ carbon atoms, } 4 \text{ oxygen atoms} = 408 \text{ \AA}^3$;

$V_{\text{polymer}} = n_{\text{block}} \times V_{\text{block}} = 23 \times 408 \text{ \AA}^3 = 9,384 \text{ \AA}^3$;

$D_{\text{polymer}} = 2 \times ((n_{\text{block}} \times V_{\text{block}}) / \pi / 4 \times 3)^{1/3} = 2 \times ((9,384 \text{ \AA}^3) / \pi / 4 \times 3)^{1/3} = 26 \text{ \AA} = 2.6 \text{ nm}$

Calculation of SMA-2021TM compact polymer diameter: $Mw_{\text{block}} = 324 \text{ g/mol}$; (2 styrene, 1 maleic acid), $Mw_{\text{SMA2021}} \approx 22,000 \text{ g/mol}$

$n_{\text{block}} = Mw_{\text{polymer}}/Mw_{\text{block}} \approx 68$

$V_{\text{block}} = 2 \text{ aromatic rings, 8 carbon atoms, 4 oxygen atoms} = 408 \text{ \AA}^3$;

$V_{\text{polymer}} = n_{\text{block}} * V_{\text{block}} = 68 \times 408 \text{ \AA}^3 = 27,744 \text{ \AA}^3$;

$D_{\text{polymer}} = 2 \times ((n_{\text{block}} \times V_{\text{block}}) / \pi / 4 \times 3)^{1/3} = 2 \times ((27,744 \text{ \AA}^3) / \pi / 4 \times 3)^{1/3} = 37 \text{ \AA} = 3.7 \text{ nm}$

Calculation of SMA-725TM compact polymer diameter: $Mw_{\text{block}} = 428 \text{ g/mol}$;
(3 styrene, 1 maleic acid), $Mw_{\text{SMA725}} \approx 130,000 \text{ g/mol}$

$n_{\text{block}} = Mw_{\text{SMA725}} / Mw_{\text{block}} \approx 308$

$V_{\text{block}} = 3 \text{ aromatic rings, 10 carbon atoms, 4 oxygen atoms} = 532 \text{ \AA}^3$;

$V_{\text{polymer}} = n_{\text{block}} * V_{\text{block}} = 308 \times 532 \text{ \AA}^3 = 163,856 \text{ \AA}^3$;

$D_{\text{polymer}} = 2 \times ((n_{\text{block}} \times V_{\text{block}}) / \pi / 4 \times 3)^{1/3} = 2 \times ((163,856 \text{ \AA}^3) / \pi / 4 \times 3)^{1/3} = 68 \text{ \AA} = 6.8 \text{ nm}$

Calculation of SMA-3000TM compact polymer diameter: $Mw_{\text{block}} = 428 \text{ g/mol}$;
(3 styrene, 1 maleic acid), $Mw_{\text{SMA3000}} \approx 9,000 \text{ g/mol}$

$n_{\text{block}} = Mw_{\text{SMA3000}} / Mw_{\text{block}} \approx 21$

$V_{\text{block}} = 3 \text{ aromatic rings, 10 carbon atoms, 4 oxygen atoms} = 532 \text{ \AA}^3$;

$V_{\text{polymer}} = n_{\text{block}} * V_{\text{block}} = 21 \times 532 \text{ \AA}^3 = 11,172 \text{ \AA}^3$;

$D_{\text{polymer}} = 2 \times ((n_{\text{block}} \times V_{\text{block}}) / \pi / 4 \times 3)^{1/3} = 2 \times ((11,172 \text{ \AA}^3) / \pi / 4 \times 3)^{1/3} = 28 \text{ \AA} = 2.8 \text{ nm}$

Calculation of SMA-IBE compact polymer diameter: $Mw_{\text{block}} = 220.2 \text{ g/mol}$;
(1 styrene, 1 maleic acid), $Mv_{\text{SMA-IBE}} \approx 65,000 \text{ g/mol}$

$n_{\text{block}} = Mv_{\text{polymer}} / Mw_{\text{block}} \approx 295$

$V_{\text{block}} = 1 \text{ aromatic ring, 6 carbon atoms, 4 oxygen atoms} = 284 \text{ \AA}^3$;

$V_{\text{polymer}} = n_{\text{block}} * V_{\text{block}} = 295 \times 284 \text{ \AA}^3 = 83,780 \text{ \AA}^3$;

$D_{\text{polymer}} = 2 \times ((n_{\text{block}} \times V_{\text{block}}) / \pi / 4 \times 3)^{1/3} = ((83,780 \text{ \AA}^3) / \pi / 4 \times 3)^{1/3} = 54 \text{ \AA} = 5.4 \text{ nm}$

Calculation of SMA-Cumene terminated compact polymer diameter:
 $Mw_{\text{block}} = 428 \text{ g/mol}$; (3 styrene, 1 maleic acid), $Mw_{\text{Cumene}} = 152 \text{ g/mol}$ $Mw_{\text{SMA-Cumene}} \approx 1500 \text{ g/mol}$

$n_{\text{block}} = Mw_{\text{SMA-Cumene}} - Mw_{\text{Cumene}} / Mw_{\text{block}} \approx 3.1$

$V_{\text{block}} = 3 \text{ aromatic rings, 10 carbon atoms, 4 oxygen atoms} = 532 \text{ \AA}^3$;

$V_{\text{polymer}} = n_{\text{block}} * V_{\text{block}} + V_{\text{Cumene}} = (3.1 \times 532) + 144 \text{ \AA}^3 = 1,819 \text{ \AA}^3$;

$$D_{\text{polymer}} = 2 \times ((n_{\text{block}} \times V_{\text{block}}) / \pi / 4 \times 3)^{1/3} = 2 \times ((1,819 \text{ \AA}^3) / \pi / 4 \times 3)^{1/3} = 15 \text{ \AA} = 1.5 \text{ nm}$$

Calculation of DIBMA compact polymer diameter: $M_{w\text{block}} = 226 \text{ g/mol}$; (18 hydrogen atoms, 12 carbon atoms, 4 oxygen atoms), $M_{w\text{DIBMA}} \approx 12,000 \text{ g/mol}$

$$n_{\text{block}} = M_{w\text{DIBMA}} / M_{w\text{block}} \approx 53$$

$$V_{\text{block}} = 12 \text{ carbon atoms, 4 oxygen atoms} = 320 \text{ \AA}^3;$$

$$V_{\text{polymer}} = n_{\text{block}} \times V_{\text{block}} = 53 \times 320 \text{ \AA}^3 = 16,960 \text{ \AA}^3;$$

$$D_{\text{polymer}} = 2 \times ((n_{\text{block}} \times V_{\text{block}}) / \pi / 4 \times 3)^{1/3} = 2 \times ((16,960 \text{ \AA}^3) / \pi / 4 \times 3)^{1/3} = 32 \text{ \AA} = 3.2 \text{ nm}$$

Calculation of Ethanol derived SMA-2000TM compact polymer diameter:

$M_{w\text{block}} = 367 \text{ g/mol}$; (22 carbon atoms, 4 oxygen atoms, 1 nitrogen atom, 25 hydrogen atoms), $M_{w\text{Ethanol derived SMA2000}} \approx 8495 \text{ g/mol}$

$$n_{\text{block}} = M_{w\text{polymer}} / M_{w\text{block}} \approx 23;$$

$$V_{\text{block}} = 2 \text{ aromatic rings, 10 carbon atoms, 4 oxygen atoms, 1 nitrogen atom} = 468 \text{ \AA}^3;$$

$$V_{\text{polymer}} = n_{\text{block}} \times V_{\text{block}} = 23 \times 468 \text{ \AA}^3 = 10,764 \text{ \AA}^3;$$

$$D_{\text{polymer}} = 2 \times ((n_{\text{block}} \times V_{\text{block}}) / \pi / 4 \times 3)^{1/3} = 2 \times ((10,764 \text{ \AA}^3) / \pi / 4 \times 3)^{1/3} = 27 \text{ \AA} = 2.7 \text{ nm}$$

Calculation of Triton x-100TM surfactant aggregate diameter:

$$(D_{\text{aggregate}}) = 2 \times ((n \times V_{\text{sur}}) / \pi / 4 \times 3)^{1/3}$$

$$N_{\text{surfactant}} = 100-155, V_{\text{surfactant}} = 10 \text{ oxygen atoms, 29 carbon atoms, 1 aromatic ring} = 864 \text{ \AA}^3$$

$$(D_{\text{aggregate}}) = 2 \times ((100 \times 864 \text{ \AA}^3) / \pi / 4 \times 3)^{1/3} = 21 \text{ \AA} = 5.2 \text{ nm}.$$

For Tx-100, $D_{\text{aggregate}} > D_{\text{SMA2000}}$; Nanonet formation is predicted.
 $D_{\text{aggregate}} < D_{\text{SMA725}^{\text{TM}}}$; Nanonet formation is incompatible with SMA725TM.
 $D_{\text{aggregate}} > D_{\text{SMA2021}}$ Nanonet formation is predicted.

Calculation of Triton x-305TM surfactant aggregate diameter:

$$(D_{\text{aggregate}}) = 2 \times ((n \times V_{\text{sur}}) / \pi / 4 \times 3)^{1/3}$$

$$N_{\text{surfactant}} = 30, V_{\text{surfactant}} = 30 \text{ oxygen atoms, 69 carbon atoms, 1 aromatic ring} = 3,684 \text{ \AA}^3$$

$$(D_{\text{aggregate}}) = 2 \times ((30 \times 3,684 \text{ \AA}^3) / \pi / 4 \times 3)^{1/3} = 59.5 \text{ \AA} \approx 6 \text{ nm}.$$

For Triton x-305TM, $D_{\text{aggregate}} > D_{\text{SMA2000}}$; Nanonet formation is predicted with SMA-2000TM. $D_{\text{aggregate}} < D_{\text{SMA725}}$; Nanonet formation is incompatible with SMA-725TM. $D_{\text{aggregate}} > D_{\text{SMA2021}}$ Nanonet formation is predicted.

Table 1: Diameters of Selected Polymers and Surfactants.			
	MW	Estimated Diameter	Measured Diameter
SMA 2000 TM	7,500 Da	2.6 nm*	4 nm
SMA 2021 TM	22,000 Da	3.7nm*	N/A
SMA 725 TM	130,000 Da	6.8 nm*	10 nm
Triton X-100 TM	650 Da	5.4 nm	5 nm
Triton X-305 TM	1500 Da	6 nm	9 nm
Triton X-45 TM	400 Da	N/A	91 nm

* Assuming fully compacted polymer in generally spherical shape.

In some illustrative embodiments of the present invention, the surfactant is selected from the group consisting of: PGL1, PGL2, PGL3, PGL4, PGL5, Triton X-45TM, Triton X-100TM, Triton X-305TM, sodium oleate, potassium oleate, LDAO, polyoxyethylene sorbitan monolaurate, polyoxyethylene sorbitan monooleate, polyoxyethylene sorbitan myristate, polyoxyethylene sorbitan stearate, polyoxyethylene sorbitan palmitate, lauramidopropyl betaine, cocoyl glycinate, glucose laurate monoester, sucrose laurate, sucrose monooleate, sucrose palmitate, glyceryl monolaurate, fructose laurate, potassium oleate, oleic acid, palmitic acid, linoleic acid, and mixtures thereof. In some embodiments, the surfactant is a mixture of PGL5 and sodium oleate. In some embodiments, the surfactant is a mixture of PGL5 and potassium oleate.

In illustrative embodiments of the present invention, the polymer is a block co-polymer. In some embodiments, the polymer has a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 4:1, from about 1:1 to about 3:1, from about 2:1 to about 3:1, or about 2:1. In some embodiments, the polymer is selected from the group consisting of: SMA-725TM, SMA 3000TM,

SMA-2000TM, and SMA-2021TM. In some embodiments, the polymer is SMA-725TM.

Nanonets according to the present invention may be made by mixing, in aqueous environment, a polymer and a surfactant. It is often advantageous to add enough surfactant such that the concentration of surfactant is at the concentrated micelle concentration (CMC). Nanonets may be prepared in a more clear and/or sterile environment and then added to an aqueous solution for treatment later, or may be provided as separate parts in a kit for mixing later. The parts of the kit may be mixed in a clean and/or sterile environment or they may be mixed *in situ* for preparation of the nanonets directly in the aqueous solution to be treated.

In illustrative embodiments, there is provided a method of treating produced water using nanonets described herein. An example of such a method is injecting a solution of nanonets into a liquid flow. The manner of injection may be through any method known to a person of skill in the art, and is often through an eductor, such as a Venturi eductor. The nanonet solution may be actively pumped into the eductor, or naturally drawn into the fluid flow by the venturi effect if a suitable pressure drop is present. The nanonet solution may be injected into the fluid flow alone, or concurrently with a gas or other water chemicals. The nanonet solution may also be injected through an injection coil followed by a static mixer. In such a case it may be necessary to first dilute the nanonet solution to facilitate mixing in a pipe.

Examples

The following examples are illustrative of some of the embodiments of the invention described herein. These examples do not limit the spirit or scope of the invention in any way.

In the following Examples 1-14, the following methods reagents and materials were used.

Reagents and Materials:

Unless otherwise stated, all chemicals were purchased from Sigma-AldrichTM Canada. Polyglycerol-10-palmitate (PGL5), Polyglycerol-10-oleate (PGL1), Polyglycerol-10-stearate (PGL3), Polyglycerol-10-laurate 1 (PGL2) and 2 (PGL4) were purchased from Shandong Binzhou New Material Technology Co.TM, Ltd. Styrene maleic anhydride block co-polymer was purchased from Cray ValleyTM (SMA-2000TM), or Jiaying Huawen Chemical Co.TM, Ltd (SMA-725TM).

Preparation of Concentrated Polymer Stock Solutions of SMA-725TM and SMA-2000TM

For preparation of styrene maleic acid co-polymers, SMA-725TM was first ground to a fine powder, while SMA-2000TM was purchased in powder form. 5% (wt/vol) of pre-hydrolyzed SMA-725TM and SMA-2000TM were subsequently incubated in 500mM potassium hydroxide at 80°C with stirring for 5 hours or until the solution had cleared. Solutions were passed through a 10 µm filter to remove non-hydrolyzed particles, then precipitated by dropwise addition of 12M HCl with stirring until pH ≤ 2. Precipitated polymer was filtered (10 µm), washed 3x with 100mL of dilute HCl (50mM), 3X with 100mL of ultrapure water, then left to dry at room temperature for 72 hours. The concentrated polymer stock solution was then prepared by resuspending 200mg of the dry polymer in ultrapure water and adjusting the pH of the suspension to 7 by dropwise addition of sodium hydroxide. The resulting clear polymer solution was then adjusted to a final volume of 10mL and final concentration of 2% (wt/vol). All polymer stock solutions were prepared according to this procedure and diluted accordingly in distilled water before use.

Polymer stock solutions prepared according to the procedure immediately above were used in the examples set forth in the following sections; even if a polymer is referred to only by its name or trade name.

Preparation of Stock Surfactant Solutions.

Stock Triton X-100™, Triton X-305™, and Triton X-45™ solutions were prepared by mixing 300mg of surfactant into a final volume of 30mL of ultrapure water. Stock solutions of PGL1, PGL3, or PGL5, respectively, were prepared by weighing out 1 g, 500mg, or 330mg, respectively. The weighed amount of the selected surfactant was resuspended in 10mL of ultrapure water and mixed with a handheld homogenizer until emulsified to produce each respective PGL surfactant stock solution. For all other surfactant stock solutions, 300mg of surfactant was dissolved into 30mL of ultrapure water to form 1% stock solutions.

Nanonet Formation Assays:

Concentrated polymer (stock solution of 0.5% wt/vol) and surfactant (stock solution of 1% wt/vol) were mixed in a microcentrifuge tube. The amount of each stock solution employed, depended upon the desired surfactant to polymer ratio in the examples that follow. The combined concentrated stock solutions in the microcentrifuge tube were then diluted to a final volume of 1mL in ultrapure water. The tubes were mixed by hand inversion 3 times, left to sit for 5 minutes, and then 1 drop (approximately 10uL) of 12M hydrochloric acid was introduced to stimulate nanonet precipitation. Tubes were mixed 3 times by inversion, left to sit at room temperature for 5 minutes. To detect the formation of nanonets, the absorbance of the tube contents was measured at 540 nm.

Size Measurement With Dynamic Light Scattering (DLS):

Polymer particle size, surfactant aggregate size, and nanonet size measurements were measured using dynamic light scattering (Zetasizer Nano ZS™). In brief, 0.5% wt/vol solutions of surfactants were prepared in Buffer A (20mM Tris-HCl {Tris(hydroxymethyl)aminomethane}, pH 7). Surfactants were mixed at a 1:1 ratio (wt/wt) with the indicated polymer and allowed to sit 5 minutes at room temperature. Additionally, polymer only and surfactant only samples, respectively, were mixed with an equivalent volume of Buffer A and incubated for an equivalent time period. After 5 minutes, large aggregates and

dust were removed from the solutions by 0.2 μ M filter, followed immediately by measuring size by dynamic light scattering (absorbance at 540 nm). During sizing, samples were diluted dropwise in Buffer A to obtain an optimum polydispersity index (<0.5) for measurements. Number values were taken as a measure of the diameter for surfactant aggregate, nanonet and polymer particle diameters.

Flocculation Speed Experiments:

1% sodium oleate nanonets were formed by addition of 1:1 (wt:wt) of the indicated SMA polymer and sodium oleate. Sodium oleate nanonet solutions were then diluted to 0.02% by weight/volume in 1mL of ultrapure water and allowed to sit for 5 minutes at room temperature (21°C) in a polystyrene cuvette. Nanonets were subsequently destabilized by acidification (1 drop 12M HCl), and the absorbance measured at 540nm. Settling of the precipitated sodium oleate nanonet floc was tracked over time. As a control, the settling behavior of acidified sodium oleate aggregates were also tracked over time.

Example 1

Size Characterization of Non-Ionic Surfactant Aggregates.

A comparison of nanonet formation with one relatively smaller polymer (SMA-2000TM), and one relatively larger polymer (SMA-725TM) SMA polymer, using the same surfactant series was undertaken. Each of these polymers are styrene maleic acid block co-polymers. The molecular weight averages for SMA-2000TM and SMA-725TM are 6,000 Da and 130,000 Da, respectively.

To examine the relationship between polymer particle size and surfactant aggregate size in nanonet formation, surfactants from each of the TritonTM X series and polyglyceryl-10-alkyl (PG) series of surfactants were tested. First, the surfactant average aggregate diameter was measured by dynamic light scattering (Table 2). For the TritonTM X series of surfactants, the higher the hydrophobic lipid balance (HLB) of the surfactant, the smaller the average aggregate diameter ($D_{TX-45} > D_{TX-305} \approx D_{TX-100}$). This may be due to the increase in

the hydrophilic head-group component of the surfactant, which may lead to increased curvature of the surfactant aggregate, and thus a smaller measured diameter of the aggregate. For the PG series of surfactants, the measured aggregate diameters did not fit this trend, possibly due to varying lengths and degrees of saturation in the surfactants' alkyl chains, which may alter monomer packing and possibly aggregate structure.

Table 2. Dynamic Light Scattering Characterization of Surfactant Average Aggregates and Activated SMA Polymers.				
Additive Surfactant	Poly Dispersity Index (PDI)	Average Aggregate Diameter (nm)	Hydrophobic Lipid Balance	Alkyl Chain Carbons
0.5% Triton X-305™	0.162	9.913	17.3	12
0.5% Triton X-45™	0.467	90.7	9.8	12
0.5% Triton X-100™	0.246	5.327	13.5	12
0.5% PGL1	0.356	46.05	10.5	18
0.5% PGL 2	0.169	96.66	12	12
0.5% PGL 3	0.541	304.9	11	16
0.5% PGL 4	0.109	69.98	14	12
0.5% PGL 5	1	557	10.5	18
0.5% Sodium Oleate	NA	392	18	18
Polymer	PDI	Average Particle Diameter (nm)	Contour Length	
0.5% SMA-2000™	0.945	4.305	14nm	
0.5% SMA-725™	0.418	10.73	349nm	

Example 2

Self-Assembly of Nanonets May Be Driven By Factors Including Polymer Average Particle Diameter, Surfactant Average Aggregate Diameter, and the Ratio (wt/wt) of Polymer to Surfactant in Solution.

Here, the analysis of nanonet formation began by seeking to determine whether any given combination of surfactant and polymer led to the co-precipitation of surfactant and polymer in a complex upon acidification of the solution (Fig. 2). Each of the PG and Triton™ series of surfactants are generally

stable in acidic solution. It is noted, however, that the PG surfactant series can slowly degrade in very acidic conditions. Upon combining solutions of a given SMA polymer and surfactant from the PG or Triton™ series, and even as polymer particles and surfactant aggregates interact, an overall negative charge prevails. That charge can be neutralized by protonation (via addition of a proton source, e.g. acid). Upon such neutralization, a voluminous precipitate immediately forms and begins to flocculate and settle out of solution. The formation of this precipitate can be measured by changes in the absorbance of the solution at 540 nm, a measure of solution turbidity. The rate and amount of formation of turbidity in any given solution is taken as a measurement of nanonet formation.

Triton X-100™, Triton X-305™, and Triton X-45™ showed co-precipitation of surfactant and polymer upon addition of surfactant (Fig. 2A, 2B, and 2C, squares). Once above the 1:1 ratio (wt/wt) of surfactant to polymer, addition of excess surfactant did not significantly increase precipitation. This suggested that once the polymer is saturated, addition of excess surfactant is not incorporated into the nanonet composition.

In the case of Triton X-305™, excess surfactant (500ppm) led to re-solubilization of the nanonet precipitate (Fig. 2A, squares). A similar trend was observed with Triton X-100™, but much higher concentrations (>500ppm) were necessary to cause solubilization of the nanonet precipitate (Fig. 2B, squares). In the case of Triton X-45™, the surfactant at high concentrations is turbid, preventing differentiation between nanonet precipitation and basal surfactant aggregates, thus re-solubilization was not detected.

At the ratio of 1:1 wt:wt, the SMA-2000™ polymer was able to form nanonets and co-precipitate with any of the Triton™ series surfactants (Fig. 2A, 2B, and 2C, squares). To next test the effect of polymer length on nanonet formation, the experiments were repeated with the higher molecular weight polymer SMA-725™. In contrast to SMA-2000™, SMA-725™ was only able to

form nanonets with the relatively larger surfactant aggregates formed by Triton X-45TM (Fig. 2A, 2B, and 2C, triangles).

These experiments were repeated using the same two SMA polymers but with the PG series of surfactants. In nearly all cases, the PG series of surfactants were able to co-precipitate with both polymers, indicating the formation of nanonets. These results are consistent with the large aggregate diameters of the PG surfactants recorded by dynamic light scattering (Table 2). The smallest surfactant aggregate diameter recorded was 46nm for polyglyceryol-10-monoleate (PGL1), nearly 4 times greater than the measured average particle diameter of the polymer only particle formed by SMA-725TM (10.7 nm, Table 2).

Example 3

Relationship of Polymer Average Particle Size to Surfactant Average Aggregate Diameter Is A Significant Factor That Drives Nanonet Formation

To understand why polymer average particle size may matter for nanonet formation, the average particle diameters of the polymer particles in a buffered solution were measured by dynamic light scattering (Table 2). Although each polymer is an extended chain having a contour length (nm) in basic conditions, the extended chain conformation does not necessarily exist under other solution conditions. For example, in buffered solution at neutral pH, the polymers will often adopt a compact particulate structure having a measurable and calculable average particle diameter.

For example, consider SMA-2000TM in dynamic light scattering measurements. Although SMA-2000TM has a contour length of 14nm, the polymer formed a compact particle having an average particle diameter of $\approx$ 4nm (Table 2). For SMA-725TM the polymer formed a particle having an average particle diameter of $\approx$ 10nm despite a contour length of nearly 350nm (Table 2).

Additionally, and also from our dynamic light scattering results, it was observed that both the Triton X-100TM and Triton X-305TM formed surfactant

aggregates having average aggregate diameters below 10nm (Table 2). By contrast, Triton X-45TM formed an aggregate having an average aggregate diameter of 90nm (Table 2). Despite all three surfactants forming nanonets with SMA-2000TM, only Triton X-45TM formed a nanonet with SMA-725TM (Fig. 2C, triangles).

From these results, if a surfactant has an average aggregate diameter that is larger than the average particle diameter of a polymer, that polymer surfactant combination will form a nanonet. Consequently, as each PG series surfactant formed an aggregate having an average aggregate diameter over at least 40nm, both SMA-725TM and SMA-2000TM were able to interact and form nanonets (Table 3).

Table 3. Nanonet Formation with PGL Series Surfactant and SMA-725 TM and SMA-2000 TM				
Surfactant	Ratio Surfactant to Polymer (wt/wt)	Control (Absorbance at 540 nm)	SMA-725 TM (Absorbance at 540 nm)	SMA-2000 TM (Absorbance at 540 nm)
PGL1	0	0.001	0.107	0.162
PGL1	0.5	0.022	0.192	0.272
PGL1	1	0.035	0.322	0.475
PGL1	1.5	0.053	0.453	0.981
PGL1	2	0.101	1.06	1.793
PGL1	5	0.195	2.333	2.245
PGL2	0	NA	0.11	0.14
PGL2	0.5	NA	0.124	0.272
PGL2	1	NA	0.141	0.22
PGL2	1.5	NA	0.14	0.223
PGL2	2	NA	0.184	0.253
PGL2	5	0.12	0.269	0.529
PGL2	10	0.562	2.063	2.249
PGL2	20	0.143	1.296	2.261
Triton X-305 TM	0	0	0.021	0.042
Triton X-305 TM	0.5	-0.001	0.051	0.031
Triton X-305 TM	1	-0.003	0.057	0.457

Table 3. Nanonet Formation with PGL Series Surfactant and SMA-725™ and SMA-2000™				
Surfactant	Ratio Surfactant to Polymer (wt/wt)	Control (Absorbance at 540 nm)	SMA-725™ (Absorbance at 540 nm)	SMA-2000™ (Absorbance at 540 nm)
Triton X-305™	2	-0.005	0.062	0.512
Triton X-305™	5	-0.002	0.068	0.02
Triton X-100™	0	-0.006	0.024	0.042
Triton X-100™	0.5	-0.009	0.047	0.075
Triton X-100™	1	-0.006	0.073	0.163
Triton X-100™	2	-0.006	0.144	1.149
Triton X-100™	5	0.008	0.131	1.728
Triton X-100™	10	-0.006	0.003	1.54
Triton X-100™	20	0	0	0.903
Triton X-45™	0	-0.01	0.022	0.043
Triton X-45™	0.5	0.017	1.333	1.781
Triton X-45™	1	0.091	1.398	1.848
Triton X-45™	2	0.139	1.724	1.866
Triton X-45™	5	0.378	1.594	NA
Triton X-45™	10	0.978	NA	NA

Example 4

Effect of Polymer Length On Nanonet Size.

To measure the size of the nanonets, we utilized dynamic light scattering. Nanonets generally had nanonet diameters smaller than the average aggregate diameter of the surfactant (Table 4). This suggests that the polymer was able to compact or partially dissociate surfactant aggregates. However, in some cases the addition of the polymers resulted in nanonets that had nanonet diameters with no change in size, or even a larger nanonet diameter when compared to the average aggregate diameter of the surfactant aggregate alone, notably in the cases of PGL 2 and Triton X-45™ (Table 4). Interestingly, these two polymers often formed different sized nanonets with the same surfactants. Surfactants with small alkyl chains (PGL4 and PGL 2, $N_{CH_2} = 12$) formed nanonets with SMA-2000™ which were 2 to 4-fold smaller than those formed by SMA-725™

(Table 4). However, and surprisingly, the longer contour length SMA-725TM polymer generally formed nanonets with smaller nanonet diameters than those formed by SMA-2000TM when surfactants with long alkyl chains were employed (Alkyl chain carbons > 12) (Table 2). This smaller nanonet size may be due to the decreased free energy requirements of a single long polymer (SMA-725TM) stabilizing a surfactant aggregate compared to the higher entropic contributions of multiple smaller polymers (SMA-2000TM) stabilizing the same surfactant aggregate.

Table 4: Dynamic Light Scattering Measurements of Various Polymer Surfactant Mixtures					
Polymer	Surfactant	Poly Dispersity Index (PDI)	Nanonet Diameter (nm)	Size Change Relative to Surfactant Aggregate*	Alkyl Chain Carbons
SMA-2000 TM	PGL 1	0.471	23.54	-49%	18
SMA-725 TM	PGL 1	0.379	24.49	-48%	18
SMA-2000 TM	PGL 2	0.269	95.38	1%	12
SMA-725 TM	PGL 2	0.188	178.4	85%	12
SMA-2000 TM	PGL 3	0.339	116.2	-62%	16
SMA-725 TM	PGL 3	0.511	24.32	-92%	16
SMA-2000 TM	PGL 4	0.52	8.074	-89%	12
SMA-725 TM	PGL 4	0.225	44.81	-36%	12
SMA-2000 TM	PGL 5	0.45	544.6	1%	18
SMA-725 TM	PGL 5	0.49	366.4	-35%	18
SMA-2000 TM	TX-100 TM	0.51	5.123	-4%	12
SMA-725 TM	TX-100 TM	ND	ND	NA	12
SMA-2000 TM	TX-305 TM	0.392	3.543	-62%	12
SMA-725 TM	TX-305 TM	ND	ND	NA	12
SMA-2000 TM	TX-45 TM	0.342	220.3	144%	12
SMA-725 TM	TX-45 TM	0.357	101.5	12%	12
SMA-2000 TM	Na-Oleate	0.426	53.24	-75%	18
SMA-725 TM	Na-Oleate	0.445	34.43	-91%	18

Example 5

This example compares polymer average particle diameter sizes to the stability of a nanonet from that polymer when the surfactant average aggregate diameter is held constant. Flocculation efficiencies (for example, rate of formation) of sodium oleate nanonets formed by SMA-2000TM and SMA-725TM were compared (Fig. 3A). Sodium oleate forms a large surfactant aggregate having an average aggregate diameter of 392 nm (Table 2), and is therefore compatible with both SMA-2000TM and SMA-725TM for nanonet formation. sodium oleate has a long alkyl chain ($N_{CH_2} = 18$). Free sodium oleate in solution will precipitate, but little flocculation or settling occurs (Fig. 3B). However, in SMA-725TM containing nanonets, upon destabilization by pH shift (Fig. 3C), the interaction between the interacting polymer and sodium oleate precipitate and rapidly aggregate. This aggregation occurs rapidly, signaling nanonet destabilization (Fig 3C). In contrast, the short chain polymer SMA-2000TM forms sodium oleate based nanonets which do not flocculate very well. This is apparent due to the comparatively slower settling of the precipitated SMA-2000TM sodium oleate based nanonets (Fig. 3D). Flocculation of destabilized nanonets was measured quantitatively by observing solution clearing after polymer destabilization (Fig. 3E). The quantitative measurements of rate and quality of flocculation, respectively, supported the qualitative observations that sodium oleate nanonets formed by a longer polymer display faster aggregation kinetics, and are therefore easier to physically remove when being used for wastewater treatment (Fig. 3E).

Example 6

Addition of Nanonets for Capture of Oil and Suspended Solids From Produced Water

In this example, flowback water samples and produced water samples were employed with nanonets. The water samples used were from typical Bakken Formation oil production activities. The water samples were measured for hydrocarbon content before and after exposure to nanonets. Nanonets

comprising SMA-2000TM and Triton X-100TM, were added to flowback water samples as well as to produced water samples. The solids in the resulting mixtures were then removed by filtration. Thereafter, the hydrocarbon content of each filtrate recovered from the treated water samples was again measured. The results are set out in Table 5 below.

Table 5: Hydrocarbon Content of Flowback and Produced Water from Bakken Formation Pre and Post Nanonet Treatment. (250 ppm nanonets, 10 micron filtration, ppb=parts per billion)				
	Flowback Water (ppb)		Produced Water (ppb)	
	PRE-NANONET ADDITION	POST-PRECIPIRATE REMOVAL	PRE-NANONET ADDITION	POST-PRECIPIRATE REMOVAL
Extractable Petroleum Hydrocarbons (C10-C19)	378,000	11,500	9,410	1,180
Extractable Petroleum Hydrocarbons (C19-C32)	276,000	11,000	8,640	1,450
Aggregate Hydrocarbons *(6-10)	1,020,000	1,430	4,44	ND

Example 7

Optimization of Nanonets for Selective Iron Remediation From Produced Water (Nanonets as a platform for selective contaminant removal)

Several different surfactants were screened to identify a suitable surfactant with which to prepare an iron selective nanonet. In addition a lipid SMA particle (Lecithin) was assayed. In a screening procedure the iron selective nanonet is identified that can selectively capture iron from water produced by unconventional oil and gas operations. Once, the surfactant is identified, the nanonet formulation may be further refined to be easily removed by dissolved air flotation and/or filtration.

Each of sodium oleate, CAPBTM, (COCAMIDOPROPYL BETAINE), PGL1, PGL2, PGL3, PGL4, PGLW, and cocoyl glycinate, were formed into nanonets using the polymer SMA-2000TM with a 5:1 wt/wt ratio (surfactant:polymer). A lipid-SMA formulation (Lecithin and SMA-2000TM) was also assayed. The

nanonets were added to solutions comprising dissolved iron at three different concentrations of divalent cations (4000 ppm, 400 ppm and 40 ppm). A first control with no nanonet added to it and a second control with KOH added to it were also tested. Two separate experiments were conducted. In the first, the solutions with nanonets or SMA-lecithin added were left overnight, then filtered. In the second, the solutions with nanonets and SMA-lecithin added were left for 10 minutes, then filtered. The controls were also filtered after the requisite elapsed time.

Sodium oleate, and polyglyceryl-10 monostearate (PGL5) displayed good targeting properties for capture of iron into SMA-2000TM based nanonets. PGL5 (PGLW) shows most consistent removal across the range of calcium concentrations. Conversely, the results when taken as a whole across all of the surfactants tested tend to display relatively better removal at 400 ppm calcium and relatively worse at 4000 ppm. The PGL series of surfactants appear to be generally insensitive to calcium concentration. Surfactants with high binding affinity for iron (e.g. cocoyl glycinate) appear to be more negatively affected by calcium ion concentration. The results are graphically displayed in Figures 4 and 5.

Example 8

Fourteen (14) different nanonets were prepared. Each nanonet comprised either SMA-2000TM or SMA-725TM and one of the following surfactants: sodium oleate, PGL2, PGL3, cocoyl glycinate. A SMA-lecithin formulation was also tested. Each of the nanonets was then added to various solutions comprising varying concentrations of Fe²⁺ and Ca²⁺ ions. In a first experiment, the treated solutions were left for 10 minutes. In a second experiment, the treated solutions were left for 12 hours. The results indicate that SMA-2000TM nanonets appear relatively better at removing iron than SMA-725TM nanonets. Increasing concentrations of divalent cations such as calcium appear to generally decrease

the iron binding capacity of nanonets. A control without nanonets and treated with only KOH was also tested. The results are graphically depicted in Figure 6.

Example 9

Four (4) nanonets were prepared using SMA-725TM as the polymer and using one of the following surfactants: PGL2, PGLW, sodium oleate, and cocoyl glycinate. A SMA-Lecithin control was also tested. Solutions comprising iron ions in a ratio of 75:25 Fe³⁺: Fe²⁺, were prepared at a concentration of 100 ppm. One of each of the four nanonets and the SMA-lethicin formulation was then added to a separate iron solution and left to sit for either 10 minutes or 12 hours. A control without nanonets and treated with only KOH was also tested. The results, graphically depicted in Figure 7 generally show that increasing the residence time of iron solution treated with nanonets generally increases the amount of iron removed.

Example 10

Six different nanonets were prepared using sodium oleate as the surfactant and three different polymers. Each polymer was used to prepare two nanonets, the first of which comprised a ratio of 1:5 wt/wt polymer:surfactant and the second comprised a ratio of 1:1 wt/wt. The results are graphically depicted in Figure 8. The figure shows generally that increasing the surfactant concentration within a nanonet increases iron removal. This suggests that iron removal is dependent, at least in part, upon surfactant concentration within the nanonet and not solely dependent on pH of a nanonet mixture.

Example 11

Nanonets comprising SMA-725TM and one of the following surfactants: PGL5, cocoyl glycinate, or sodium oleate and a SMA lecithin-formulation were prepared and each added to a separate sample of produced water. The produced water comprised 500ppm oil, >100,000ppm, total dissolved solids (TDS), and >10,000 ppm divalent cations. A first control with no nanonet and a

second control without nanonets and treated with only KOH were also tested. The results are shown in Figures 9 and 10. Generally, the results indicate that sodium oleate based nanonets provide a relatively better combination of oil removal and iron removal in real produced water samples.

Example 12

Sodium Oleate Nanonets Can Be Tuned by Small Addition of Polyglyceryl 10 Monostearate to Enhance Flotation (Nanonet OPGW)

An OPGW nanonet comprised of SMA-725TM, 75% sodium oleate, 25% polyglyceryl-10 monostearate, 1:1 wt ratio polymer:surfactant was prepared. The nanonets were then added to produced water samples having varying levels of elevated divalent cations (10 ppm iron, 50 ppm iron, and 100 ppm iron) and left for 10 min. The samples were then filtered and the iron content (Figure 11A), colour (Figure 11B) and the turbidity (Figure 11C) of the filtrate from the treated water samples were tested. A first control with no nanonet added and a second control without nanonets and treated with only KOH were also tested. The results generally indicate that the OPGW nanonet provides rapid removal of both iron and oil and gas from produced water samples and in the presence of elevated divalent cations.

Example 13

OPGW Nanonets Enhance Dissolved Air Flotation (DAF) Based Removal of Both Iron and Hydrocarbons From Produced Water.

The OPGW nanonet was prepared as set out above and added to produced water samples that were then treated by dissolved air flotation (DAF), followed by filtration. Prior to the addition of nanonets and again after filtration, the samples were tested for iron content, colour and turbidity. A control with no nanonet added was also tested. Total iron concentration of the filtrate from treated water samples was determined by the Ferrozine method. A yellow colour is indicative of dissolved hydrocarbons and colloidal iron was measured by

absorbance at 350nm. Turbidity was measured by absorbance at 540nm. The results are set out graphically in Figure 12.

Example 14

Treatment of Produced Water With OPGW Nanonets (SMA-725TM, 75% sodium oleate, 25% Polyglyceryl-10 Monostearate, 1:1 wt ratio polymer:surfactant), a first control with untreated produced water, and a second control without nanonets and only filtration were tested for metal concentration and for extractable petroleum hydrocarbons. All samples were vigorously mixed with addition of air for 10 minutes. The results are set out in Figure 13.

In the following Examples 15-21, the following methods reagents and materials were used.

Reagents and Materials Examples 15-21:

Poly(styrene-co-maleic anhydride) polymer was purchased from Jiaying Huawen Chemical Co.TM, Ltd (SMA-725TM, Lot no. S190405A01), or Cray ValleyTM (SMA-2021TM, Lot no. 204222A; SMA-2000TM, Lot no. PS16072901; and SMA-3000TM, Lot no. PS16111302). The polymers SMA-725TM and SMA-2021TM were ground to a fine powder before use. Poly(diisobutylene-*alt*-maleic acid) (DIBMATM, Lot no. 4217473) was purchased from Anatrache, while the polymers poly(styrene-co-maleic anhydride), cumene terminated (SMA-cumene terminated, Lot no. 08728HN) and poly(styrene-co-maleic anhydride), partial isobutyl ester (SMA-IBE, Lot no. 09918JN) were purchased from Sigma Aldrich.

Non-ionic surfactants 4-(1,1,3,3-tetramethylbutyl)phenyl-polyethylene glycol TritonTM X-100, Lot no. SLBX5574; TritonTM X-305 (70% in H₂O), Lot no. SLBX0207; TritonTM X-45, Lot no. SLBX0207; CAS no. 9002-93-1), sorbitan laurate (Span 20TM; CAS no. 1338-39-2, Lot no. MKCF5665), polyethylene glycol sorbitan monolaurate (Tween 20TM; CAS no. 9005-64-5, Lot no. SLBW9887) polyethylene glycol sorbitan monopalmitate (Tween 40TM; CAS no.

9005-66-7, Lot no. MKBV6853V), polyethylene glycol sorbitan monostearate (Tween 60TM; CAS no. 9005-67-8, Lot no. MKBX0810V), polyoxyethylenesorbitan tristearate (Tween 65TM; CAS no. 9005-71-1, Lot no. MKCG4378), polyethylene glycol sorbitan monooleate (Tween 80TM; CAS no. 9005-65-6, Lot no. BCBV5152), and polyethylene glycol sorbitan trioleate (Tween 85TM; CAS no. 9005-70-3, Lot no. MKCG2988), (Z)-9-octadecenoic acid sodium salt (sodium oleateTM; CAS no. 143-19-1, Lot no. SZBG1110V), *p*-toluenesulfonic acid sodium salt hydrate (sodium *p*-toluenesulfinate hydrateTM; CAS no. 207801-20-5, Lot no. MKCD1005), 1-octanesulfonic acid sodium salt monohydrate (sodium 1-octanesulfonate monohydrateTM; CAS no. 207596-29-0, Lot no. BCBW1986), 9,10-dihydro-9,10-dioxo-2-anthracenesulfonic acid sodium salt monohydrate (Anthraquinone-2-sulfonic acid sodium salt monohydrateTM; CAS no. 153277-35-1, Lot no. MKCD1028), *o*-ethylxanthic acid potassium salt (Potassium ethyl xanthogenateTM; CAS no. 140-89-6, Lot no. 44706), dodecylbenzenesulfonic acid sodium salt (Sodium dodecylbenzene sulfonateTM, CAS no. 25155-30-0, Lot no. STBH2778), and 1,2-Hexanediol (CAS no. 6920-22-5, Lot no. STBH6345) were purchased from Sigma Aldrich.

Poly(oxy-1,2-ethanediol)-alpha-isotridecyl-omega-hydroxy phosphate (Lakeland PAE 136TM, CAS no. 73038-25-2, Lot no. W475) was purchased from Lakeland Labs, Ltd. (Z)-9-Octadecenoic acid (oleic acidTM, CAS no. 112-80-0, Lot no. SJ1-111) was purchased from Vantage. Polyethylene glycol monoalkyl ether (Genapol X-080TM, CAS no. 9043-30-5) was purchased from Fluka, while lauramidopropyl betaine (TC-LAB 35TM, CAS no. 86438-78-0) and potassium cocoyl glycinate (Amin GCK 30HTM, CAS no. 301341-58-2) were purchased from Tinci. Sodium-1-dodecanesulfonate (dodecane-1-sulfonic acid sodium saltTM, CAS no. 2386-53-0, Lot no. K49293246 806) was purchased from Merck KGaA. Alkylpolyglucoside (coco glucosideTM, CAS no. 141464-42-8, Lot no.

16072-A52) was purchased from New Directions. Decaglyceryl monostearate (polyglycerol-10-stearateTM, CAS no. 79777-30-3), decaglyceryl monolaurate (polyglycerol-10-laurateTM, CAS no. 34406-66-1), (polyglycerol-10-palmitateTM), and triglyceryl monooleate (polyglycerol-10-oleateTM, CAS no. 11094-60-3) were obtained from Shandong Binzhou GIN&ING New Material Technology Co., Ltd. Sodium (C₁₀₋₁₆) benzenesulfonate (Bio Soft D-40TM, CAS no. 68081-81-2, Lot no. 8252764), sodium (C₁₄₋₁₆) olefin sulfonate (Bio Terge AS-40TM, CAS no. 68439-57-6, Lot no. 8242348), sodium lauryl sulfoacetate (Lathanol LAL PowderTM, CAS no. 1847-58-1, Lot no. 8170061), sodium lauroyl lactylate (Stepan SLL FBTM, CAS no. 13557-75-0, Lot no. 8086194), lauryl lactyl lactate (Stepan Mild L3TM, CAS no. 910661-93-7, Lot no. 8080185), alkylbenzenesulfonic acid (C₁₁₋₁₃) (Stepan Sulfonic 100TM, CAS no. 68608-88-8, Lot no. TK-265), sodium tridecyl ether sulfate (Cedepal TD-403 MFLDTM, CAS no. 25446-78-0, Lot no. 7606923), disodium cocoamphodipropionate (Amphosol 2CSFTM, CAS no. 68604-71-7, Lot no. S77072810), and isopropylamine alkylbenzenesulfonic acid (C₉₋₁₇ Br) (Ninate 411TM, CAS no. 68649-00-3, Lot no. 8082275) were purchased from Stepan Company. VDISTILL DV53TM Distilled Soy Fatty Acid (CAS no. 68308-53-2, Lot no. 829608) and VDISTILL DV 63TM Distilled Canola Fatty Acid (Lot no. 833603) were purchased from Vantage. *n*-Decyl-β-D-thiomaltoside (*n*-decyl-β-D-thiomaltopyranosideTM, CAS no. 148565-56-4, Lot no. 4216901), 1H, 1H, 2H-perfluorooctyl-β-D-maltopyranoside (octyl maltoside, fluorinatedTM CAS no. 118680-70-9, Lot no. 4217259), *n*-dodecyl-β-D-thiomaltoside (*n*-dodecyl-β-D-thiomaltopyranosideTM, CAS no. 148565-58-6, Lot no. 4216186), 1,2,5,6-tetradecyl-β-D-glucopyranoside-3,4-O-di-dodecyl-D-mannitol (perGlu-bisdodecyl mannitolTM, Lot no. 4217252), 1,2,5,6-tetradecyl-β-D-glucopyranoside-3,4-O-di-tridecyl-d-mannitol (perGlu-bistridecyl mannitolTM, Lot no. 4217317), 1,2,5,6-tetradecyl-β-D-glucopyranoside-3,4-O-Di-tetradecyl-d-

mannitol (perGlu-bistetradecyl mannitolTM, Lot no. 4217150), 3,6,9,12,15,18,21,24-octaoxahexatriacontan-1-ol (octaethylene glycol monododecyl etherTM, CAS no. 3055-98-9, Lot no. 4217388), *n*-decyl- β -D-thioglucoside (*n*-decyl- β -D-thioglucopyranosideTM, CAS no. 98854-16-1, Lot no. 127098), *n*-nonyl- β -D-thioglucoside (*n*-nonyl- β -D-thioglucopyranosideTM, CAS no. 98854-15-0, Lot no. 4216868), hexaethylene glycol monodecyl ether (decyl hexaethylene glycol etherTM, CAS no. 5168-89-0, Lot no. 4157194), *n*-heptyl- β -D-thioglucoside (*n*-heptyl- β -D-thioglucopyranosideTM, CAS no. 85618-20-8, Lot no. 4216927), *n*-octyl- β -D-thiomaltoside (*n*-octyl- β -D-thiomaltopyranosideTM, CAS no. 148616-91-5, Lot no. 4216064), decyl hexaethylene glycol ether (hexaethylene glycol mono-octyl etherTM, CAS no. 4440-54-4, Lot no. 4216205), hexaethylene glycol monododecyl ether (dodecyl hexaethylene glycol ether, CAS no. 3055-96-7, Lot no. 4215962), and dodecyl heptaethylene glycol ether (heptaethylene glycol monododecyl etherTM, CAS no. 3055-97-8, Lot no. 4216865) were purchased from Anatrace.

N-octanoyl-*L*-methionine (CAS no. 35440-75-6), *n*-decanoyl-*L*-methionine (CAS no. 51570-51-5), *n*-dodecanoyl-*L*-methionine (CAS no. 35440-74-5), *n*-octanoyl-*L*-threonine (CAS no. 91694-75-6), *n*-decanoyl-*L*-threonine (CAS no. 30664-76-7), *n*-dodecanoyl-*L*-threonine (CAS no. 14379-57-8), and *n*-dodecanoyl-*L*-histidine (CAS no. 55258-11-2) were synthesized by GreenCentre Canada.

The subsequent Anatrace surfactants may be abbreviated as following: *n*-decyl- β -D-thiomaltopyranoside as nDDTP-D335, octyl maltoside, fluorinated as PFDMP-O310F, *n*-dodecyl- β -D-thiomaltopyranoside as nDDDT-P342, perGlu-bisdodecyl mannitol as DDDM-MNA-C12, perGlu-bistridecyl mannitol as pGBTM-MNA-C13, perGlu-bistetradecyl mannitol as pGBTEM-MNA-C14, octaethylene glycol monododecyl ether as OEGMDE-O330, *n*-decyl- β -D-thioglucopyranoside as nDDTGP-D323, *n*-nonyl- β -D-thioglucopyranoside as nNDTP-N335, hexaethylene glycol mono-octyl ether as HEGMOE-H350, *n*-heptyl- β -D-

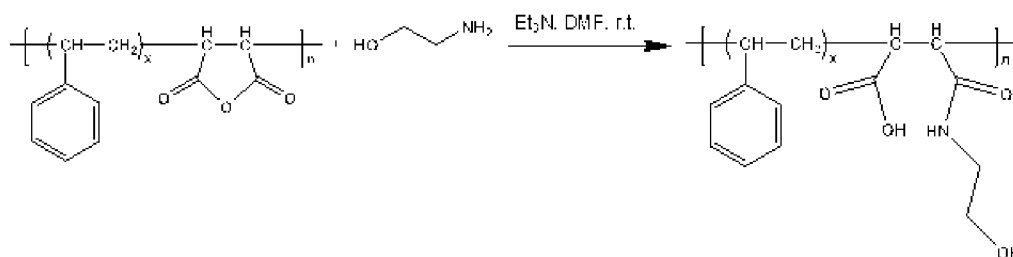
thioglucopyranoside as nHDTP-H301, *n*-octyl- β -D-thiomaltopyranoside as nODTTP-O320, hexaethylene glycol monodecyl ether as HEGMDE-H360, heptaethylene glycol monododecyl ether as HEGMDDE-H370, hexaethylene glycol monododecyl ether as HXEGMDDE-H375.

Hydrolysis of polymers:

SMA-725TM (Jiaxing Huawen Chemical Co.TM) and SMA-2021TM (Cray ValleyTM) were first ground to a fine powder, while SMA-2000TM (Cray ValleyTM), SMA-3000TM (Cray ValleyTM), DIBMA (AnatraceTM), SMA-cumene terminated (Sigma AldrichTM), and SMA-IBE (Sigma AldrichTM) were purchased in powder form.

In a 3-neck round-bottom flask connected with a condenser was added a 500mM potassium hydroxide (KOH) solution in deionized water and heated to 50 °C. To the warm KOH solution was added 5% (wt/vol) SMA-XTM (X = SMA-725TM, SMA-2000TM, SMA-3000TM, SMA-2021TM, cumene terminated, IBE) and the reaction mixture was heated to reflux at 100 °C for 3h. During reflux, the polymer dissolved completely and for the reaction with SMA-725TM and SMA-2021TM a clear yellow solution was obtained. The reaction mixture was cooled down to ambient temperature and subsequently filtered to remove non-hydrolyzed particles. The filtrate was precipitated by dropwise addition of 12 M HCl with stirring until a pH of ≤ 2 was achieved. The precipitate was filtered using a 10 μ m filter, washed 3x with 100 mL of dilute 50 mM HCl solution and 3x with 100 mL of deionized water. The isolated colorless polymer was dried at ambient temperature for 2 hours and lyophilized over night for ca. 12 hours.

Preparation of Derivatized SMA-2000TM



Scheme 1. Reaction of SMA-2000™ with ethanolamine.

In a reaction vessel with a magnetic stir bar, 0.200g SMA-2000™ powder was dissolved in 10 mL dimethylformamide to give a clear yellow solution. To this reaction mixture was added 20 μ L triethylamine and 99 μ L ethanolamine. A colour change from yellow to colourless solution was observed. The reaction mixture was stirred for 24 h at ambient temperature. Upon addition of 51 mL acidic aqueous solution a colorless suspension was obtained and the supernatant was decanted. To the colorless precipitate was added 2 mL tetrahydrofuran and the precipitate dissolved overnight and was let to evaporate over two days. Yield: 0.234g. FTIR (ATR, cm^{-1}): 3380, 3027, 2930, 2887, 1769, 1717, 1653, 1596, 1492, 1452, 1387, 1176, 1061, 1033, 916, 760, 699.

Infra-red spectroscopy

Fourier-transform infrared spectroscopy was performed on commercially available poly(styrene-co-maleic anhydride) polymers and poly(styrene-co-maleic acid) polymers using an Agilent Technologies Cary 630 spectrometer. Equipped with a diamond ATR (attenuated total reflectance). Spectra were recorded in the range of 400-4000 cm^{-1} with an average of 4 scans and a resolution of 1 cm^{-1} . The C=O maleic anhydride vibration stretching modes were observed at 1775 cm^{-1} and 1855 cm^{-1} , while the C=O maleic acid stretching mode was detected at 1705 cm^{-1} (see Killian *et al. Chemistry and Physics of Lipids* (2019), 218, 85-90).

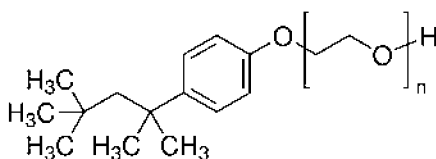
Preparation of 2% polymer stock solution:

200 mg of anhydrous polymer was suspended in deionized water and the pH adjusted to 7-8 by dropwise addition of 500mM potassium hydroxide solution. Polymer solutions were sonicated to obtain a homogenous solution and adjusted to a final volume of 10 mL. The polymer solutions were filtered using a 0.22 μm Nylon filter.

Preparation of 1% surfactant stock solution:

150mg of surfactant was dissolved into 15mL of deionized water to form separate 1% stock solutions of each surfactant. The following surfactants *N*-octanoyl-*L*-methionine, *N*-dodecanoyl-*L*-methionine, *N*-decanoyl-*L*-threonine, *N*-dodecanoyl-*L*-threonine, and *N*-dodecanoyl-*L*-histidine were dissolved in deionized water and the pH was adjusted to 8 to ensure complete dissolution of surfactants. The surfactant solutions were filtered using a 0.22 μm Nylon filter.

The chemical structure of model surfactant Triton™ X series is as follows in Formula (i):



Formula (i)

Triton™ X-100 with $n=9-10$; Triton™ X-305 with $n=30$; Triton™ X-45 with $n=5$.

Particle Sizing on DLS:

Particle sizing with intensity values were measured by dynamic light scattering on a Malvern Model ZEN1600 NanoSizer-S™ instrument (Malvern Instruments Inc, Westborough, MA, USA) at 25 °C with a 173° scattering angle. Number intensity values for hydrolyzed polymer and surfactant samples were obtained using a Malvern Zetasizer Nano ZS™ at 25 °C with a 173° scattering angle.

Surfactant solutions and hydrolyzed polymer solutions were prepared in 20mM Tris-HCl {Tris(hydroxymethyl)aminomethane} pH=7 buffer solution at ambient temperature and diluted in Tris-HCl buffer in concentrations ranging from 1% to 0.1%. The samples were filtered using a 0.22 μ M Nylon or 0.45 μ M Nylon filter, followed immediately by sizing in disposable cuvettes. Solutions were diluted to obtain an optimum dispersity index (< 0.5) for surfactant aggregate diameter and polymer particle diameter measurements. Surfactant nDDTGP-D323 was prepared in 20mM TRIS pH 7 and methanol (1:1 ratio). In one instance, intensity values were taken as surfactant aggregate or polymer particle diameter. In another instance, number values were taken as surfactant aggregate or polymer particle diameter.

Table 6. Dynamic Light Scattering Characterization using Intensity Values of Anionic Surfactant Aggregates					
Surfactant	Alkyl Chain Length	Head Group	Concentration	Aggregate Diameter [nm] (Intensity Mean d)	Poly Dispersity Index (PDI)
Sodium 1-Octane Sulfonate	8	Sulfonate	10%	1.562	0.367
Amin GCK 30H	18, 8	Acyl glycinate	2%	7.286	0.3
Anthraquinone-2-sulfonic mono hydrate, sodium	0	Sulfonate	1%	1.008	0.343
Stepan Bio Soft D-40™	0	Sulfonate	1%	5.13	0.322
Stepan Bio Terge AS-40™	14	Sulfonate	0.50%	6.434	0.181
Cedepal TD-403 MFLD™	13	Ether sulfate	0.50%	6.421	0.235
Dodecane-1-sulfonic acid	12	Sulfonate	0.50%	5.108	0.227
Lakeland PAE 136	13	Phosphate ester	0.25%	7.924	0.161
Lathanol LAL™	12	Sulfoacetate	1%	6.814	0.151
n-Decanoyl-L-Methionine	10	Amino acid	1%	85.47	0.172

Table 6. Dynamic Light Scattering Characterization using Intensity Values of Anionic Surfactant Aggregates					
Surfactant	Alkyl Chain Length	Head Group	Concentration	Aggregate Diameter [nm] (Intensity Mean d)	Poly Dispersity Index (PDI)
n-Decanoyl-L-Threonine	10	Amino acid	0.50%	90.68	0.213
n-Dodecanoyl-L-Methionine	12	Amino Acid	1%	2.32	0.138
n-Dodecanoyl-L-Histidine	12	Amino acid	1%	99.96	0.249
n-Dodecanoyl-L-Threonine	12	Amino acid	0.50%	99.77	0.203
n-Octanoyl-L-Methionine	8	Amino Acid	1%	116.6	0.239
n-Octanoyl-L-Threonine	8	Amino acid	1%	119	0.372
Sodium Dodecylbenzene Sulfonate	12	Sulfonate	2%	4.235	0.53
Ninate 411™	12	Sulfonate	0.50%	216.2	0.288
p-Toluene Sulfonic Acid	1	Sulfonate	10%	0.8236	0.32
Potassium Ethyl Xanthogenate	2	Xanthogenate	10%	190.7	0.281
Sodium-1-Pentasulfonate	5	Sulfonate	0.25	0.8036	0.573
Stepan Mild L3™	12	Lactylate	1%	372.9	0.067
Stepan SLL FB™	12	Lactylate	0.10%	136.3	0.144
Stepan Sulfonic Acid™	2	Sulfonate	2%	2.821	0.186

Table 7. Dynamic Light Scattering Characterization using Intensity Values of Non-ionic Surfactant Aggregates							
Surfactant	Alkyl Chain Length	Head Group	HLB	Concentration	CMC [mM]	Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
Apo 10	10	Phosphine oxide	NA	0.50%	NA	5.835	0.259
Apo 12	12	Phosphine oxide	NA	0.50%	NA	82.41	0.272
Coco Gglucoside	18	Glucopyranoside	13.1	2%	NA	27.32	0.154
Fluoro Octyl Maltoside O310F	8	Maltoside-sugar	NA	0.50%	NA	23.83	0.172
Genapol X-080	13	Poly ethylene glycol	12	0.10%	0.06-0.15	10.58	0.147
HEGMDE H370	12	Ethylene glycol	NA	1%	0.069	8.218	0.14
HEGMDE H360	10	Ethylene glycol	NA	0.50%	0.9	6.033	0.158
HEGMOE H350	8	Ethylene glycol	NA	2%	10	5.505	0.25
HXEGMDDE H375	12	Ethylene glycol	NA	1%	NA	9.351	0.079
MNA C12	12	Glucopyranoside	NA	0.50%	0.004	6.122	0.068
MNA C13	13	Glucopyranoside	NA	0.50%	0.002	6.423	0.046
MNA C14	14	Mannitol	NA	0.50%	0.001	7.548	0.217
nDDTP D342	12	Thiomaltopyranoside	NA	0.50%	0.05	11.67	0.131
nDDTGP D323	10	Thioglucopyranoside	NA	1%	0.9	262.2	0.176
nDDTP D335	10	Thiomaltopyranoside	NA	0.50%	0.9	6.881	0.078
nHDTTP H301	7	Thioglucopyranoside	NA	3%	29	8.935	0.011
nNDTP N335	9	Thioglucopyranoside	NA	0.50%	2.9	137	0.357
nODTTP O320	8	Thiomaltopyranoside	NA	2%	8.5	6.022	0.324
OEGMDE O330	12	Ethylene glycol	NA	0.50%	0.09	8.099	0.165
Polyglycerol-10-Laurate	12	Polyglycerol fatty acid ester	12	1%	NA	114.8	0.226

Table 7. Dynamic Light Scattering Characterization using Intensity Values of Non-ionic Surfactant Aggregates							
Surfactant	Alkyl Chain Length	Head Group	HLB	Concentration	CMC [mM]	Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
Polyglycerol-10-Oleate	18	Polyglycerol fatty acid ester	10.5	0.50%	NA	358	0.254
Polyglycerol-10-Palmitate	16	Polyglycerol fatty acid ester	10.5	0.50%	NA	111.8	0.226
Polyglycerol-10-Stearate	18	Polyglycerol fatty acid ester	11	0.50%	NA	112.3	0.172
Span 20™	12	Sorbitan	8.6	0.10%	NA	1061	0.472
Triton X-100™	8	Ethylene glycol	13.4	0.50%	0.24	10.07	0.089
Triton X-305™	8	Ethylene glycol	17.3	0.50%	0.65	10.66	0.181
Triton X-45™	8	Ethylene glycol	9.8	0.10%	0.1	156.3	0.325
Tween 20™	12	Polysorbate	16.7	0.50%	0.05	9.173	0.072
Tween 40™	16	Polysorbate	15.6	0.50%	NA	9.691	0.079
Tween 60™	18	Polysorbate	14.9	0.10%	0.022	134.2	0.637
Tween 65™	18	Polysorbate	10.5	0.50%	NA	114.5	0.286
Tween 80™	18	Polysorbate	15	0.50%	0.012	10.42	0.026
Tween 85™	16	Polysorbate	11	0.50%	NA	189.8	0.252

Table 8. Dynamic Light Scattering Characterization using Intensity and Number Values of Hydrolyzed Polymer Aggregates						
	Calculated Aggregate Diameter [nm]	Concentration	Measured Aggregate Diameter [nm] (Intensity Mean d)	Poly Dispersity Index (PDI)	Aggregate Diameter [nm] (Number Mean d)	Poly Dispersity Index (PDI)
Polymer						
SMA-2000™	2.6	0.10%	5.242	0.113	4.633	0.292

Table 8. Dynamic Light Scattering Characterization using Intensity and Number Values of Hydrolyzed Polymer Aggregates							
Polymer	Calculated Aggregate Diameter [nm]	Concentration	Measured Aggregate Diameter [nm] (Intensity Mean d)	Poly Dispersity Index (PDI)	Concentration	Aggregate Diameter [nm] (Number Mean d)	Poly Dispersity Index (PDI)
SMA-2021™	3.7	0.25%	6.627	0.274	0.10%	8.383	0.169
SMA-3000™	2.8	0.25%	3.713	0.213	0.01%	4.68	0.355
SMA-725™	6.8	0.10%	15.03	0.219	0.10%	10.16	0.283
SMA-cumene terminated	1.5	0.10%	7.123	0.164	0.50%	3.325	0.297
SMA-IBE	5.4	0.25%	9.759	0.209	0.01%	4.256	0.869
DIBMA	3.2	0.50%	76.74*	0.135	0.10%	5.704	0.271
*DIBMA sample in 20mM Tris pH=14. DIBMA molecular weight 10,000-12,000 g/mol.							

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants						
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
Apo 10	5.835	DIBMA	92.7	3.2	69.11	0.178
Apo 12	82.41	DIBMA	92.7	3.2	57.88	0.446
Coco Glucoside	27.32	DIBMA	92.7	3.2	84.87	0.245

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
Genapol X 080	10.58	DIBMA	92.7	3.2	91.23	0.005	0.252
HEGMDDE H370	8.218	DIBMA	92.7	3.2	29.05	0.005	0.528
HEGMDE H360	6.033	DIBMA	92.7	3.2	110.2	0.005	0.351
HXEGMDDE H375	9.351	DIBMA	92.7	3.2	62.11	0.005	0.268
DDDM-MNA-C12	6.122	DIBMA	92.7	3.2	84.44	0.005	0.407
pGBTM-MNA-C13	6.423	DIBMA	92.7	3.2	126.4	0.005	0.578
pGBTEM-MNA-C14	7.548	DIBMA	92.7	3.2	89.21	0.005	0.597
nDDDTp D342	11.67	DIBMA	92.7	3.2	96.78	0.005	0.305
nDDTGP D323	262.2	DIBMA	92.7	3.2	93.11	0.005	0.234
OEGMDE O330	8.099	DIBMA	92.7	3.2	79.04	0.005	0.175

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
Polyglycerol-10-mono-laurate	114.8	DIBMA	92.7	3.2	81.35	0.005	0.246
Polyglycerol-10-mono-oleate	358	DIBMA	92.7	3.2	287.3	0.005	0.279
Span 20™	1061	DIBMA	92.7	3.2	117.3	0.005	0.149
Triton X-100™	10.07	DIBMA	92.7	3.2	120.9	0.005	0.334
Triton X-45™	156.3	DIBMA	92.7	3.2	84.06	0.005	0.311
Tween 20™	9.173	DIBMA	92.7	3.2	87.81	0.005	0.278
Tween 65™	114.5	DIBMA	92.7	3.2	89.57	0.005	0.229
Tween 85™	189.8	DIBMA	92.7	3.2	155.7	0.005	0.246
Apo 10	5.835	SMA-2000™	5.242	2.6	4.889	0.005	0.188
Apo12	82.41	SMA-2000™	5.242	2.6	4.387	0.005	0.421
Coco Glucoside	27.32	SMA-2000™	5.242	2.6	7.668	0.0025	0.467
Genapol X 080	10.58	SMA-2000™	5.242	2.6	107.6	0.005	0.165
HEGMDDDE H370	8.218	SMA-2000™	5.242	2.6	4.938	0.005	0.419

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
HEGMDE H360	6.033	SMA-2000™	5.242	2.6	3.87	0.0025	0.147
HEGMOE H350	5.505	SMA-2000™	5.242	2.6	92.13	0.005	0.146
HXEGMDDE H375	9.351	SMA-2000™	5.242	2.6	179	0.0025	0.26
pGBTM-MNA-C13	6.423	SMA-2000™	5.242	2.6	6.47	0.00125	0.221
pGBTEM-MNA-C14	7.548	SMA-2000™	5.242	2.6	6.38	0.005	0.404
nDDDTp D342	11.67	SMA-2000™	5.242	2.6	113.3	0.0025	0.211
nDDTGp D323	262.2	SMA-2000™	5.242	2.6	143.3	0.005	0.264
nDDTP D335	6.881	SMA-2000™	5.242	2.6	168.3	0.005	0.249
OEGMDE O330	8.099	SMA-2000™	5.242	2.6	121.9	0.0025	0.236
Polyglycerol-10-mono-laurate	114.8	SMA-2000™	5.242	2.6	101.2	0.005	0.3

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
Polyglycerol-10-mono-oleate	358	SMA-2000™	5.242	2.6	123.1	0.005	0.172
Span 20™	1061	SMA-2000™	5.242	2.6	97.93	0.005	0.293
Triton X-100™	10.07	SMA-2000™	5.242	2.6	139.7	0.00125	0.188
Triton X-305™	10.66	SMA-2000™	5.242	2.6	121.7	0.0025	0.468
Triton X-45™	156.3	SMA-2000™	5.242	2.6	90.85	0.0025	0.201
Tween 20™	9.173	SMA-2000™	5.242	2.6	97.91	0.0025	0.389
Tween 40™	9.691	SMA-2000™	5.242	2.6	133	0.0025	0.605
Tween 60™	134.2	SMA-2000™	5.242	2.6	6.555	0.00125	0.173
Tween 65™	114.5	SMA-2000™	5.242	2.6	164	0.0025	0.25
Tween 80™	10.42	SMA-2000™	5.242	2.6	130	0.0025	0.416
Tween 85™	189.8	SMA-2000™	5.242	2.6	9.903	0.00125	0.4
Apo 10	5.835	SMA-2021™	6.627	3.7	8.087	0.005	0.142
Apo 12	82.41	SMA-2021™	6.627	3.7	11.56	0.005	0.166
Coco Glucoside	27.32	SMA-2021™	6.627	3.7	13.26	0.005	0.323
HEGMDDE H370	8.218	SMA-2021™	6.627	3.7	10.53	0.005	0.298

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
HEGMDE H360	6.033	SMA-2021™	6.627	3.7	8.217	0.0025	0.514
HXEGMDDE H375	9.351	SMA-2021™	6.627	3.7	6.601	0.005	0.309
pGBTM-MNA-C13	6.423	SMA-2021™	6.627	3.7	8.065	0.005	0.104
pGBTEM-MNA-C14	7.548	SMA-2021™	6.627	3.7	7.101	0.005	0.16
nDDTGP D323	262.2	SMA-2021™	6.627	3.7	8.654	0.0025	0.248
nDDDTTP D342	11.67	SMA-2021™	6.627	3.7	8.19	0.0025	0.278
OEGMDE O330	8.099	SMA-2021™	6.627	3.7	5.706	0.005	0.27
Polyglycerol-10-mono-laurate	114.8	SMA-2021™	6.627	3.7	22.34	0.005	0.238
Polyglycerol-10-mono-oleate	358	SMA-2021™	6.627	3.7	120.7	0.005	0.276

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
Triton X-100™	10.07	SMA-2021™	6.627	3.7	5.144	0.005	0.152
Triton X-45™	156.3	SMA-2021™	6.627	3.7	6.222	0.005	0.093
Tween 40™	9.691	SMA-2021™	6.627	3.7	9.262	0.005	0.337
Tween 60™	134.2	SMA-2021™	6.627	3.7	8.687	0.00125	0.317
Tween 80™	10.42	SMA-2021™	6.627	3.7	10.39	0.005	0.441
Tween 85™	189.8	SMA-2021™	6.627	3.7	121.4	0.005	0.304
Apo 10	5.835	SMA-3000™	3.713	2.8	163.1	0.005	0.252
Apo 12	82.41	SMA-3000™	3.713	2.8	221	0.005	0.293
Coco Glucoside	27.32	SMA-3000™	3.713	2.8	16.23	0.005	0.199
Genapol X080	10.58	SMA-3000™	3.713	2.8	53.86	0.005	0.429
HEGMDDE H370	8.218	SMA-3000™	3.713	2.8	43.06	0.005	0.137
HEGMDE H360	6.033	SMA-3000™	3.713	2.8	79.51	0.005	0.457
HXEGMDDE H375	9.351	SMA-3000™	3.713	2.8	79.11	0.005	0.498
DDDM-MNA-C12	6.122	SMA-3000™	3.713	2.8	6.911	0.005	0.151

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
pGBTM-MNA-C13	6.423	SMA-3000™	3.713	2.8	7.306	0.005	0.212
pGBTEM-MNA-C14	7.548	SMA-3000™	3.713	2.8	7.266	0.005	0.085
nDDDTp D342	11.67	SMA-3000™	3.713	2.8	10.04	0.005	0.241
nDDTGP D323	262.2	SMA-3000™	3.713	2.8	201.3	0.005	0.466
OEGMDE O330	8.099	SMA-3000™	3.713	2.8	27	0.005	0.404
Polyglycerol-10-mono-laurate	114.8	SMA-3000™	3.713	2.8	20.32	0.005	0.131
Polyglycerol-10-mono-oleate	358	SMA-3000™	3.713	2.8	218.7	0.005	0.263
Polyglycerol-10-mono-palmitate	111.8	SMA-3000™	3.713	2.8	165.4	0.0005	0.391
Span 20™	1061	SMA-3000™	3.713	2.8	102.8	0.005	0.175

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants						
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
Triton X-100™	10.07	SMA-3000™	3.713	2.8	181.2	0.295
Triton X-45™	156.3	SMA-3000™	3.713	2.8	249	0.318
Tween 60™	114.5	SMA-3000™	3.713	2.8	356.4	0.806
Tween 85™	189.8	SMA-3000™	3.713	2.8	167.8	0.422
Apo 12	82.41	SMA-IBE	9.759	5.4	11	0.31
Coco Glucoside	27.32	SMA-IBE	9.759	5.4	9.499	0.236
HEGMDDE H370	8.218	SMA-IBE	9.759	5.4	8.579	0.189
HEGMDE H360	6.033	SMA-IBE	9.759	5.4	11.05	0.288
HXEGMDDE H375	9.351	SMA-IBE	9.759	5.4	12.07	0.278
nDDDTp D342	11.67	SMA-IBE	9.759	5.4	13.19	0.326
nDDTP D335	6.881	SMA-IBE	9.759	5.4	13.2	0.311
Polyglycerol-10-mono-oleate	358	SMA-IBE	9.759	5.4	95.55	0.52
Triton X-45™	156.3	SMA-IBE	9.759	5.4	6.882	0.292

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
Tween 85™	189.8	SMA-IBE	9.759	5.4	82.56	0.00125	0.201
Genapol X 080	10.58	SMA-725™	15.03	6.8	19.55	0.005	0.319
nDDTP D342	11.67	SMA-725™	15.03	6.8	9.736	0.005	0.301
nDDTGP D323	262.2	SMA-725™	15.03	6.8	12.29	0.005	0.336
Polyglycerol-10-mono-oleate	358	SMA-725™	15.03	6.8	240.5	0.005	0.451
Span 20™	1061	SMA-725™	15.03	6.8	98.31	0.005	0.561
HEGMDDE H370	8.218	SMA-725™	15.03	6.8	14.64	0.0025	0.231
HEGMDE H360	6.033	SMA-725™	15.03	6.8	11.75	0.0025	0.111
HXEGMDDE H375	9.351	SMA-725™	15.03	6.8	11.44	0.005	0.437
pGBTEM-MNA-C14	7.548	SMA-725™	15.03	6.8	55.4	0.005	0.429

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
Polyglycerol-10-mono-palmitate	111.8	SMA-725™	15.03	6.8	14.62	0.0025	0.234
Triton X-100™	10.07	SMA-725™	15.03	6.8	10.5	0.005	0.475
Triton X-45™	156.3	SMA-725™	15.03	6.8	14.02	0.0025	0.328
Tween 65™	114.5	SMA-725™	15.03	6.8	14.71	0.0025	0.233
Tween 80™	10.42	SMA-725™	15.03	6.8	9.102	0.005	0.608
Apo 10	5.835	SMA-Cumene terminated	7.123	1.5	4.232	0.005	0.14
Apo 12	82.41	SMA-Cumene terminated	7.123	1.5	4.923	0.005	0.266
PFDMP O310F	23.83	SMA-Cumene terminated	7.123	1.5	4.484	0.005	0.228
Genapol X080	10.58	SMA-Cumene terminated	7.123	1.5	5.366	0.005	0.25

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
HEGMDDE H370	8.218	SMA-Cumene terminated	7.123	1.5	4.557	0.005	0.069
HEGMDE H360	6.033	SMA-Cumene terminated	7.123	1.5	4.55	0.005	0.398
HXEGMDDE H375	9.351	SMA-Cumene terminated	7.123	1.5	5.153	0.005	0.118
DDDM-MNA-C12	6.122	SMA-Cumene terminated	7.123	1.5	4.801	0.005	0.238
pGBTM-MNA-C13	6.423	SMA-Cumene terminated	7.123	1.5	4.561	0.005	0.237
nDDTTP D342	11.67	SMA-Cumene terminated	7.123	1.5	4.652	0.005	0.224
nDDTGP D323	262.2	SMA-Cumene terminated	7.123	1.5	4.424	0.005	0.221

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
nDDTP D335	6.881	SMA-Cumene terminated	7.123	1.5	4.659	0.005	0.248
OEGMDE O330	8.099	SMA-Cumene terminated	7.123	1.5	4.706	0.005	0.232
Polyglycerol-10-mono-laurate	114.8	SMA-Cumene terminated	7.123	1.5	4.869	0.005	0.248
Polyglycerol-10-mono-oleate	358	SMA-Cumene terminated	7.123	1.5	127.9	0.005	0.305
Span 20™	1061	SMA-Cumene terminated	7.123	1.5	91.93	0.005	0.292
Triton X-100™	10.07	SMA-Cumene terminated	7.123	1.5	4.643	0.005	0.228
Triton X-45™	156.3	SMA-Cumene terminated	7.123	1.5	5.305	0.005	0.103

Table 9. Dynamic Light Scattering Characterization of Nanonet Aggregates with Nonionic Surfactants						
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
Tween 65™	114.5	SMA-Cumene terminated	7.123	1.5	4.677	0.276
Tween 85™	189.8	SMA-Cumene terminated	7.123	1.5	8.387	0.349

Table 10. Dynamic Light Scattering Characterization of Nanonet Aggregates with Anionic Surfactants						
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
Stepan Bio-Soft D-40™	5.13	DIBMA	92.7	3.2	70.62	0.5
Lakeland PAE136	7.924	DIBMA	92.7	3.2	93.72	0.197
Stepan Lathanol LAL™	6.814	DIBMA	92.7	3.2	83.64	0.32

Table 10. Dynamic Light Scattering Characterization of Nanonet Aggregates with Anionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
n-Decanoyl-L-Threonine	90.68	DIBMA	92.7	3.2	92.49	0.005	0.283
n-Dodecanoyl-L-Threonine	99.77	DIBMA	92.7	3.2	98.7	0.005	0.259
n-Decanoyl-L-Methionine	85.47	DIBMA	92.7	3.2	95.33	0.005	0.246
n-Dodecanoyl-L-Methionine	2.312	DIBMA	92.7	3.2	104.2	0.005	0.34
Stepan Ninat 411™	216.2	DIBMA	92.7	3.2	163.6	0.005	0.196
n-Octanoyl-L-Methionine	116.6	DIBMA	92.7	3.2	95	0.005	0.3
Stepan Mild L3™	372.9	DIBMA	92.7	3.2	92.22	0.005	0.185
Stepan Bio-Terge AS-40™	6.434	SMA 2000™	5.242	2.6	132.7	0.0025	0.452
Stepan Bio-Soft D-40™	5.13	SMA 2000™	5.242	2.6	132.5	0.0025	0.354
Dodecane-1-sulfonic acid sodium salt	5.108	SMA 2000™	5.242	2.6	8.054	0.000625	0.454

Table 10. Dynamic Light Scattering Characterization of Nanonet Aggregates with Anionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
Lakeland PAE 136	7.924	SMA 2000™	5.242	2.6	143	0.0025	0.203
Stepan Lathanol LAL™	6.814	SMA 2000™	5.242	2.6	6.541	0.005	0.153
n-Decanoyl-L-Methionine	85.47	SMA 2000™	5.242	2.6	82.64	0.00125	0.399
n-Dodecanoyl-L-Methionine	2.312	SMA 2000™	5.242	2.6	5.89	0.0025	0.203
n-Decanoyl-L-Threonine	90.68	SMA 2000™	5.242	2.6	152.4	0.00125	0.347
n-Dodecanoyl-L-Histidine	99.96	SMA 2000™	5.242	2.6	94.7	0.005	0.288
Stepan Mild L3™	372.9	SMA 2000™	5.242	2.6	58.51	0.0025	0.494
n-Dodecanoyl-L-Threonine	99.77	SMA 2000™	5.242	2.6	124.6	0.005	0.302
Sodium dodecylbenzene sulfonate	4.235	SMA 2000™	5.242	2.6	2.775	0.005	0.458
Stepan Ninat 411™	216.2	SMA 2000™	5.242	2.6	131.4	0.0025	0.435

Table 10. Dynamic Light Scattering Characterization of Nanonet Aggregates with Anionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
Stepan SLL-FB™	136.3	SMA 2000™	5.242	2.6	147.7	0.0025	0.312
Stepan Sulfonic acid™	2.821	SMA 2000™	5.242	2.6	5.508	0.0025	0.416
Sodium dodecane-1-sulfonic acid salt	5.108	SMA 2021™	6.627	3.7	8.378	0.0025	0.361
Lakeland PAE 136™	7.924	SMA 2021™	6.627	3.7	6.351	0.0025	0.235
Stepan Lathanol LAL™	6.814	SMA 2021™	6.627	3.7	4.209	0.005	0.393
n-Decanoyl-L-Methionine	85.47	SMA 2021™	6.627	3.7	8.205	0.0025	0.341
Cedepal TD-403 MFLD™	6.421	SMA 2021™	6.627	3.7	7.653	0.0025	0.311
Stepan Mild L3™	372.9	SMA 2021™	6.627	3.7	4.174	0.0025	0.42
n-Dodecanoyl-L-Methionine	2.312	SMA 2021™	6.627	3.7	7.181	0.0025	0.406
n-Decanoyl-L-Methionine	85.47	SMA 3000™	3.713	2.8	173.2	0.005	0.179
n-Dodecanoyl-L-Methionine	2.312	SMA 3000™	3.713	2.8	185.3	0.005	0.24

Table 10. Dynamic Light Scattering Characterization of Nanonet Aggregates with Anionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
n-Decanoyl-L-Threonine	90.68	SMA 2021™	6.627	3.7	129.3	0.0025	0.41
n-Dodecanoyl-L-Threonine	99.77	SMA 2021™	6.627	3.7	8.825	0.005	0.332
Sodium dodecylbenzene sulfonate	4.235	SMA 2021™	6.627	3.7	6.732	0.005	0.147
Stepan Ninat 411™	216.2	SMA 2021™	6.627	3.7	27.73	0.005	0.397
Stepan SLL-FB™	136.3	SMA 2021™	6.627	3.7	8.91	0.005	0.151
Stepan Sulfonic acid™	2.821	SMA 2021™	6.627	3.7	7.86	0.005	0.153
Lakeland PAE 136	7.924	SMA 3000™	3.713	2.8	127.7	0.005	0.153
n-Dodecanoyl-L-Threonine	99.77	SMA 3000™	3.713	2.8	126.3	0.005	0.155
Stepan Ninat 411™	216.2	SMA 3000™	3.713	2.8	104.6	0.005	0.269
Stepan Mild L3™	372.9	SMA 3000™	3.713	2.8	139.1	0.005	0.233

Table 10. Dynamic Light Scattering Characterization of Nanonet Aggregates with Anionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
Stepan SLL-FB™	136.3	SMA 3000™	3.713	2.8	7.09	0.005	0.24
Stepan Sulfonic acid™	2.821	SMA 3000™	3.713	2.8	178.8	0.005	0.381
n-Decanoyl-L-Methionine	85.47	SMA IBE	9.759	5.4	14.43	0.005	0.29
n-Dodecanoyl-L-Methionine	2.312	SMA IBE	9.759	5.4	13.37	0.005	0.323
Stepan Mild L3™	372.9	SMA IBE	9.759	5.4	13.98	0.005	0.29
Stepan SLL-FB™	136.3	SMA IBE	9.759	5.4	13.59	0.005	0.3
Amin GCK30H	7.286	SMA 725™	15.03	6.8	126.6	0.00125	0.65
Stepan Bio-Terge AS-40™	6.434	SMA 725™	15.03	6.8	131.1	0.00125	0.689
n-Dodecanoyl-L-Histidine	99.96	SMA 725™	15.03	6.8	92.95	0.00125	0.555
Stepan Lathanol LAL™	6.814	SMA 725™	15.03	6.8	122.7	0.0025	0.647
n-Decanoyl-L-Methionine	85.47	SMA 725™	15.03	6.8	107.2	0.0025	0.538
n-Dodecanoyl-L-Methionine	2.312	SMA 725™	15.03	6.8	124.3	0.005	0.692

Table 10. Dynamic Light Scattering Characterization of Nanonet Aggregates with Anionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
Sodium dodecane-1-sulfonic acid salt	5.108	SMA 725™	15.03	6.8	126	0.005	0.682
Lakeland 136 PAE	7.924	SMA 725™	15.03	6.8	132.1	0.0025	0.658
n-Dodecanoyl-L-Threonine	99.77	SMA 725™	15.03	6.8	126.1	0.005	0.687
Stepan Mild L3™	372.9	SMA 725™	15.03	6.8	102.9	0.0025	0.601
Stepan SLL-FB™	136.3	SMA 725™	15.03	6.8	99.11	0.0025	0.577
Lakeland 136 PAE	7.924	SMA Cumene terminated	7.123	1.5	4.65	0.005	0.232
n-Dodecanoyl-L-Threonine	99.77	SMA Cumene terminated	7.123	1.5	4.464	0.005	0.235
Stepan Mild L3™	372.9	SMA Cumene terminated	7.123	1.5	4.621	0.005	0.236
Stepan SLL-FB™	136.3	SMA Cumene terminated	7.123	1.5	4.551	0.005	0.229

Table 10. Dynamic Light Scattering Characterization of Nanonet Aggregates with Anionic Surfactants							
Surfactant	Surfactant Aggregate Diameter [nm] (Intensity Mean d)	Polymer	Polymer Aggregate Diameter [nm]	Calculated Polymer Aggregate Diameter [nm]	Nanonet Aggregate Diameter [nm]	Concentration	Poly Dispersity Index (PDI)
n-Decanoyl-L-Methionine	85.47	SMA Cumene terminated	7.123	1.5	3.699	0.005	0.288
n-Dodecanoyl-L-Methionine	2.312	SMA Cumene terminated	7.123	1.5	3.903	0.005	0.122
n-Dodecanoyl-L-Histidine	99.96	SMA Cumene terminated	7.123	1.5	83.07	0.0025	0.403

Gel Permeation Chromatography (GPC)

The polymer molecular weight distribution analysis of commercially available poly(styrene-co-maleic anhydride) polymers were determined by gel permeation chromatography (GPC) on a Viscotek GPCmax VE-2001 using a conventional calibration of polystyrene standards at 2 mg/mL using tetrahydrofuran (THF) as eluent at 1 mL/min. Before injection, the samples were filtered using a 0.22 μ m Nylon filter. The measurements were carried out by GreenCentre Canada.

Table 11. Molecular Weight Distribution Data (M_n , M_w , M_z , M_p , $\bar{D}$) of Different Polymer Samples					
	SMA-725™	SMA-2000™	SMA-2021™	SMA-3000™	SMA-Cumene terminated
M_n - (Daltons)	48,216	3,268	10,029	4,667	3,983
M_w - (Daltons)	97,987	5,793	23,727	8,767	7,708
M_z - (Daltons)	170,089	9,020	84,311	14,042	12,598
M_p - (Daltons)	81,853	4,839	17,096	7,648	6,486
$\bar{D}$ (M_w/M_n)	2.032	1.773	2.366	1.879	1.935

Nanonet Formation Experiment – Method #1

To each well of a 96 well plate was added 200 ppm of 2% polymer stock solution followed by addition of a 1% surfactant stock solution selected from one of the following six concentrations (1000 ppm, 500 ppm, 250 ppm, 125 ppm, 50 ppm, and 0 ppm). The polymer and surfactant mixture were diluted in deionized water to a final volume of 200 μ L in each well. To the mixture was added 10 μ L of 12M hydrochloric acid (HCl) as precipitation agent. Nanonet formation was observed based on degree of turbidity of the mixture in each well. The degree of turbidity (an indicator of nanonet formation) was determined by absorbance measurements at 540 nm on a Molecular Devices-SPECTRAmax M2. Each surfactant and polymer combination was run in triplicate.

Nanonet Formation Efficiency – Method #2

Method #2 describes the nanonet complexation efficiency of Triton™ X-containing surfactants (Triton™ X-100, Triton™ X-305, and Triton™ X-45) with SMA-containing polymers (SMA-725™, SMA-2000™, SMA-2021™, SMA-3000™). The purpose was to identify the surfactant loading capacity of the polymer and to study the effect of the length of polymer. For the range of surfactant concentrations employed, a two-fold ratio of the Nanonet formation factor (Nf) for each selected surfactant and selected polymer was used. A total of six concentrations were selected. The Nanonet formation factor (Nf) were calculated by method #4 and are highlighted below. For each titration, the polymer concentration was 400 ppm (20 µL).

Nanonet Titration:

Titration of surfactant and polymer at given concentrations (see below) was performed using 1.5 mL centrifuge tubes, followed by dilution in deionized water to a final volume of 1.0 mL. The resulting nanonet was obtained by the addition of 10 µL 12M HCl. The titration was performed in triplicate. In addition, a surfactant control in deionized water and a separate control containing a mixture of surfactant and HCl were prepared. A portion (ca. 200 µL) of the prepared nanonet, surfactant control, and surfactant control with HCl were transferred into a 96-well plate and the absorption was measured at 540 nm. The remaining solution of the prepared nanonets, surfactant control with and without acid were sedimented using a Sigma™ 4-16 centrifuge at 2300 rpm (816 rcf) for 10 min. The supernatant (ca. 200 µL) was transferred into a 96-well UV plate and the absorption was measured at 280 nm. An average of the absorption values was recorded.

These are the following surfactant and polymer concentrations employed: SMA-2000™ (400 ppm, 20 µL), Triton™ X-100 (1000, 500, 250, 100, 64.8, 0 ppm); SMA-725™ (400 ppm, 20 µL), Triton™ X-100 (1000, 500, 295.16, 250, 100, 0 ppm); SMA-3000™ (400 ppm, 20 µL), Triton™ X-100 (1000, 500, 250,

100, 13.4, 0 ppm); SMA-2021™ (400 ppm, 20 µL), Triton™ X-100 (1000, 500, 250, 91.22, 0 ppm).

SMA-2000™ (400 ppm, 20 µL), Triton™ X-305 (1000, 500, 250, 231.58, 100, 0 ppm); SMA-725™ (400 ppm, 20 µL), Triton™ X-305 (1000, 500, 473.76, 250, 100, 0 ppm); SMA-3000™ (400 ppm, 20 µL), Triton™ X-305 (1000, 500, 447.88, 250, 100, 0 ppm); SMA-2021™ (400 ppm, 20 µL), Triton™ X-305 (1000, 500, 436.88, 250, 100, 0 ppm).

SMA-2000™ (400 ppm, 20 µL), Triton™ X-45 (1000, 616.18, 500, 250, 100, 0 ppm); SMA-725™ (400 ppm, 20 µL), Triton™ X-45 (1000, 500, 250, 100, 92.92, 0 ppm); SMA-3000™ (400 ppm, 20 µL), Triton™ X-45 (1000, 500, 250, 99.7, 0 ppm); SMA-2021™ (400 ppm, 20 µL), Triton™ X-305 (1000, 500, 250, 158.4, 100, 0 ppm).

Zn²⁺ and Cu²⁺ Metal Ion Removal from Seawater – Method #3

Surfactant and hydrolyzed polymer solutions in deionized water were mixed into 96-well plates to give a total of 1 mL 0.5% nanonet library stock solutions. 199 different nanonets were prepared. Each well of a 96- well plate contained a unique 1mL nanonet solution.

Colorimetric determination of Zinc and Copper concentrations using Zincon:

As proof of principle of the optimized nanonet library, seawater was spiked with 10ppm of Zinc or Copper and treated with nanonets. The treatment procedure was as follows. To prepare the metal spiked seawater solutions, a volume of 175 µL of the 100nM Zn²⁺ solution (28,765 ppm Zn²⁺) or 315 µL of the 100nM Cu²⁺ solution was added to 500 mL seawater, yielding a 10.07 ppm Zn²⁺ and 10.03 ppm Cu²⁺ solution, respectively. A volume of 960 µL of this sample was then treated with 40 µL of each optimized nanonet library solution (Table 9 and Table 10). As a positive control for metal removal, separate 960uL aliquots of the metal spiked seawater samples were treated with 40 µL of 1% NaOH solution to precipitate the Zn or Cu as hydroxides. To ensure metal stability in the seawater, additional 960uL aliquots of the metal spiked seawater solutions

were also mixed with 40 μ L distilled water. The solutions were individually mixed and each centrifuged ($1,000 \times g$) for approximately 5 minutes to sediment precipitated nanonets. Five μ L of the supernatant was analyzed applying the method for Zn^{2+} and Cu^{2+} metal ion removal.

Preparation of Calibration Curves for Method #3

Preparation of calibration standards

For dissolved Zn^{2+} metal ion, the calibration standards were prepared in 0.10, 0.26, 0.52, 1.03, 1.75, 2.52 ppm concentrations, and for dissolved Cu^{2+} , the calibration standards were prepared in 0.10, 0.25, 0.5, 1.0, 1.74, 2.49 ppm concentrations.

Preparation of calibration curves for dissolved Zn^{2+} and Cu^{2+}

Into one set of 1.5 mL conical centrifuge tubes, 25 μ L aliquots of the Zn^{2+} calibration standard solutions were added. Into another set of 1.5 mL conical centrifuge tubes, 25 μ L aliquots of the Cu^{2+} calibration standard solutions were added. To both sets of tubes, 950 μ L of boric acid buffer pH 9 were added and the tubes were mixed followed by the addition of 25 μ L of 1.6 M Zincon solution (75% dye content). The solutions were mixed and allowed to sit for 5 minutes. Aliquots of 200 μ L of the samples were transferred into 96-well flat bottom plates suitable for visible spectrophotometry and the samples were measured at 615 nm. Separate calibration curves were constructed for Zn^{2+} and Cu^{2+} .

Preparation of Calibration Curves for Iron Removal

The following procedure is based upon M. J. Verschoor, L. A. Molot, *Limnol. Oceanogr-Meth.*, (2013), 11, 113-125. A Comparison of Three Colorimetric Methods of Ferrous and Total Reactive Iron Measurement in Freshwaters. (doi:10.4319/lom.2013.11.113). The procedure therein was modified and adapted to quantify total and dissolved iron in water samples. Solutions of 20,000 ppm Fe^{2+} , 10% ascorbic acid and 2.5 mg/mL ferrozine were prepared in water. The 20,000 ppm Fe^{2+} solution was diluted to appropriate

concentrations to prepare the calibration standards. Both Fe^{2+} and Fe^{3+} were quantitated using the calibration curve prepared for Fe^{2+} in the range of 0.1 – 2.0 ppm in deionized water.

Preparation of calibration standards

The calibration standards were prepared in 0.10, 0.25, 0.50, 1.0 and 2.50 ppm concentrations by serial dilutions of the 20,000 ppm Fe^{2+} stock solution with deionized water.

Preparation of calibration curve

Into 1.5 mL conical centrifuge tubes were added 1 mL of the Fe^{2+} calibration standards and 100 μL of 10% ascorbic acid. The solutions were mixed and allowed to sit for 10 minutes. Twenty-five μL of the 2.5 mg/mL ferrozine solution was added and the samples were allowed to sit for 10 min. Aliquots of 200 μL of the samples were transferred to into 96-well flat bottom plates suitable for visible spectrophotometry and analyzed at 562 nm.

The same procedure was applied to measuring total iron and dissolved iron concentrations.

Residual Zn^{2+} and Cu^{2+} concentrations were analyzed by through colorimetric determination by complexation with Zincon. The Zincon method is adapted from a procedure described in (E. Säbel, J. M. Neureuther, S. Siemann. *Anal. Biochem.*, (2010), 397, 2, 218-226, (doi: 10.1016/j.ab.2009.10.037). A Spectrophotometric Method for the Determination of Zinc, Copper, and Cobalt ions in metalloproteins using Zincon Crystal). The procedure disclosed therein was modified and adapted to quantify dissolved Zn^{2+} and Cu^{2+} metal ions in water samples

Theoretical Calculations – Method #4:

The coprecipitation of 7 hydrolyzed polymers and 58 surfactants with varying concentrations resulted in nanonets with various degree of turbidity. The turbidity was measured and the rise in absorption values was modelled as

surfactant was added to polymer. First baseline absorption values a_p and a_s were calculated, which were estimated as linear functions of concentration for a given polymer or surfactant when measured alone. Then baseline-corrected absorption values (A') were calculated by subtracting a_p and a_s from raw absorption values A , i.e. $A' = A - a_s - a_p$.

For each combination of polymer and surfactant, a curve $A'(S, P_0)$ was constructed, where S is a varying concentration of surfactant and P_0 is a fixed concentration of polymer. The peak of this curve was identified and a sigmoid was fit to the pre-peak section of the curve (R , *optim* function). The estimated sigmoid coefficient allowed quantification of the peak height, transition midpoint, and transition rate.

For a given combination of one surfactant and one polymer, nanonet formation was defined as any sigmoid fit with height greater than 0.2. Further, any sigmoids with midpoint past 500ppm were removed, as the titration ended at 1000 ppm and the sigmoid curve required at least 3 data points to be a valid curve. Visual inspection confirmed that the sigmoid fits of A' were a useful method to detect nanonet formation. In total, optimized nanonets occurring between 199 out of 434 possible surfactant and polymer combinations ($62 * 7$) were detected.

Synthesis of Surfactants

Nuclear magnetic resonance (NMR) spectra were performed in the GreenCentre Canada facilities. ^1H and $^{13}\text{C}\{^1\text{H}\}$ NMR spectra were recorded on a VarianTM 400 MHz spectrometer at room temperature. Chemical shifts are reported in part per million (ppm) using tetra-methyl silane (TMS) as standard [^1H (CDCl_3 : 7.26 ppm; $\text{DMSO}-d_6$: 2.50 ppm) and ^{13}C [CDCl_3 : 77.16 ppm; $\text{DMSO}-d_6$: 39.52 ppm]]. Multiplicity of chemical shifts reported as follows: s = singlet, d = doublet, dd = doublet of doublets, t = triplet, m = multiplet, and br = broad. NMR spectra were processed using MestReNova software. Fourier Transform Infrared Spectroscopy (FTIR) spectra were recorded on an Agilent Cary 630 FTIR Spectrometer equipped with a diamond ATR (attenuated total reflectance) and a

ZnSe window with a spectral range of 650-4000- cm^{-1} . The synthetic route of *N*-Dodecanoyl-L-Methionine is referenced in (3) G. Bonacucina *et al. Colloids and Surfaces A: Physicochem. Eng. Aspects*, **2016**, 492, 38-46. Chemical-Physical Properties and Cytotoxicity of *N*-Decanoyl Amino Acid-Based Surfactants: Effect of Polar Heads.

Synthesis of *N*-Dodecanoyl-L-Methionine

To a 100 mL round-bottom flask with a magnetic stir bar, were added methionine (1.50 g, 10 mmol), H_2O (20 mL), and Na_2CO_3 (1.57g, 15 mmol). THF (10 mL) was added and the mixture turned cloudy. *n*-Dodecanoyl chloride (3.0 mL, 13 mmol) was added dropwise and the mixture was stirred at room temperature for 4 h. After the stated time, THF was evaporated under reduced pressure and the reaction mixture was acidified until pH 2.0 (with 2 N HCl). A precipitate was formed and filtered using a Buchner funnel, with #2 filter paper and washed with water (3 x 10 mL), and acetone (about 5-10 mL). The solid was dried in a vacuum oven at 40 °C, overnight. The solid was recrystallized from hexanes. Then, the precipitate was filtered and washed with hexanes (3 x 10 mL), to afford pure product, as a white solid (1.786 g, 54% yield). ^1H NMR (400 MHz, CDCl_3) δ 6.83 (d, J = 7.6 Hz, 1H, NHCO), 4.57 (dd, J = 12.4, 4.8 Hz, 1H), 2.54 (t, J = 7.6 Hz, 2H), 2.23 (t, J = 7.6 Hz, 2H), 2.19 – 2.13 (m, 1H), 2.09 (s, 3H), 2.04 – 1.95 (m, 1H), 1.63 – 1.56 (m, 2H), 1.31 – 1.25 (m, 15H), 0.87 (t, J = 6.8 Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 175.7 (COOH), 174.7 (CONH), 52.3 (HC-NH), 36.6 (O=C-CH_2), 32.0 ($\text{O=C-CH}_2\text{-CH}_2$), 31.2 (CH_2), 30.3 (CH_2), 29.8 (CH_2), 29.7 (CH_2), 29.7 (CH_2), 29.5 (CH_2), 29.5 (CH_2), 29.4 (CH_2), 25.8 (CH_2), 22.8 (CH_2), 15.6 (SCH_2), 14.3 (CH_3); FT-IR (ATR, cm^{-1}): 3303, 2938, 2916, 2849, 1702, 1706, 1646, 1535, 1419, 1226, 685.

Synthesis of *N*-Decanoyl-L-Methionine

To a 100 ml round-bottom flask with a magnetic stir bar, were added methionine (1.50 g, 10 mmol), H_2O (20 mL) and Na_2CO_3 (1.6 g, 15 mmol). Upon addition of THF (10 mL) to the reaction mixture a cloudy solution formed, *n*-

Decanoyl chloride (2.7 mL, 13 mmol) was added dropwise and the mixture was stirred at room temperature for 4 h. The solvent THF was evaporated under reduced pressure and the reaction mixture was acidified until pH 2.0 (with 2 N HCl). At pH 3-4, some precipitate was observed at pH 2 an oil on top of reaction mixture was formed. The reaction mixture was extracted with hexanes (30 mL), and EtOAc (ethyl acetate) (2 x 20 mL). The organic phases were combined and the solvent was removed under reduced pressure. An oil was isolated and stored in the fridge until a solid was obtained. The solid was recrystallized in hexanes, filtered and washed with hexanes (3 x 10 mL), to afford a pure colourless solid (2.516 g, 83% yield). ^1H NMR (400 MHz, CDCl_3) δ 10.18 (br, 1H), 6.55 (d, J = 7.6 Hz, 1H, NHCO), 4.71 (dd, J = 12.8, 5.6 Hz, 1H), 2.55 (t, J = 7.4 Hz, 2H), 2.36 – 2.17 (m, 3H), 2.10 (s, 3H), 2.08 – 1.99 (m, 1H), 1.68 – 1.58 (m, 2H), 1.28 – 1.25 (m, 12H), 0.87 (t, J = 6.8 Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 175.1 (COOH), 174.7 (CONH), 51.9 (HC–NH), 36.6 (O=C–CH₂), 32.0 (O=C–CH₂–CH₂), 31.2 (CH₂), 30.1 (CH₂), 29.6 (CH₂), 29.4 (CH₂), 29.4 (CH₂), 29.3 (CH₂), 29.3 (CH₂), 25.8 (CH₂), 22.8 (CH₂), 15.5 (SCH₂), 14.2 (CH₃); FT-IR (ATR, cm^{-1}): 3314, 2916, 2849, 1706, 1650, 1535, 1441, 1419, 1255, 1229, 920, 685.

Synthesis of N-Octanoyl-L-Methionine

To a 100 ml round-bottom flask with a magnetic stir bar, were added methionine (1.50 g, 10 mmol), H₂O (20 mL), and Na₂CO₃ (1.6 g, 15 mmol). Upon addition of THF (10 mL) to the reaction mixture a cloudy solution formed. n-Octanoyl chloride (2.2 mL, 13 mmol) was added dropwise and the mixture was stirred at ambient temperature for 4 h. The solvent THF was evaporated under reduced pressure on a Schlenk line and the reaction mixture was acidified until pH 2.0 (with 2 N HCl). An oil on top of reaction mixture was formed. The reaction mixture was extracted with EtOAc (3 x 20 mL). The organic phases were combined and the solvent was removed under reduced pressure. An oil was obtained and stored inside the fridge to solidify. The solid was recrystallized in hexanes, filtered, and washed with hexanes (3 x 10 mL), to afford a pure colourless solid (1.194 g, 43% yield). ^1H NMR (400 MHz, CDCl_3) δ 10.54 (br,

1H), 6.58 (d, $J = 7.6$ Hz, 1H, NHCO), 4.70 (dd, $J = 12.6, 5.0$ Hz, 1H), 2.54 (t, $J = 7.4$ Hz, 2H), 2.25 (t, $J = 7.6$ Hz, 2H), 2.21 – 2.17 (m, 1H), 2.09 (s, 3H), 2.08 – 2.00 (m, 1H), 1.63 – 1.60 (m, 2H), 1.28 – 1.25 (m, 8H), 0.86 (t, $J = 6.6$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 175.0 (COOH), 174.7 (CONH), 51.9 (HC-NH), 36.5 (O=C-CH_2), 31.8 ($\text{O=C-CH}_2\text{-CH}_2$), 31.2 (CH_2), 30.1 (CH_2), 29.2 (CH_2), 29.1 (CH_2), 25.7 (CH_2), 22.7 (CH_2), 15.5 (SCH_2), 14.2 (CH_3); FTIR (ATR, cm^{-1}): 3329, 2942, 2919, 2856, 1698, 1612, 1549, 1445, 1248, 1225, 1188, 1128, 957.

Synthesis of N-Dodecanoyl-L-Histidine

To a 100 mL round-bottom flask with a magnetic stir bar, histidine (1.55 g, 10 mmol), H_2O (20 mL), and Na_2CO_3 (1.38 g, 13 mmol) were added followed by the addition of THF (10 mL). The reaction mixture was cooled down to 0 °C. Upon which the mixture turned cloudy. N-dodecanoyl chloride (2.5 mL, 11 mmol) was added dropwise and the mixture was stirred at ambient temperature overnight. Before complete addition of acyl chloride, the solution became cloudy and viscous (gel-like). After 4 h of reaction, the solvent THF was evaporated under reduced pressure and the reaction mixture was acidified until pH 2.0 (with 2 N HCl). The precipitate was filtered and washed with acetone (3 x 5 mL) and CHCl_3 (3 x 5 mL). The ^1H NMR spectrum of the crude product showed residual dodecanoic acid, and the crude was recrystallization in 95% EtOH. The solids were filtered and washed with 95% EtOH. The colourless solid obtained was contaminated with about 12% dodecanoic acid (1.197 g, about 35% yield). ^1H NMR (400 MHz, $\text{DMSO}-d_6$): δ 8.01 (d, $J = 7.6$ Hz, 1H, NHCO), 7.54 (s, 1H, N=CHN), 6.78 (s, 1H, NCH=), 4.41-4.35 (m, 1H, NCHCOOH), 2.94-2.77 (m, 2H, CH_2), 2.05 (t, $J = 7.4$ Hz, 2H, COCH_2), 1.41-1.38 (m, 2H), 1.27-1.20 (m, 16H, CH_2), 0.85 (t, $J = 6.8$ Hz, 3H, CH_3). ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$): δ 173.3 (COOH), 172.1 (CONH), 134.7 (NC=N), 52.1 (HC-NH), 35.2 (O=C-CH_2), 31.4 ($\text{O=C-CH}_2\text{-CH}_2$), 29.1 (CH_2), 29.1 (CH_2), 29.0 (CH_2), 28.9 (CH_2), 28.8 (CH_2), 28.6 (CH_2), 25.2 (CH_2), 22.1 (CH_2), 14.0 (CH_3); FTIR (ATR, cm^{-1}): 3329, 3146, 2945, 2919, 2852, 1639, 1546, 1397, 1184, 983, 834, 685.

Synthesis of N-Dodecanoyl-L-Threonine

To a 100 mL round-bottom flask with a magnetic stir bar, were added threonine (1.20 g, 10 mmol), H₂O (20 mL), and Na₂CO₃ (1.60 g, 15 mmol). Then, THF (10 mL) was added and the mixture turned cloudy. n-dodecanoyl chloride (3.0 mL, 13 mmol) was added dropwise and the mixture was stirred at ambient temperature for 4 h. The solvent THF was evaporated under reduced pressure and the reaction mixture was acidified until pH 2.0 (with 2 N HCl). The colourless precipitate was filtered and washed with deionized water (3 x 10 mL) and dried in a vacuum oven at 40 °C for 3 days. The colourless solid (1.10 g, 36%) was purified by recrystallization from hexanes. ¹H NMR (400 MHz, DMSO-*d*₆): δ 7.75 (d, *J* = 8.8 Hz, 1H, NHCO), 4.20 (dd, *J* = 8.8, 3.2 Hz, 1H, CH–NH), 4.13–4.07 (m, 1H, CH–OH), 2.26–2.11 (m, 2H, O=C–CH₂), 1.49–1.46 (m, 2H, CH₂), 1.23 (m, 15H, CH₂), 1.04 (d, *J* = 6.4 Hz, 3H, CHOH–CH₃), 0.85 (t, *J* = 6.8 Hz, 3H, CH₃); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 172.7 (COOH), 172.4 (CONH), 66.5 (HC–OH), 57.4 (HC–NH), 35.1 (O=C–CH₂), 32.0 (O=C–CH₂–CH₂), 29.1 (CH₂), 29.0 (CH₂), 28.9 (CH₂), 28.8 (CH₂), 28.7 (CH₂), 25.4 (CH₂), 22.5 (CH₂), 20.5 (HOHC–CH₃), 14.1 (CH₃); FTIR (ATR, cm^{–1}): 3426, 3362, 3314, 2942, 2919, 2852, 1709, 1612, 1605, 1538, 1463, 1385, 1382, 1207, 1151, 1088, 871, 674.

Synthesis of N-Decanoyl-L-Threonine

To a 100 mL round-bottom flask with a magnetic stir bar, were added threonine (1.20 g, 10 mmol), H₂O (20 mL) and NaOH (0.6 g, 15 mmol). Then, THF (10 mL) was added, followed by the dropwise addition of n-decanoyl chloride (2.7 mL, 13 mmol) and the mixture was stirred at ambient temperature overnight. A suspension was obtained and the solvent THF was evaporated under reduced pressure and the reaction mixture was acidified until pH 2 (with 2 N HCl). No precipitate was observed. The reaction mixture was extracted with EtOAc (3 x 20 mL). The organic phases were combined and the solvent removed under reduced pressure. An oil was obtained and purified using a short plug of silica, eluted with Hex/EtOAc 6:4 to remove decanoic acid, and then with EtOAc, to remove the product, a colourless solid (1.326 g, 49% yield). ¹H NMR (400

MHz, CDCl₃): δ 7.00 (d, J = 8.8 Hz, 1H, NHCO), 4.54 (d, J = 6.4 Hz, 1H, CH-NH), 4.43-4.41 (m, 1H, CH-OH), 2.29 (t, J = 7.6 Hz, 2H, O=C-CH₂), 1.64-1.60 (m, 2H, CH₂), 1.28-1.25 (m, 15H, CH₂), 1.20 (d, J = 6.4 Hz, 3H, CHOH-CH₃), 0.87 (t, J = 6.6 Hz, 3H, CH₃); ¹³C NMR (100 MHz, CDCl₃): δ 175.5 (COOH), 174.3 (CONH), 67.7 (HC-OH), 57.8 (HC-NH), 36.5 (O=C-CH₂), 32.0 (O=C-CH₂-CH₂), 32.0 (CH₂), 29.7 (CH₂), 29.7 (CH₂), 29.6 (CH₂), 29.5 (CH₂), 29.4 (CH₂), 29.4 (CH₂), 25.9 (CH₂), 22.8 (CH₂), 19.6 (HOHC-CH₃), 14.2 (CH₃); FTIR (ATR, cm⁻¹): 3426, 3363, 3314, 2938, 2919, 2852, 1709, 1616, 1605, 1538, 1419, 1382, 1277, 1270, 1199, 1151, 1087, 875, 675.

Synthesis of N-Octanoyl-L-Threonine

To a 100 ml round-bottom flask with a magnetic stir bar were added threonine (1.20 g, 10 mmol), H₂O (20 mL), and Na₂CO₃ (1.60 g, 15 mmol). Then, THF (10 mL) was added followed by the dropwise addition of n-octanoyl chloride (2.2 mL, 13 mmol). The reaction mixture was stirred at ambient temperature for 4 h. After the stated time, THF was evaporated under reduced pressure and the reaction mixture was acidified until pH 2.0 (with 2 N HCl). No precipitate was observed. The reaction mixture was extracted with 3 x EtOAc (20 mL each). The organic phases were combined and the solvent removed under reduced pressure to afford an oil. The oil was purified using a short plug of silica, eluted with Hex/EtOAc 6:4 to remove decanoic acid, and then with EtOAc to remove the product. (1.039 g, 42% yield). ¹H NMR (400 MHz, CDCl₃): δ 7.01 (d, J = 8.4 Hz, 1H, NHCO), 4.54 (dd, J = 8.4, 2.4 Hz, 1H, CH-NH), 4.42-4.40 (m, 1H, CH-OH), 2.29 (t, J = 7.6 Hz, 2H, O=C-CH₂), 1.63-1.60 (m, 2H, CH₂), 1.30-1.25 (m, 10H, CH₂), 1.19 (d, J = 6.0 Hz, 3H, CHOH-CH₃), 0.86 (t, J = 6.8 Hz, 3H, CH₃); ¹³C NMR (100 MHz, CDCl₃): δ 176.3 (COOH), 173.7 (CONH), 67.7 (HC-OH), 57.5 (HC-NH), 36.4 (O=C-CH₂), 32.5 (O=C-CH₂-CH₂), 29.1 (CH₂), 25.8 (CH₂), 22.7 (CH₂), 19.4 (HOHC-CH₃), 14.0 (CH₃); FTIR (ATR, cm⁻¹): 3303, 2942, 2927, 2856, 1724, 1631, 1534, 1419, 1415, 1382, 1210, 1117, 1084, 1080, 857, 674.

Example 15

Nanonet Formation

To rapidly screen for the formation of nanonets using any particular surfactant and polymer combination, a titration based approach was utilized. Increasing amounts of surfactant were added to a fixed amount of polymer. Subsequent destabilization of the polymer by addition of concentrated hydrochloric acid lead to precipitation of both surfactant and polymer, indicating nanonet formation. The amount of nanonet formation was measured by an increase in absorbance at 540nm relative to polymer only and surfactant only controls. This method can be utilized as a rapid, automated approach to screening polymer-surfactant interactions for the purposes of observing the onset of nanonet formation in any given combination of polymer(s) and surfactants(s) (see Reagents and Materials Examples 15-21, method #1).

Formation of Nanonets With The Triton™ Series of Surfactants and Validation of a Screening Method.

To test nanonet formation with multiple different surfactants and polymers, a screening method was designed utilizing co-precipitation of surfactant and polymer as a measure of nanonet formation. To test the screening method, nanonet formation was first measured between model surfactants, selected from the Triton™ series, with styrene maleic acid block co-polymers of varying length and block composition (Figure 15). The Triton™ series of surfactants correspond to amphipathic molecules with a hydrophobic alkyl chain of 14 carbons, and a hydrophilic head group composed of repeating oxyethylene groups (see Formula (i) above). By increasing the number of repeating oxyethylene groups, the HLB value of the surfactant increases and surfactant aggregate size of the resulting surfactant decreases. This trend allows for creation of surfactants with similar chemistry but radically different physical wetting, foaming, and aggregation properties (Stubicar and Petres, *Micelle Formation by Tritons in Aqueous Solutions*, (1981), *Croatia Chemica Acta*, 54 (3) 255-266)

Utilizing the SMA polymers and the Triton™ series of surfactants, the screening method for the onset of nanonet formation may be validated. What follows below is a comparison of nanonet formation using relatively smaller, and relatively larger, SMA polymer particle sizes in combination with relatively smaller and relatively larger surfactant aggregate sizes within the Triton™ series of surfactants. Thus, many possible combinations of relative size could be observed. The Triton™ surfactants employed were as follows (along with measured surfactant aggregate sizes): (Triton X-45™: 156 nm), and a small micelle forming surfactant (Triton X-100™: 10 nm). Polymer particle sizes are detailed below along with results.

A clear increase in absorbance indicative of nanonet formation was observed with Triton X-100™ and SMA-2000™ ($D_{\text{calculated}}$: 2.6 nm, D_{observed} : 5.2nm) and SMA-3000™ ($D_{\text{calculated}}$: 2.8 nm, D_{observed} : 3.7 nm) (Figure 15). Conversely, the relatively larger SMA-725™ ($D_{\text{calculated}}$: 6.8 nm, D_{observed} : 10-15nm) showed little increase in absorbance with Triton X-100™. All three polymers showed strong increase in absorbance with the larger surfactant aggregate formed by Triton X-45™. To confirm that this increase in absorbance corresponded to nanonet formation (and not mere precipitation of a given component), The precipitate was removed from solution by centrifugation. Residual surfactant left in the solution was measured by UV absorbance of the Triton™ surfactants for SMA 2000 and SMA 725 (Table 21). The results demonstrated that the increase in formed precipitate was proportional to a corresponding decrease in surfactant concentration in the supernatant. This result validates co-precipitation as a valid screening method for measuring nanonet formation(see Reagents and Materials Examples 15-21, method #2). The results obtained from the screening method, support the proposition that nanonet formation is influenced by a size mismatch between a minimal polymer particle size and surfactant aggregate size. For example, this proposition may be stated as follows: ($D_{\text{polymer}} < D_{\text{surfactant aggregate}} = \text{nanonet}$), and may be visualized

through the relatively simple rapid, automated screening method detailed above. Illustrated results are set out in Figure 15.

Example 16

Conversion of the Nanonet Screening Method To A High Throughput Method and Measurement of Nanonet Formation.

To better measure nanonet formation between various surfactants and numerous polymers, a relatively more automated and quantitative scoring method for detecting nanonet formation was developed. Consider first the simple screening method described above and validated. In that validation, nanonet formation was assumed to occur between a single block co-polymer and a single surfactant aggregate. What follows is the description of a computer-assisted model developed to predict, standardize, and score the formation of nanonets.

A binding curve of an X-Y plot of absorbance at 540 nm versus surfactant concentration was fit with a sigmoidal function after correction for surfactant only artifacts (Figure 16). From the sigmoid fit, a nanonet formation factor (Nf) was determined. Nf is taken from the sigmoid fit as the half maximal value of the sigmoid. Nf may also be referred to as the midway point, half-maximal concentration, or critical aggregation concentration. Sigmoid fitting also provided values for the absolute height of the sigmoid, and the slope of the sigmoid curve.

Sigmoid curves with a steeper slope may result from rapid association of all available surfactant with the polymer. This has been described as a hallmark of relatively high cooperativity of polymer surfactant binding. (Goddard E.D. Polymer/Surfactant Interaction-Its relevance to Detergent Systems. JAOCS. Vol 71, 1994). Similarly, sigmoid curves with a relatively higher maximum absorbance may be indicative of association of more of the available surfactant with the polymer, and may also indicate more efficient nanonet formation. From these two sigmoid curve parameters (slope and maximum absorbance), the area under the sigmoid curve was used as a score for nanonet formation (Figure 17). The nanonet formation factor (Nf) is taken to be the point at which half of the available polymer was became associated with surfactant.

To summarize, in this automated method, nanonet formation was construed to begin based upon a sigmoid fit of parameters taken from absorbance versus surfactant concentration plots. A nanonet score was calculated using the area under the sigmoid curve (AUC). If no sigmoid was detected, Nanonet formation was determined to have not occurred. Each polymer and surfactant combination were scored independently. The resulting AUC values were scaled between 0 and 1. The resulting score, so scaled, may be indicative of nanonet formation. The scaled score captures two aspects. First, it captures the height of the absorption profile, which may indicate relatively greater quantities of nanonets formed. Second, it captures the surfactant concentration range (for a fixed polymer concentration) over which nanonets are formed. Illustrated results are set out in Figure 17.

Example 17

Nanonet Formation Occurs With Both Anionic and Non-Ionic Surfactants.

In this section, a study of the influence of the chemical structure of the surfactant headgroup on nanonet formation was undertaken. Nanonet formation with surfactants containing various non-ionic and ionic headgroups was examined. Headgroups were classified by the functional chemical group forming the primary interactions with water in solution, the number of repeating functional groups, and linkage to the hydrophobic alkyl chain. Except for combinations shown below (Table 13 and 14), the formation of nanonets was detected with all other surfactants tested (Tables 12 A-D).

Table 12A. Positively Scoring Nanonets

Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
1,2-Hexanediol	DIBMA	0.1	901.088435	55.485365	0.08909591
Amphosol 2CSF™	DIBMA	0.77027187	595.08416	92.7298755	0.01197528
Apo 10	DIBMA	0.84774592	438.416181	95.641022	0.02734266
Apo 12	DIBMA	1.02744699	84.1216144	20.6225583	0.05375417
Stepan Bio Soft D-40™	DIBMA	0.50324237	154.38828	15.7339089	0.23726898
Cedepal TD-403 MFLD™	DIBMA	0.16418752	155.336851	21.5040037	0.1078598
Coco Glucoside	DIBMA	0.54794297	117.587414	11.5384967	0.07657559
Dodecane-1-sulfonic acid	DIBMA	0.1	113.103271	22.0604016	0.06354805
PFDMP-O310F	DIBMA	0.62275273	653.987409	69.6324963	0.02728391
Genapol X080	DIBMA	0.71496667	20.9693868	1	0.52045649
HEGMDDE H370	DIBMA	0.77233692	22.9310378	6.94012383	0.31281476
HEGMDE H360	DIBMA	0.73495559	15.6419746	1	0.22670991
HEGMOE H350	DIBMA	0.21723333	31.5858482	1.67734031	0.14408076
HXEGMDDE H375	DIBMA	0.61635555	48.9368401	1	0.16375448
Lakeland PAE 136™	DIBMA	0.61441111	45.4594876	2.03653556	0.31629426
Stepan Lathanol LAL™	DIBMA	0.26535556	50.1592844	1	0.17633476
MNA C12	DIBMA	0.35946774	96.4098691	21.0225132	0.21215451
MNA C13	DIBMA	0.44239109	127.190065	19.3785602	0.21842851
MNA C14	DIBMA	0.37788666	107.360025	27.9020346	0.22240426
n-Decanoyl-L-Methionine	DIBMA	0.72673333	22.5959793	1	0.42891738
n-Decanoyl-L-Threonine	DIBMA	0.72474944	326.332201	21.5716367	0.15178313
n-Dodecanoyl-L-Histidine	DIBMA	0.26962716	112.97279	55.8505974	0.14662091
n-Dodecanoyl-L-Methionine	DIBMA	0.41636667	19.6303579	1.00005512	0.31618239

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
n-Dodecanoyl-L-Threonine	DIBMA	0.68847777	122.622926	1	0.13991125
n-Octanoyl-L-Methionine	DIBMA	0.30027282	480.892315	122.694584	0.10060373
Sodium dodecylbenzene sulfonate	DIBMA	0.17219996	53.1225689	4.99559612	0.14861309
nDDTP D342	DIBMA	1.08387992	143.724993	28.3862408	0.32460789
nDDTGP D323	DIBMA	1.00681667	162.687526	1	0.81488173
nDDTP D335	DIBMA	1.02878536	658.069895	111.181966	0.04566893
Ninate 411™	DIBMA	0.30978354	155.42648	25.6747016	0.16191695
nNDTP N335	DIBMA	0.86906633	565.917927	17.1427281	0.02297988
OEGMDE O330	DIBMA	0.6817	12.4022252	1	0.40518747
Poly-10- laurate	DIBMA	0.42863333	56.1286226	3.54827919	0.25251619
Poly-10- oleate	DIBMA	0.8523417	42.6516533	3.03531455	0.09171199
Poly-10-stearate	DIBMA	0.51043339	503.701113	4.14415278	0.03461897
Span 20™	DIBMA	0.24467942	43.3092107	6.61452179	0.08822107
Stepan Mild L3™	DIBMA	0.35162074	130.031097	66.1989563	0.10881984
Stepan SLL FB™	DIBMA	0.14626668	559.237685	19.3401413	0.02347724
Stepan sulfonic acid™	DIBMA	0.19633275	47.7084451	5.07352302	0.19511718
TC Lab 35	DIBMA	0.71271471	334.20971	69.445712	0.04069248
Triton X-100™	DIBMA	0.84218245	88.2041539	37.3457156	0.3181006
Triton X-305™	DIBMA	0.1	91.8020987	72.9726761	0.06771337
Triton X-45™	DIBMA	0.97430833	44.3902689	2.65271092	0.21510792
Tween 20™	DIBMA	0.2062	59.4954843	2.56038525	0.16107583
Tween 40™	DIBMA	0.1	8.45029038	1	0.10140582
Tween 60™	DIBMA	0.12510002	35.0104779	1	0.11411969
tween 65™	DIBMA	0.39536667	27.3521657	1	0.20251538

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
Tween 80™	DIBMA	0.1	42.5149373	4.84881196	0.10169357
Tween 85™	DIBMA	1.1473	50.0724917	1	0.7354596
1-Octane sulfonate	SMA 2000™	0.1	534.878493	583.851199	0.02086525
Amin GCK 30H	SMA 2000™	0.12479326	36.4305728	13.0012012	0.09493266
Amphosol 2CSF™	SMA 2000™	0.89738859	526.085102	85.0942317	0.12821088
Apo 10	SMA 2000™	0.66099518	235.351463	49.4562223	0.04355634
Apo 12	SMA 2000™	0.83502697	43.9420198	3.6786668	0.11565243
Stepan Bio Soft D-40™	SMA 2000™	0.79162791	137.979405	85.4989689	0.4041504
Stepan Bio Terge AS-40™	SMA 2000™	0.92848624	253.779429	165.263877	0.46161255
Coco Glucoside	SMA 2000™	1.26892037	332.677184	125.515767	0.38850904
Dodecane-1-sulfonic acid	SMA 2000™	0.56516733	160.502073	24.1015342	0.28669766
PFDMP-O310F	SMA 2000™	1.01532665	680.265245	38.2248183	0.01984661
Genapol X080	SMA 2000™	1.0753076	11.7682243	3.36750959	0.77029655
HEGMDDE H370	SMA 2000™	1.09769487	21.7341727	3.09134026	0.50035298
HEGMDE H360	SMA 2000™	1.12000001	14.0557595	1	0.10236606
HEGMOEH350	SMA 2000™	0.91326667	29.6281823	1.50144269	0.5367388
HXEGMDDE H375	SMA 2000™	1.28238258	11.1512028	1.64726731	0.48667566
Lakeland PAE 136™	SMA 2000™	0.66803889	5.39012428	1	0.37215477
Stepan Lathanol LAL™	SMA 2000™	0.74126316	91.8029172	14.1917398	0.48785472
MNA C13	SMA 2000™	0.6319667	128.761567	3.711762	0.37348738
MNA C14	SMA 2000™	0.99491667	392.376055	1	0.46130594
n-Decanoyl-L-Methionine	SMA 2000™	0.73880069	49.794164	4.61655421	0.43712689
n-Decanoyl-L-Threonine	SMA 2000™	0.73130093	456.577364	38.4224003	0.03601437
n-Dodecanoyl-L-Histidine	SMA 2000™	1.61427321	374.522858	139.368208	0.50129881

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
n-Dodecanoyl-L-Methionine	SMA 2000™	0.4084	57.0809993	2.23986642	0.1950195
n-Dodecanoyl-L-Threonine	SMA 2000™	0.9988012	75.5389069	22.2511234	0.30337225
n-Octanoyl-L-Methionine	SMA 2000™	0.8677396	595.722748	128.683455	0.1129965
Sodium dodecylbenzene sulfonate	SMA 2000™	0.82733333	58.4154015	3.15064529	0.52388549
nDDTP D342	SMA 2000™	1.1908412	133.315281	28.3191444	0.66418528
nDDTGP D323	SMA 2000™	1.10939784	41.8240721	9.60983666	0.97587682
nDDTP D335	SMA 2000™	1.20761612	378.515512	44.2671138	0.01731967
Ninate 411™	SMA 2000™	0.56074691	124.95851	7.4506132	0.28406656
nNDTP N335	SMA 2000™	1.22379909	556.256227	16.7117816	0.01241129
OEGMDE O330	SMA 2000™	1.09319861	14.89518	1.89476483	0.65447037
Poly-10-laurate	SMA 2000™	1.20588317	86.2204713	42.8109052	0.59840684
Poly-10-oleate	SMA 2000™	0.52048013	333.405039	40.5367025	0.43078348
Poly-10-stearate	SMA 2000™	1.07071039	682.310738	67.9169494	0.02250116
Span 20™	SMA 2000™	0.88136828	210.360538	40.0411291	0.23162938
Stepan Mild L3™	SMA 2000™	0.75077124	258.170262	149.886645	0.14475193
Stepan SLL FB™	SMA 2000™	0.97317757	77.8264966	32.5773193	0.6772525
Stepan sulfonic acid™	SMA 2000™	0.58338839	30.4316496	13.1990135	0.4432083
TC Lab 35	SMA 2000™	0.97102218	165.127063	35.7111399	0.41573918
Triton X-100™	SMA 2000™	1.090175	30.9867367	1	0.72468072
Triton X-305™	SMA 2000™	0.42093514	52.7835175	15.5976649	0.30439504
Triton X-45™	SMA 2000™	0.95685104	309.862067	45.4754577	0.36300713
Tween 20™	SMA 2000™	0.91024998	34.984391	10.9797071	0.57074161
Tween 40™	SMA 2000™	0.85543611	3.17731708	1	0.68636157
Tween 60™	SMA 2000™	0.9335923	39.2492665	10.5156914	0.73452283

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
Tween 65™	SMA 2000™	1.00891099	88.268573	44.9631639	0.50517267
Tween 80™	SMA 2000™	0.6415	5.33232732	1.15717371	0.55961401
Tween 85™	SMA 2000™	1.16957282	27.4118067	6.90087914	0.77074868
VDISTILL DV53™	SMA 2000™	0.13904954	71.4810149	11.2021876	0.10627094
VDISTILL DV63™	SMA 2000™	0.15760566	123.096414	4.36983785	0.08516374
Sodium-1 octane sulfonate	SMA 2021™	0.1	935.389666	161.242254	0.00725711
Amin GCK 30H	SMA 2021™	0.16113336	127.913092	1	0.16408753
Amphosol 2CSF™	SMA 2021™	1.13313179	699.363304	130.974204	0.04151624
Anthraquinone-2-sulfonic acid	SMA 2021™	0.1	144.475114	4.22812193	0.08545977
Apo 10	SMA 2021™	0.40881668	424.402176	82.9898363	0.02397257
Apo 12	SMA 2021™	0.43479445	45.8318637	9.10508678	0.04698531
Cedepal TD-403 MFLD™	SMA 2021™	0.37948356	202.197224	355.854349	0.19519247
Coco Glucoside	SMA 2021™	0.98464268	195.488268	98.3074408	0.54331317
Dodecane-1-sulfonic acid	SMA 2021™	0.31287633	231.295941	87.00054	0.18335223
PFDMP-O310F	SMA 2021™	0.4981352	512.350393	14.1993402	0.06167963
Genapol X080	SMA 2021™	2	575.161187	228.267432	0.61372737
HEGMDDE H370	SMA 2021™	1.3322001	28.222288	1	0.72265722
HEGMDDE H360	SMA 2021™	1.23872227	15.2556935	1	0.34880255
HEGMOE H350	SMA 2021™	1.26373384	48.4702544	1	0.83467596
HXEGMDDE H375	SMA 2021™	1.28879888	19.5782657	2.56428128	0.23248461
Lakeland PAE 136™	SMA 2021™	1.4386872	85.0790029	63.634503	0.73781132
Stepan Lathanol LAL™	SMA 2021™	0.57771661	141.186295	51.5380326	0.30928375
MNA C13	SMA 2021™	0.2409385	134.849306	46.1029803	0.17162141
MNA C14	SMA 2021™	0.78835514	283.952241	73.5407791	0.37225011

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
n-Decanoyl-L-Methionine	SMA 2021™	0.891	88.0421719	1	0.49088847
n-Decanoyl-L-Threonine	SMA 2021™	0.4390166	392.356052	40.0057236	0.05193664
n-Dodecanoyl-L-Methionine	SMA 2021™	0.41149996	123.17186	1.25778314	0.25692248
n-Dodecanoyl-L-Threonine	SMA 2021™	0.86790988	50.8818456	4.26347048	0.29433431
n-Octanoyl-L-Methionine	SMA 2021™	0.60579899	516.821604	12.9652078	0.17133589
Sodium dodecylbenzene sulfonate	SMA 2021™	1.09185243	161.669071	19.5260699	0.26996932
nDDTP D342	SMA 2021™	1.20839103	245.77649	55.2888738	0.6628966
nDDTGP D323	SMA 2021™	0.67210572	271.14231	245.389902	0.42141363
nDDTP D335	SMA 2021™	1.2873531	603.00528	115.949197	0.0447591
Ninate 411™	SMA 2021™	0.9306941	171.064771	20.4236571	0.29365403
nNDTP N335	SMA 2021™	0.86373574	547.727127	15.5023812	0.03540623
OEGMDE O330	SMA 2021™	1.32343889	21.8346265	2.90574343	0.62121758
Poly-10-laurate	SMA 2021™	1.28228728	155.971849	34.137995	0.52399
Poly-10-oleate	SMA 2021™	1.18861032	83.4270066	13.4234048	0.8530148
Poly-10-palmitate	SMA 2021™	0.35150268	543.456395	119.620995	0.10326259
Poly-10-stearate	SMA 2021™	0.3782	568.843671	1	0.01032824
Span 20™	SMA 2021™	0.35694164	0	560.882957	0.27224066
Stepan Mild L3™	SMA 2021™	0.69871255	244.417729	105.835039	0.53848898
Stepan SLL FB™	SMA 2021™	0.92909918	164.645031	36.1794065	0.66869796
Stepan sulfonic acid™	SMA 2021™	0.55220886	55.2137116	37.4305648	0.34163289
TC Lab 35	SMA 2021™	0.96554937	389.055194	51.5484316	0.16979021
Triton X-100™	SMA 2021™	0.42769999	33.9152557	5.81523546	0.32101287
Triton X-305™	SMA 2021™	0.14214001	59.2815788	24.7028952	0.10990348
Triton X-45™	SMA 2021™	0.58975329	76.5263381	39.5915588	0.33915197

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
Tween 40™	SMA 2021™	0.36146692	118.71608	85.3871419	0.24266693
Tween 60™	SMA 2021™	0.67845333	127.223016	1	0.44119457
Tween 80™	SMA 2021™	1.21108659	128.01887	38.4517763	0.78075637
Tween 85™	SMA 2021™	0.99793553	44.3368596	37.5045817	0.60323297
Amphosol 2CSF™	SMA 3000™	0.2431068	387.142003	84.7206518	0.05874185
Anthraquinone-2-sulfonic acid	SMA 3000™	0.1	267.220564	203.433114	0.01958907
Apo 10	SMA 3000™	0.59850259	345.652245	63.7069632	0.0446669
Apo 12	SMA 3000™	0.64	22.0604935	1	0.45617241
Coco Glucoside	SMA 3000™	1.5392872	392.820112	89.9918433	0.5250622
PFDMP-O310F	SMA 3000™	0.35033333	978.524449	1	0.05425814
Genapol X080	SMA 3000™	1.42288709	38.7011311	33.5395704	1.04843213
HEGMDDE H370	SMA 3000™	0.95577965	23.3898301	4.16284629	0.38737013
HEGMDE H360	SMA 3000™	0.99145558	14.9430288	1	0.27724382
HEGMOE H350	SMA 3000™	0.58346666	30.226552	1.32455222	0.38455735
HXEGMDDE H375	SMA 3000™	1.03194568	6.42321656	1	0.40506607
Lakeland PAE 136™	SMA 3000™	0.65675001	13.8265829	2.05437227	0.53884736
Stepan Lathanol LAL™	SMA 3000™	0.15055264	72.3648955	32.957779	0.11609036
MNA C12	SMA 3000™	0.44480852	213.732813	67.2106088	0.14180333
MNA C13	SMA 3000™	0.37384604	137.945378	24.257433	0.15204811
MNA C14	SMA 3000™	0.34348388	148.758063	27.4801783	0.19699695
n-Decanoyl-L-Methionine	SMA 3000™	0.40006551	45.0739894	3.13635836	0.3975135
n-Decanoyl-L-Threonine	SMA 3000™	0.66326463	577.018149	101.155496	0.03172253
n-Dodecanoyl-L-Histidine	SMA 3000™	0.17466666	50.2447083	1.55294276	0.09766373
n-Dodecanoyl-L-Methionine	SMA 3000™	0.52365556	90.1723376	1	0.2635066

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
n-Dodecanoyl-L-Threonine	SMA 3000™	0.91735595	145.757221	20.6970266	0.27636447
n-Octanoyl-L-Methionine	SMA 3000™	0.70518455	603.060198	78.2084217	0.28379335
Sodium dodecylbenzene sulfonate	SMA 3000™	0.1	98.6810362	46.037207	0.13681818
nDDTP D342	SMA 3000™	0.94715999	291.719517	77.3415987	0.31719493
nDDTGP D323	SMA 3000™	0.90149358	42.8290789	7.39689134	0.73194673
nDDTP D335	SMA 3000™	1.03777981	540.1365	67.2903046	0.03244535
Ninate 411™	SMA 3000™	0.32081235	107.423919	52.4429783	0.16493285
nNDTP N335	SMA 3000™	1.01744385	674.951314	56.9789227	0.01531551
nODTTP O320	SMA 3000™	0.1	471.641835	112.970763	0.0610347
OEGMDE O330	SMA 3000™	1.01130002	13.7605991	1	0.64921569
Poly-10-laurate	SMA 3000™	0.73904923	43.3998996	11.3077173	0.49205051
Poly-10-oleate	SMA 3000™	0.37827778	461.91294	1	0.34235745
Poly-10-palmitate	SMA 3000™	0.56899066	42.8717573	11.3588766	0.49671037
Poly-10-stearate	SMA 3000™	0.87370102	573.481425	18.2860756	0.00817572
Span 20™	SMA 3000™	0.88478417	223.911109	59.231853	0.16455893
Stepan Mild L3™	SMA 3000™	0.64376341	248.456385	60.6747749	0.08284202
Stepan SLL FB™	SMA 3000™	0.78988212	71.5162006	26.1873518	0.68163018
Stepan sulfonic acid™	SMA 3000™	0.24164413	28.317891	13.237126	0.22898645
TC Lab 35	SMA 3000™	0.22991524	346.941575	125.389064	0.09589022
Triton X-100™	SMA 3000™	0.6009527	3.28892177	1	0.48131015
Triton X-305™	SMA 3000™	0.14412718	70.8706163	31.6534128	0.11452151
Triton X-45™	SMA 3000™	0.77245801	48.4308877	9.94655414	0.42715311
Tween 65™	SMA 3000™	2	132.474221	243.533814	0.73905917
Tween 85™	SMA 3000™	2	210.604204	168.795922	0.76177281

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
Amin GCK 30H	SMA-IBE	0.13840166	55.1971085	12.9955696	0.08879885
Amphosol 2CSF™	SMA-IBE	0.1	595.782092	359.850059	0.02127168
Apo 10	SMA-IBE	0.27167398	522.590715	179.741526	0.03888101
Apo 12	SMA-IBE	0.55703334	49.9902832	1.00232534	0.2472827
Stepan Bio Soft D-40™	SMA-IBE	0.1	331.505939	57.327707	0.05184201
Coco Glucoside	SMA-IBE	0.21820058	43.4533724	5.83931635	0.09625545
Fluoro octyl maltoside O310F	SMA-IBE	0.1	479.469366	123.671052	0.04984429
HEGMDDE H370	SMA-IBE	0.51316586	109.786116	17.767388	0.07687877
HEGMDE H360	SMA-IBE	0.70257438	271.535997	21.286082	0.14827892
HXEGMDDE H375	SMA-IBE	0.83539321	17.972628	4.13175952	0.26012348
Lakeland PAE 136™	SMA-IBE	0.14100002	54.8904151	2.98756606	0.15685401
MNA C14	SMA-IBE	0.43062183	603.240819	256.721295	0.09519527
n-Decanoyl-L-Methionine	SMA-IBE	0.91993334	50.0123255	3.27012462	0.52108808
n-Decanoyl-L-Threonine	SMA-IBE	0.51336231	658.954775	70.475536	0.01496219
n-Dodecanoyl-L-Histidine	SMA-IBE	0.11277886	114.538291	57.6895038	0.08483761
n-Dodecanoyl-L-Methionine	SMA-IBE	0.47576667	92.4348088	1	0.27032538
n-Dodecanoyl-L-Threonine	SMA-IBE	0.14354495	318.504378	155.735051	0.0714816
n-Octanoyl-L-Methionine	SMA-IBE	0.62686283	716.038034	111.041836	0.07385598
nDDDT D342	SMA-IBE	0.22859992	54.0350613	2.33743662	0.10447126
nDDTGP D323	SMA-IBE	0.14623362	499.220019	10.1196651	0.22788446
nDDTP D335	SMA-IBE	0.27080924	212.359061	65.2471625	0.09548108
OEGMDE O330	SMA-IBE	0.11736715	54.4395671	6.07004333	0.07830438
Poly-10-laurate	SMA-IBE	0.11284842	75.6808608	17.7297431	0.03695515
Poly-10-oleate	SMA-IBE	0.39399999	49.7886456	1.03559627	0.151117336

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
Span 20™	SMA-IBE	0.11276668	504.450638	4.56821982	0.09276794
Stepan Mild L3™	SMA-IBE	0.42979985	123.215334	2.75949469	0.23430997
Stepan SLL FB™	SMA-IBE	0.50575003	49.8392414	1.00617269	0.10171128
Stepan sulfonic acid™	SMA-IBE	0.12123375	56.4878144	5.84092656	0.12913625
Triton X-305™	SMA-IBE	0.1	369.852806	310.374151	0.05229663
Triton X-45™	SMA-IBE	0.3194	31.9394658	1	0.34639209
Tween 60™	SMA-IBE	0.12768971	790.283457	281.753355	0.04439502
Tween 65™	SMA-IBE	0.17028942	244.053429	7.32272882	0.06460263
Tween 85™	SMA-IBE	0.2088	25.346497	1	0.21842755
1,2-Hexanediol	SMA 725™	0.16665356	78.0526029	29.96572	0.0786097
Amin GCK 30H	SMA 725™	0.21016623	380.083452	165.338014	0.19224575
Amphosol 2CSF™	SMA 725™	0.1	398.10186	164.474318	0.04635902
Apo 10	SMA 725™	0.34625609	595.290306	129.398525	0.0322409
Apo 12	SMA 725™	0.16243333	5.17360531	1.36617645	0.1467061
Stepan Bio Soft D-40™	SMA 725™	1.18217395	628.407481	244.949233	0.40729597
Stepan Bio Terge AS-40™	SMA 725™	0.81420252	373.201656	182.869165	0.39245615
Dodecane-1-sulfonic acid	SMA 725™	0.37391949	61.5385496	11.5744684	0.49054484
PFDMP-O310F	SMA 725™	0.12143325	497.138141	16.0950889	0.03517421
Genapol X080	SMA 725™	2	437.146479	188.834499	0.81568587
HEGMDDE H370	SMA 725™	0.40963279	21.1965285	2.60828794	0.3036369
HEGMDDE H360	SMA 725™	0.94983345	16.0164725	1	0.49167483
HXEGMDDE H375	SMA 725™	0.31032122	6.42321847	1	0.2859397
Lakeland PAE 136™	SMA 725™	0.40092	6.7971917	2.13221812	0.45672292
Stepan Lathanol LAL™	SMA 725™	1.19795721	457.059606	133.528247	0.50554515

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
MNA C12	SMA 725™	0.1142333	55.1225816	2.58801463	0.09770116
MNA C14	SMA 725™	0.23660233	150.054301	22.1077259	0.16548523
n-Decanoyl-L-Methionine	SMA 725™	0.58480067	47.3254363	2.68989215	0.3245131
n-Decanoyl-L-Threonine	SMA 725™	0.58337096	689.059826	63.3478749	0.01010457
n-Dodecanoyl-L-Histidine	SMA 725™	0.21162515	265.115704	107.553749	0.08328081
n-Dodecanoyl-L-Methionine	SMA 725™	0.88116667	91.8738612	1	0.52147673
n-Dodecanoyl-L-Threonine	SMA 725™	0.70376668	127.100453	1.95061246	0.18621891
n-Octanoyl-L-Methionine	SMA 725™	0.67963151	629.742403	171.134141	0.1470039
nDDTP D342	SMA 725™	0.56658422	468.144931	104.513987	0.16931911
nDDTGP D323	SMA 725™	0.52058349	295.081064	126.495496	0.50181278
nDDTP D335	SMA 725™	0.1	398.778329	175.312356	0.05487862
Ninate 411™	SMA 725™	0.10989205	80.3623182	34.3296293	0.08815548
nNDTP N335	SMA 725™	0.24651841	699.874309	111.827067	0.02225363
OEGMDE O330	SMA 725™	0.19725853	6.4233049	1	0.17818649
Poly-10-laurate	SMA 725™	1.22644872	727.140102	150.984716	0.37399647
Poly10-oleate	SMA 725™	0.80395887	415.317226	163.718284	0.32417095
Poly-10-palmitate	SMA 725™	0.55378333	220.557022	1	0.34232087
Poly-10-stearate	SMA 725™	0.60420715	666.142483	37.0384979	0.01112445
Span 20™	SMA 725™	0.62848573	283.912697	107.582548	0.07211668
Stepan Mild L3™	SMA 725™	0.70260988	92.7403599	23.4580717	0.14619491
Stepan SLL FB™	SMA 725™	0.79445623	88.9172839	27.0557157	0.55123733
Triton X-100™	SMA 725™	0.51049104	47.0513106	16.1200228	0.38193929
Triton X-305™	SMA 725™	0.1225251	34.2615725	24.79289	0.12988317
Triton X-45™	SMA 725™	0.59014722	42.8041791	9.32698688	0.34259304

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
Tween 65™	SMA 725™	0.97100901	58.7517888	4.05623277	0.72301593
Tween 80™	SMA 725™	0.47732613	15.04405	6.27578768	0.39298946
VDISTILL DV53™	SMA 725™	0.39229689	87.5663909	28.213039	0.23295534
Amin GCK 30H	SMA Cumene	0.16903289	56.1552878	40.0446791	0.17652471
Amphosol 2CSF™	SMA Cumene	0.18654925	530.682194	105.573694	0.06440189
Apo 10	SMA Cumene	0.58177855	411.551568	114.32331	0.16331519
Apo 12	SMA Cumene	1.0038667	51.5463032	1	0.68938642
Coco Glucoside	SMA Cumene	0.73385895	629.210189	118.220275	0.06106245
PFDMP-O310F	SMA Cumene	0.42783337	132.346965	1.94842086	0.32052902
Genapol X080	SMA Cumene	1.07026667	27.4976239	1	0.81008762
HEGMDDE H370	SMA Cumene	1.23098434	113.823501	15.9102869	0.69184844
HEGMDE H360	SMA Cumene	1.11430738	153.280331	26.3897685	0.0453947
HEGMOE H350	SMA Cumene	0.15213026	374.958386	184.294742	0.0348972
HXEGMDDE H375	SMA Cumene	1.24640003	14.0977381	1	0.78171728
Lakeland PAE 136™	SMA Cumene	0.99696666	50.2713157	1.66259192	0.57835253
MNA C12	SMA Cumene	0.23067437	98.3880237	33.0449879	0.15126198
MNA C13	SMA Cumene	0.22119994	49.6118337	1	0.13610065
MNA C14	SMA Cumene	0.16250006	26.9122469	16.5961981	0.1679783
n-Decanoyl-L-Methionine	SMA Cumene	0.92776667	90.2420667	1	0.49123986
n-Decanoyl-L-Threonine	SMA Cumene	0.65673647	648.371055	79.4597391	0.01326363
n-Dodecanoyl-L-Histidine	SMA Cumene	0.21763416	45.1571325	9.392309	0.15028436
n-Dodecanoyl-L-Methionine	SMA Cumene	0.7225	94.3490016	1	0.3879571
n-Dodecanoyl-L-Threonine	SMA Cumene	0.6923	63.335923	3.13250227	0.25086018
n-Octanoyl-L-Methionine	SMA Cumene	1.15366359	677.077237	73.1710311	0.03997618

Table 12A. Positively Scoring Nanonets					
Surfactant	Polymer	Height	Midway (ppm)	Slope coefficient	Average error (all data)
Sodium dodecylbenzene sulfonate	SMA Cumene	0.1	33.2633392	9.9021511	0.17315999
nDDTP D342	SMA Cumene	1.07132204	278.516346	50.4682219	0.1172323
nDDTGP D323	SMA Cumene	1.16926667	99.7568612	1.0000009	0.39835953
nDDTP D335	SMA Cumene	0.23069617	258.761008	43.8357899	0.102913
nNDTP N335	SMA Cumene	1.0257	843.880546	1	0.25040799
OEGMDE O330	SMA Cumene	1.20103346	15.8664634	1	0.73328604
Poly-10-laurate	SMA Cumene	0.4544219	73.626828	13.3065884	0.32536092
Poly-10-oleate	SMA Cumene	0.85908063	56.01628	22.6214454	0.44477444
Poly-10-stearate	SMA Cumene	0.779727	660.645528	38.4206269	0.00859737
Span 20™	SMA Cumene	0.75731112	129.430534	3.84638646	0.37532063
Stepan Mild L3™	SMA Cumene	0.40778623	310.08723	51.3347099	0.15323633
Stepan SLL FB™	SMA Cumene	1.05814906	53.2020865	2.27103651	0.87997254
Stepan sulfonic acid™	SMA Cumene	0.12353299	23.2191036	6.38311742	0.16510284
TC Lab 35	SMA Cumene	0.31656598	308.812572	62.0939333	0.15751957
Triton X-100™	SMA Cumene	0.66506667	4.67141262	1	0.56561508
Triton X-305™	SMA Cumene	0.15114334	55.3408846	43.0219002	0.12379949
Triton X-45™	SMA Cumene	1.44670135	54.6973778	5.58346883	0.76893364
Tween 40™	SMA Cumene	0.1	40.2306461	5.76851145	0.11282389
Tween 65™	SMA Cumene	0.57494502	22.7324121	11.1524571	0.40483246
Tween 80™	SMA Cumene	0.13393333	26.8482731	1	0.12413461
Tween 85™	SMA Cumene	0.65256667	26.3387849	1	0.4528309

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
1,2-Hexanediol	DIBMA	0.01079653	0.15517241	C6	NA	1
Amphosol 2CSF™	DIBMA	0.31306068	0.51724138	C18	NA	2
Apo 10	DIBMA	0.47591154	0.70689655	C10	NA	1
Apo 12	DIBMA	0.94118313	0.98275862	C12	NA	1
Stepan Bio Soft D-40™	DIBMA	0.42579884	0.67241379	NA- benzene C6 ring	NA	0
Cedepal TD-403 MFLD™	DIBMA	0.13876273	0.27586207	C13	NA	1
Coco Glucoside	DIBMA	0.48378552	0.72413793	C18	NA	1
Dodecane-1-sulfonic acid	DIBMA	0.08872691	0.18965517	C12	NA	1
PFDMP-O310F	DIBMA	0.21608623	0.39655172	C8	NA	1
Genapol X080	DIBMA	0.70033174	0.82758621	C13	Y	1
HEGMDE H370	DIBMA	0.75483262	0.87931034	C12	NA	1
HEGMDE H360	DIBMA	0.72382691	0.86206897	C10	NA	1
HEGMOEH350	DIBMA	0.21048045	0.36206897	C8	NA	1
HXEGMDDE H375	DIBMA	0.58650124	0.75862069	C12	NA	1
Lakeland PAE 136™	DIBMA	0.5867875	0.77586207	C13	NA	1
Lathanol LAL™	DIBMA	0.25217819	0.44827586	C12	NA	1
MNA C12	DIBMA	0.3249164	0.53448276	C12	NA	2
MNA C13	DIBMA	0.38633275	0.62068966	C13	NA	2
MNA C14	DIBMA	0.33728709	0.55172414	C14	NA	2
n-Decanoyl-L-Methionine	DIBMA	0.71067545	0.84482759	C10	NA	1
n-Decanoyl-L-Threonine	DIBMA	0.48860273	0.74137931	C10	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
n-Dodecanoyl-L-Histidine	DIBMA	0.23744623	0.43103448	C12	NA	1
n-Dodecanoyl-L-Methionine	DIBMA	0.40840142	0.65517241	C12	NA	1
n-Dodecanoyl-L-Threonine	DIBMA	0.60439885	0.79310345	C12	NA	1
n-Octanoyl-L-Methionine	DIBMA	0.1558324	0.29310345	C8	NA	1
Sodium dodecylbenzene sulfonate	DIBMA	0.16313833	0.31034483	C12	NA	1
nDDDTp D342	DIBMA	0.92845061	0.94827586	C12	NA	1
nDDTGP D323	DIBMA	0.84352356	0.93103448	C10	NA	1
nDDTP D335	DIBMA	0.35712153	0.56896552	C10	NA	1
Ninate 411™	DIBMA	0.26177156	0.48275862	C12	NA	1
nNDDTP N335	DIBMA	0.37768065	0.5862069	C9	NA	1
OEGMDE O330	DIBMA	0.67358625	0.81034483	C12	NA	1
Poly-10-laurate	DIBMA	0.40478905	0.63793103	C12	NA	1
Poly-10-oleate	DIBMA	0.81641409	0.9137931	C18	NA	1
Poly-10-stearate	DIBMA	0.25358274	0.46551724	C18	NA	1
Span 20™	DIBMA	0.23420274	0.4137931	C12	NA	1
Stepan Mild L3™	DIBMA	0.30304127	0.5	C12	NA	1
Stepan SLL FB™	DIBMA	0.06454197	0.17241379	C12	NA	1
Stepan sulfonic acid™	DIBMA	0.18706412	0.32758621	C12	Y	1
TC Lab 35	DIBMA	0.4744805	0.68965517	C12	NA	1
Triton X-100™	DIBMA	0.76552289	0.89655172	C8	Y	1
Triton X-305™	DIBMA	0.08905549	0.20689655	C8	Y	1
Triton X-45™	DIBMA	0.93154568	0.96551724	C8	Y	1
Tween 20™	DIBMA	0.19403513	0.34482759	C12	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
Tween 40™	DIBMA	0.09920496	0.24137931	C16	NA	1
Tween 60™	DIBMA	0.12078276	0.25862069	C18	NA	1
tween 65™	DIBMA	0.38475022	0.60344828	C18	NA	3
Tween 80™	DIBMA	0.09579844	0.22413793	C18	NA	1
Tween 85™	DIBMA	1.09042548	1	C16	NA	3
1-Octane sulfonate	SMA 2000™	0.04864068	0.11864407	C8	NA	1
Amin GCK 30H	SMA 2000™	0.12021731	0.15254237	C18, C8	NA	1,1
Amphosol 2CSF™	SMA 2000™	0.4258668	0.33898305	C18	NA	2
Apo 10	SMA 2000™	0.50548314	0.38983051	C10	NA	1
Apo 12	SMA 2000™	0.7987517	0.66101695	C12	NA	1
Stepan Bio Soft D-40™	SMA 2000™	0.67057294	0.55932203	NA- benzene C6 ring	NA	0
Stepan Bio Terge AS-40™	SMA 2000™	0.66514278	0.54237288	C14	NA	1
Coco Glucoside	SMA 2000™	0.83736484	0.69491525	C18	NA	1
Dodecane-1-sulfonic acid	SMA 2000™	0.47472229	0.3559322	C12	NA	1
PFDMP-O310F	SMA 2000™	0.3251518	0.22033898	C8	NA	1
Genapol X080	SMA 2000™	1.06309757	0.88135593	C13	Y	1
HEGMDE H370	SMA 2000™	1.07438368	0.91525424	C12	NA	1
HEGMDE H360	SMA 2000™	1.10481756	0.96610169	C10	NA	1
HEGMOE H350	SMA 2000™	0.88666487	0.74576271	C8	NA	1
HXEGMDDE H375	SMA 2000™	1.2687219	1	C12	NA	1
Lakeland PAE 136™	SMA 2000™	0.66477033	0.52542373	C13	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
Stepan Lathanol LAL™	SMA 2000™	0.67356793	0.57627119	C12	NA	1
MNA C13	SMA 2000™	0.55090966	0.44067797	C13	NA	2
MNA C14	SMA 2000™	0.60503265	0.47457627	C14	NA	2
n-Decanoyl-L-Methionine	SMA 2000™	0.70238207	0.61016949	C10	NA	1
n-Decanoyl-L-Threonine	SMA 2000™	0.39777096	0.30508475	C10	NA	1
n-Dodecanoyl-L-Histidine	SMA 2000™	0.99824148	0.83050847	C12	NA	1
n-Dodecanoyl-L-Methionine	SMA 2000™	0.38529232	0.28813559	C12	NA	1
n-Dodecanoyl-L-Threonine	SMA 2000™	0.92313503	0.81355932	C12	NA	1
n-Octanoyl-L-Methionine	SMA 2000™	0.35486668	0.27118644	C8	NA	1
Sodium dodecylbenzene sulfonate	SMA 2000™	0.77941799	0.6440678	C12	NA	1
nDDTP D342	SMA 2000™	1.03238155	0.84745763	C12	NA	1
nDDTGP D323	SMA 2000™	1.06342351	0.89830508	C10	NA	1
nDDTP D335	SMA 2000™	0.75110832	0.62711864	C10	NA	1
Ninate 411™	SMA 2000™	0.49095719	0.37288136	C12	NA	1
nNDTP N335	SMA 2000™	0.54366513	0.42372881	C9	NA	1
OEGMDE O330	SMA 2000™	1.07746121	0.93220339	C12	NA	1
Poly-10-laurate	SMA 2000™	1.09611792	0.94915254	C12	NA	1
Poly-10-oleate	SMA 2000™	0.34720409	0.25423729	C18	NA	1
Poly-10-stearate	SMA 2000™	0.34135375	0.23728814	C18	NA	1
Span 20™	SMA 2000™	0.6962221	0.59322034	C12	NA	1
Stepan Mild L3™	SMA 2000™	0.53967414	0.40677966	C12	NA	1
Stepan SLL FB™	SMA 2000™	0.89518363	0.76271186	C12	NA	1
Stepan sulfonic acid™	SMA 2000™	0.56522096	0.45762712	C12	Y	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
TC Lab 35	SMA 2000™	0.8108317	0.6779661	C12	NA	1
Triton X-100™	SMA 2000™	1.05693912	0.86440678	C8	Y	1
Triton X-305™	SMA 2000™	0.39871505	0.3220339	C8	Y	1
Triton X-45™	SMA 2000™	0.66079039	0.50847458	C8	Y	1
Tween 20™	SMA 2000™	0.87847365	0.72881356	C12	NA	1
Tween 40™	SMA 2000™	0.85312531	0.71186441	C16	NA	1
Tween 60™	SMA 2000™	0.89719482	0.77966102	C18	NA	1
Tween 65™	SMA 2000™	0.91446164	0.79661017	C18	NA	3
Tween 80™	SMA 2000™	0.63839541	0.49152542	C18	NA	1
Tween 85™	SMA 2000™	1.13795748	0.98305085	C16	NA	3
VDISTILL DV53™	SMA 2000™	0.12917714	0.16949153	C18, C18	NA	11
VDISTILL DV63™	SMA 2000™	0.13828377	0.18644068	C18, C18	NA	11
Sodium-1-octane sulfonate	SMA 2021™	0.01470987	0.16814159	C8	NA	1
Amin GCK 30H	SMA 2021™	0.14060286	0.22123894	C18, C8	NA	1,1
Amphosol 2CSF™	SMA 2021™	0.35471071	0.45132743	C18	NA	2
Anthraquinone-2-sulfonic acid	SMA 2021™	0.08560249	0.18584071	NA aromatic	NA	0
Apo 10	SMA 2021™	0.23534902	0.2920354	C10	NA	1
Apo 12	SMA 2021™	0.41506008	0.52212389	C12	NA	1
Cedepal TD-403 MFLD™	SMA 2021™	0.25601338	0.36283186	C13	NA	1
Coco Glucoside	SMA 2021™	0.78031574	0.73451327	C18	NA	1
Dodecane-1-sulfonic acid	SMA 2021™	0.23883696	0.30973451	C12	NA	1
PFDMP O310F	SMA 2021™	0.2431645	0.32743363	C8	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
Genapol X080	SMA 2021™	0.88125934	0.78761062	C13	Y	1
HEGMDE H370	SMA 2021™	1.29526847	0.98230088	C12	NA	1
HEGMDE H360	SMA 2021™	1.22044406	0.92920354	C10	NA	1
HEGMOE H350	SMA 2021™	1.20311221	0.91150442	C8	NA	1
HXEGMDE H375	SMA 2021™	1.26420953	0.94690265	C12	NA	1
Lakeland PAE 136™	SMA 2021™	1.29580436	1	C13	NA	1
Stapan Lathanol LAL™	SMA 2021™	0.49459328	0.55752212	C12	NA	1
MNA C13	SMA 2021™	0.20799401	0.27433628	C13	NA	2
MNA C14	SMA 2021™	0.56369824	0.6460177	C14	NA	2
n-Decanoyl-L-Methionine	SMA 2021™	0.81299992	0.75221239	C10	NA	1
n-Decanoyl-L-Threonine	SMA 2021™	0.26698434	0.38053097	C10	NA	1
n-Dodecanoyl-L-Methionine	SMA 2021™	0.36102049	0.46902655	C12	NA	1
n-Dodecanoyl-L-Threonine	SMA 2021™	0.82418296	0.7699115	C12	NA	1
n-Octanoyl-L-Methionine	SMA 2021™	0.29301188	0.39823009	C8	NA	1
Sodium dodecylbenzene sulfonate	SMA 2021™	0.91587432	0.82300885	C12	NA	1
nDDTP D342	SMA 2021™	0.91122883	0.80530973	C12	NA	1
nDDTGP D323	SMA 2021™	0.45133615	0.53982301	C10	NA	1
nDDTP D335	SMA 2021™	0.51566509	0.57522124	C10	NA	1
Ninate 411™	SMA 2021™	0.7719462	0.69911504	C12	NA	1
nNDTP N335	SMA 2021™	0.39107611	0.48672566	C9	NA	1
OEGMDE O330	SMA 2021™	1.29520205	0.96460177	C12	NA	1
Poly-10-laurate	SMA 2021™	1.08248266	0.87610619	C12	NA	1
Poly-10-oleate	SMA 2021™	1.09001173	0.89380531	C18	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
Poly-10-palmitate	SMA 2021™	0.16112024	0.23893805	C16	NA	1
Poly-10-stearate	SMA 2021™	0.16325242	0.25663717	C18	NA	1
Span 20™	SMA 2021™	0.24952947	0.34513274	C12	NA	1
Stapan Mild L3™	SMA 2021™	0.52137185	0.61061947	C12	NA	1
Stapan SLL FB™	SMA 2021™	0.77624393	0.71681416	C12	NA	1
Stapan sulfonic acid™	SMA 2021™	0.51778871	0.59292035	C12	Y	1
TC Lab 35	SMA 2021™	0.59035451	0.66371681	C12	NA	1
Triton X-100™	SMA 2021™	0.41340161	0.50442478	C8	Y	1
Triton X-305™	SMA 2021™	0.13348571	0.20353982	C8	Y	1
Triton X-45™	SMA 2021™	0.54179759	0.62831858	C8	Y	1
Tween 40™	SMA 2021™	0.31191032	0.4159292	C16	NA	1
Tween 60™	SMA 2021™	0.59247768	0.68141593	C18	NA	1
Tween 80™	SMA 2021™	1.05503229	0.85840708	C18	NA	1
Tween 85™	SMA 2021™	0.94429643	0.84070796	C16	NA	3
Amphosol 2CSF™	SMA 3000™	0.1489152	0.34545455	C18	NA	2
Anthraquinone-2-sulfonic acid	SMA 3000™	0.06904032	0.25454545	NA aromatic	NA	0
Apo 10	SMA 3000™	0.39176322	0.58181818	C10	NA	1
Apo 12	SMA 3000™	0.62620128	0.70909091	C12	NA	1
Coco Glucoside	SMA 3000™	0.93381513	0.87272727	C18	NA	1
PFDMP O310F	SMA 3000™	0.00769877	0.2	C8	NA	1
Genapol X080	SMA 3000™	1.35561809	0.96363636	C13	Y	1
HEGMDE H370	SMA 3000™	0.93388926	0.89090909	C12	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
HEGMDE H360	SMA 3000™	0.97713596	0.90909091	C10	NA	1
HEGMOE H350	SMA 3000™	0.56612221	0.67272727	C8	NA	1
HXEGMDDE H375	SMA 3000™	1.02583227	0.94545455	C12	NA	1
Lakeland PAE 136™	SMA 3000™	0.64799652	0.72727273	C13	NA	1
Stepan Lathanol LAL™	SMA 3000™	0.13921714	0.30909091	C12	NA	1
MNA C12	SMA 3000™	0.34875182	0.52727273	C12	NA	2
MNA C13	SMA 3000™	0.32243256	0.49090909	C13	NA	2
MNA C14	SMA 3000™	0.2925184	0.47272727	C14	NA	2
n-Decanoyl-L-Methionine	SMA 3000™	0.382233	0.56363636	C10	NA	1
n-Decanoyl-L-Threonine	SMA 3000™	0.28167062	0.43636364	C10	NA	1
n-Dodecanoyl-L-Histidine	SMA 3000™	0.16597792	0.36363636	C12	NA	1
n-Dodecanoyl-L-Methionine	SMA 3000™	0.47669814	0.6	C12	NA	1
n-Dodecanoyl-L-Threonine	SMA 3000™	0.78408718	0.83636364	C12	NA	1
n-Octanoyl-L-Methionine	SMA 3000™	0.28058523	0.41818182	C8	NA	1
Sodium dodecylbenzene sulfonate	SMA 3000™	0.08967674	0.27272727	C12	NA	1
nDDTP D342	SMA 3000™	0.66968033	0.74545455	C12	NA	1
nDDTGP D323	SMA 3000™	0.86331518	0.85454545	C10	NA	1
nDDTP D335	SMA 3000™	0.47780789	0.61818182	C10	NA	1
Ninate 411™	SMA 3000™	0.28448763	0.45454545	C12	NA	1
nNDTP N335	SMA 3000™	0.33141816	0.50909091	C9	NA	1
nODTTP O320	SMA 3000™	0.05281838	0.23636364	C8	NA	1
OEGMDE O330	SMA 3000™	0.99788957	0.92727273	C12	NA	1
Poly-10-laurate	SMA 3000™	0.70717371	0.78181818	C12	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
Poly-10-oleate	SMA 3000™	0.20373552	0.38181818	C18	NA	1
Poly-10-palmitate	SMA 3000™	0.54474114	0.65454545	C16	NA	1
Poly-10-stearate	SMA 3000™	0.37308656	0.54545455	C18	NA	1
Span 20™	SMA 3000™	0.68594116	0.76363636	C12	NA	1
Stepan Mild L3™	SMA 3000™	0.48349829	0.63636364	C12	NA	1
Stepan SLL FB™	SMA 3000™	0.73250601	0.8	C12	NA	1
Stepan sulfonic acid™	SMA 3000™	0.23457864	0.4	C12	Y	1
TC Lab 35	SMA 3000™	0.14866913	0.32727273	C12	NA	1
Triton X-100™	SMA 3000™	0.59926377	0.69090909	C8	Y	1
Triton X-305™	SMA 3000™	0.13352979	0.29090909	C8	Y	1
Triton X-45™	SMA 3000™	0.73537752	0.81818182	C8	Y	1
Tween 65™	SMA 3000™	1.5270864	1	C18	NA	3
Tween 85™	SMA 3000™	1.49791049	0.98181818	C16	NA	3
Amin GCK 30H	SMA-IBE	0.13080691	0.63636364	C18, C8	NA	1,1
Amphosol 2CSF™	SMA-IBE	0.04431075	0.37373737	C18	NA	2
Apo 10	SMA-IBE	0.13055103	0.61616162	C10	NA	1
Apo 12	SMA-IBE	0.5294656	0.95959596	C12	NA	1
Stepan Bio Soft D-40™	SMA-IBE	0.06688198	0.45454545	NA- benzene C6 ring	NA	0
Coco Glucoside	SMA-IBE	0.20882744	0.75757576	C18	NA	1
Fluoro octyl maltoside O310F	SMA-IBE	0.05203224	0.39393939	C8	NA	1
HEGMIDE H370	SMA-IBE	0.45706561	0.89898989	C12	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
HEGMDE H360	SMA-IBE	0.51215139	0.93939394	C10	NA	1
HXEGMDDE H375	SMA-IBE	0.82075754	0.97979798	C12	NA	1
Lakeland PAE 136™	SMA-IBE	0.13333097	0.65656566	C13	NA	1
MNA C14	SMA-IBE	0.18234299	0.69696967	C14	NA	2
n-Decanoyl-L-Methionine	SMA-IBE	0.8743853	1	C10	NA	1
n-Decanoyl-L-Threonine	SMA-IBE	0.17561649	0.67676768	C10	NA	1
n-Dodecanoyl-L-Histidine	SMA-IBE	0.09908736	0.51515152	C12	NA	1
n-Dodecanoyl-L-Methionine	SMA-IBE	0.43202715	0.87878788	C12	NA	1
n-Dodecanoyl-L-Threonine	SMA-IBE	0.09546425	0.49494949	C12	NA	1
n-Octanoyl-L-Methionine	SMA-IBE	0.18338342	0.71717172	C8	NA	1
nDDTTP D342	SMA-IBE	0.21636181	0.79797978	C12	NA	1
nDDTGP D323	SMA-IBE	0.07330399	0.47474747	C10	NA	1
nDDTTP D335	SMA-IBE	0.21277188	0.77777778	C10	NA	1
OEGMDE O330	SMA-IBE	0.11103633	0.55555556	C12	NA	1
Poly-10-laurate	SMA-IBE	0.10433733	0.53535354	C12	NA	1
Poly-10-oleate	SMA-IBE	0.37458027	0.83838384	C18	NA	1
Span 20™	SMA-IBE	0.05593784	0.41414141	C12	NA	1
Stapan Mild L3™	SMA-IBE	0.37705681	0.85858586	C12	NA	1
Stapan SLL FB™	SMA-IBE	0.48079671	0.91919192	C12	NA	1
Stapan sulfonic acid™	SMA-IBE	0.1144461	0.57575758	C12	Y	1
Triton X-305™	SMA-IBE	0.05866763	0.43434343	C8	Y	1
Triton X-45™	SMA-IBE	0.30935824	0.81818182	C8	Y	1
Tween 60™	SMA-IBE	0.03869585	0.35353535	C18	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
Tween 65™	SMA-IBE	0.12881485	0.5959596	C18	NA	3
Tween 85™	SMA-IBE	0.20361205	0.73737374	C16	NA	3
1,2-Hexanediol	SMA 725™	0.15337869	0.40740741	C6	NA	1
Amin GCK 30H	SMA 725™	0.12788222	0.37037037	C18, C8	NA	1,1
Amphosol 2CSF™	SMA 725™	0.0592602	0.24074074	C18	NA	2
Apo 10	SMA 725™	0.14177449	0.38888889	C10	NA	1
Apo 12	SMA 725™	0.1616708	0.44444444	C12	NA	1
Stepan Bio Soft D-40™	SMA 725™	0.47579589	0.77777778	NA- benzene C6 ring	NA	0
Stepan Bio Terge AS-40™	SMA 725™	0.49735177	0.81481481	C14	NA	1
Dodecane-1-sulfonic acid	SMA 725™	0.35107569	0.59259259	C12	NA	1
PFDMP-O310F	SMA 725™	0.06112487	0.25925926	C8	NA	1
Genapol X080	SMA 725™	1.10987513	1	C13	Y	1
HEGMIDDE H370	SMA 725™	0.40115456	0.68518519	C12	NA	1
HEGMIDE H360	SMA 725™	0.93509538	0.98148148	C10	NA	1
HXEGMIDDE H375	SMA 725™	0.30848282	0.57407407	C12	NA	1
Lakeland PAE 136™	SMA 725™	0.39836812	0.66666667	C13	NA	1
Stepan Lathanol LAL™	SMA 725™	0.64861222	0.90740741	C12	NA	1
MNA C12	SMA 725™	0.10799358	0.33333333	C12	NA	2
MNA C14	SMA 725™	0.20121167	0.5	C14	NA	2
n-Decanoyl-L-Methionine	SMA 725™	0.55741713	0.83333333	C10	NA	1
n-Decanoyl-L-Threonine	SMA 725™	0.1819542	0.46296296	C10	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
n-Dodecanoyl-L-Histidine	SMA 725 TM	0.15380139	0.42592593	C12	NA	1
n-Dodecanoyl-L-Methionine	SMA 725 TM	0.80065107	0.94444444	C12	NA	1
n-Dodecanoyl-L-Threonine	SMA 725 TM	0.6146695	0.87037037	C12	NA	1
n-Octanoyl-L-Methionine	SMA 725 TM	0.26170605	0.53703704	C8	NA	1
nDDDT D342	SMA 725 TM	0.30132153	0.55555556	C12	NA	1
nDDTGP D323	SMA 725 TM	0.36140299	0.62962963	C10	NA	1
nDDTP D335	SMA 725 TM	0.05901839	0.22222222	C10	NA	1
Ninate 411 TM	SMA 725 TM	0.10077397	0.31481481	C12	NA	1
nNDTP N335	SMA 725 TM	0.07587077	0.27777778	C9	NA	1
OEGMIDE O330	SMA 725 TM	0.19608992	0.48148148	C12	NA	1
Poly-10-laurate	SMA 725 TM	0.36182575	0.64814815	C12	NA	1
Poly10-oleate	SMA 725 TM	0.46410887	0.74074074	C18	NA	1
Poly-10-palmitate	SMA 725 TM	0.43191942	0.7037037	C16	NA	1
Poly-10-stearate	SMA 725 TM	0.20202389	0.51851852	C18	NA	1
Span 20 TM	SMA 725 TM	0.44580726	0.72222222	C12	NA	1
Stepan Mild L3 TM	SMA 725 TM	0.6374942	0.88888889	C12	NA	1
Stepan SLL FB TM	SMA 725 TM	0.72343785	0.92592593	C12	NA	1
Triton X-100 TM	SMA 725 TM	0.48630719	0.7962963	C8	Y	1
Triton X-305 TM	SMA 725 TM	0.11772017	0.35185185	C8	Y	1
Triton X-45 TM	SMA 725 TM	0.5651288	0.85185185	C8	Y	1
Tween 65 TM	SMA 725 TM	0.914446	0.96296296	C18	NA	3
Tween 80 TM	SMA 725 TM	0.47014245	0.75925926	C18	NA	1
VDISTILL DV53 TM	SMA 725 TM	0.35766349	0.61111111	C18, C18	NA	11

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
Amin GCK 30H	SMA Cumene	0.15815307	0.37037037	C18, C8	NA	1,1
Amphosol 2CSF™	SMA Cumene	0.08774462	0.22222222	C18	NA	2
Apo 10	SMA Cumene	0.34123615	0.59259259	C10	NA	1
Apo 12	SMA Cumene	0.95262301	0.87037037	C12	NA	1
Coco Glucoside	SMA Cumene	0.27572738	0.55555556	C18	NA	1
PFDMP O310F	SMA Cumene	0.37142484	0.61111111	C8	NA	1
Genapol X080	SMA Cumene	1.04137201	0.90740741	C13	Y	1
HEGMDE H370	SMA Cumene	1.09147006	0.94444444	C12	NA	1
HEGMDE H360	SMA Cumene	0.94397663	0.83333333	C10	NA	1
HEGMOE H350	SMA Cumene	0.09265332	0.24074074	C8	NA	1
HXEGMDE H375	SMA Cumene	1.22945181	0.98148148	C12	NA	1
Lakeland PAE 136™	SMA Cumene	0.94734632	0.85185185	C13	NA	1
MNA C12	SMA Cumene	0.20772105	0.42592593	C12	NA	2
MNA C13	SMA Cumene	0.21033641	0.46296296	C13	NA	2
MNA C14	SMA Cumene	0.1577351	0.35185185	C14	NA	2
n-Decanoyl-L-Methionine	SMA Cumene	0.84450697	0.81481481	C10	NA	1
n-Decanoyl-L-Threonine	SMA Cumene	0.23185822	0.51851852	C10	NA	1
n-Dodecanoyl-L-Histidine	SMA Cumene	0.20789949	0.44444444	C12	NA	1
n-Dodecanoyl-L-Methionine	SMA Cumene	0.6546941	0.72222222	C12	NA	1
n-Dodecanoyl-L-Threonine	SMA Cumene	0.64879869	0.7037037	C12	NA	1
n-Octanoyl-L-Methionine	SMA Cumene	0.37412274	0.62962963	C8	NA	1
Sodium dodecylbenzene sulfonate	SMA Cumene	0.09669148	0.27777778	C12	NA	1
nDDDTp D342	SMA Cumene	0.77326272	0.77777778	C12	NA	1

Table 12B. Positively Scoring Nanonets						
Surfactant	Polymer	Sigmoid AUC (nanonet score)	Score polymer specific	Alkyl chain length	Alkyl chain branched	Number of alkyl chains
nDDTGP D323	SMA Cumene	1.05320893	0.92592593	C10	NA	1
nDDTP D335	SMA Cumene	0.17108907	0.40740741	C10	NA	1
nNDTP N335	SMA Cumene	0.16064457	0.38888889	C9	NA	1
OEGMIDE O330	SMA Cumene	1.18257782	0.96296296	C12	NA	1
Poly-10-laurate	SMA Cumene	0.42116849	0.64814815	C12	NA	1
Poly-10-oleate	SMA Cumene	0.80985223	0.7962963	C18	NA	1
Poly-10-stearate	SMA Cumene	0.26499802	0.53703704	C18	NA	1
Span 20™	SMA Cumene	0.65967059	0.74074074	C12	NA	1
Stepan Mild L3™	SMA Cumene	0.28149156	0.57407407	C12	NA	1
Stepan SLL FB™	SMA Cumene	1.0023824	0.88888889	C12	NA	1
Stepan sulfonic acid™	SMA Cumene	0.12070749	0.2962963	C12	Y	1
TC Lab 35	SMA Cumene	0.21883052	0.48148148	C12	NA	1
Triton X-100™	SMA Cumene	0.66228879	0.75925926	C8	Y	1
Triton X-305™	SMA Cumene	0.14128455	0.33333333	C8	Y	1
Triton X-45™	SMA Cumene	1.36829352	1	C8	Y	1
Tween 40™	SMA Cumene	0.09602644	0.25925926	C16	NA	1
Tween 65™	SMA Cumene	0.56141023	0.66666667	C18	NA	3
Tween 80™	SMA Cumene	0.13040442	0.31481481	C18	NA	1
Tween 85™	SMA Cumene	0.63570514	0.68518519	C16	NA	3

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
1,2-Hexanediol	DIBMA	Anionic	-1	Y	N	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
Amphosol 2CSF™	DIBMA	Amphoteric	1	N	Y	0
Apo 10	DIBMA	Non-ionic	0	Y	N	0
Apo 12	DIBMA	Non-ionic	0	Y	N	0
Stepan Bio Soft D-40™	DIBMA	Anionic	-1	Y	Y	0
Cedepal TD-403 MFLD™	DIBMA	Anionic	-1	Y	N	0
Coco Glucoside	DIBMA	Non-ionic	0	N C6 ring	Y	0
Dodecane-1-sulfonic acid	DIBMA	Anionic	-1	Y	N	0
PFDMP O310F	DIBMA	Non-ionic	0	N	Y	0
Genapol X080	DIBMA	Non-ionic	0	Y	N	0
HEGMDDE H370	DIBMA	Non-ionic	0	Y	N	0
HEGMDE H360	DIBMA	Non-ionic	0	Y	N	0
HEGMOEH350	DIBMA	Non-ionic	0	Y	N	0
HXEGMDDE H375	DIBMA	Non-ionic	0	Y	N	0
Lakeland PAE 136™	DIBMA	Anionic	-1	Y	N	0
Stepan Lathanol LAL™	DIBMA	Anionic	-1	Y	N	0
MNA C12	DIBMA	Non-ionic	0	N	Y	0
MNA C13	DIBMA	Non-ionic	0	N	Y	0
MNA C14	DIBMA	Non-ionic	0	N	Y	0
n-Decanoyl-L-Methionine	DIBMA	Anionic	-1	N	Y	0
n-Decanoyl-L-Threonine	DIBMA	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Histidine	DIBMA	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Methionine	DIBMA	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Threonine	DIBMA	Anionic	-1	N	Y	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
n-Octanoyl-L-Methionine	DIBMA	Anionic	-1	N	Y	0
Sodium dodecylbenzene sulfonate	DIBMA	Anionic	-1	N	Y	0
nDDTP D342	DIBMA	Non-ionic	0	N	Y	0
nDDTGP D323	DIBMA	Non-ionic	0	N	Y	0
nDDTP D335	DIBMA	Non-ionic	0	N	Y	0
Ninate 411™	DIBMA	Anionic	-1	N	Y	0
nndtp n335	DIBMA	Non-ionic	0	N	N	0
oegmde o330	DIBMA	Non-ionic	0	Y	N	0
Poly-10-laurate	DIBMA	Non-ionic	0	N	Y	0
Poly-10-oleate	DIBMA	Non-ionic	0	N	Y	1
Poly-10-stearate	DIBMA	Non-ionic	0	N	Y	0
Span 20™	DIBMA	Non-ionic	0	N	N	0
Stepan Mild L3™	DIBMA	Anionic	-1	Y	N	0
Stepan SLL FB™	DIBMA	Anionic	-1	N	Y	0
Stepan sulfonic acid™	DIBMA	Anionic	-1	Y	N	0
TC Lab 35	DIBMA	Amphoteric	1	N	Y	0
Triton X-100™	DIBMA	Non-ionic	0	N	Y	0
Triton X-305™	DIBMA	Non-ionic	0	N	Y	0
Triton X-45™	DIBMA	Non-ionic	0	N	Y	0
Tween 20™	DIBMA	Non-ionic	0	N	Y (4)	0
Tween 40™	DIBMA	Non-ionic	0	N	Y (4)	0
Tween 60™	DIBMA	Non-ionic	0	N	Y (4)	0
tween 65™	DIBMA	Non-ionic	0	N	Y (4)	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
Tween 80™	DIBMA	Non-ionic	0	N	Y (4)	1
Tween 85™	DIBMA	Non-ionic	0	N	Y (4)	0
1-Octane sulfonate	SMA 2000™	Anionic	-1	Y	N	0
Amin GCK 30H	SMA 2000™	Anionic	-1	Y	N	1
Amphosol 2CSF™	SMA 2000™	amphoteric	1	N	Y	0
Apo 10	SMA 2000™	Non-ionic	0	Y	N	0
Apo 12	SMA 2000™	Non-ionic	0	Y	N	0
Stepan Bio Soft D-40™	SMA 2000™	Anionic	-1	Y	Y	0
Stepan Bio Terge AS-40™	SMA 2000™	Anionic	-1	Y	N	1
Coco Glucoside	SMA 2000™	Non-ionic	0	N C6 ring	Y	0
Dodecane-1-sulfonic acid	SMA 2000™	Anionic	-1	Y	N	0
PFDMP O310F	SMA 2000™	Non-ionic	0	N	Y	0
Genapol X080	SMA 2000™	Non-ionic	0	Y	N	0
HEGMDE H370	SMA 2000™	Non-ionic	0	Y	N	0
HEGMDE H360	SMA 2000™	Non-ionic	0	Y	N	0
HEGMOEH350	SMA 2000™	Non-ionic	0	Y	N	0
HXEGMDDE H375	SMA 2000™	Non-ionic	0	Y	N	0
Lakeland PAE 136™	SMA 2000™	Anionic	-1	Y	N	0
Stepan Lathanol LAL™	SMA 2000™	Anionic	-1	Y	N	0
MNA C13	SMA 2000™	Non-ionic	0	N	Y	0
MNA C14	SMA 2000™	Non-ionic	0	N	Y	0
n-Decanoyl-L-Methionine	SMA 2000™	Anionic	-1	N	Y	0
n-Decanoyl-L-Threonine	SMA 2000™	Anionic	-1	N	Y	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
n-Dodecanoyl-L-Histidine	SMA 2000™	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Methionine	SMA 2000™	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Threonine	SMA 2000™	Anionic	-1	N	Y	0
n-Octanoyl-L-Methionine	SMA 2000™	Anionic	-1	N	Y	0
Sodium dodecylbenzene sulfonate	SMA 2000™	Anionic	-1	N	Y	0
nDDTP D342	SMA 2000™	Non-ionic	0	N	Y	0
nDDTGP D323	SMA 2000™	Non-ionic	0	N	Y	0
nDDTP D335	SMA 2000™	Non-ionic	0	N	Y	0
Ninate 411™	SMA 2000™	Anionic	-1	N	Y	0
nNDTP N335	SMA 2000™	Non-ionic	0	N	N	0
OEGMDE O330	SMA 2000™	Non-ionic	0	Y	N	0
Poly-10-laurate	SMA 2000™	Non-ionic	0	N	Y	0
Poly-10-oleate	SMA 2000™	Non-ionic	0	N	Y	1
Poly-10-stearate	SMA 2000™	Non-ionic	0	N	Y	0
Span 20™	SMA 2000™	Non-ionic	0	N	N	0
Stepan Mild L3™	SMA 2000™	Anionic	-1	Y	N	0
Stepan SLL FB™	SMA 2000™	Anionic	-1	N	Y	0
Stepan sulfonic acid™	SMA 2000™	Anionic	-1	Y	N	0
TC Lab 35	SMA 2000™	Amphoteric	1	N	Y	0
Triton X-100™	SMA 2000™	Non-ionic	0	N	Y	0
Triton X-305™	SMA 2000™	Non-ionic	0	N	Y	0
Triton X-45™	SMA 2000™	Non-ionic	0	N	Y	0
Tween 20™	SMA 2000™	Non-ionic	0	N	Y (4)	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
Tween 40™	SMA 2000™	Non-ionic	0	N	Y(4)	0
Tween 60™	SMA 2000™	Non-ionic	0	N	Y(4)	0
Tween 65™	SMA 2000™	Non-ionic	0	N	Y(4)	0
Tween 80™	SMA 2000™	Non-ionic	0	N	Y(4)	1
Tween 85™	SMA 2000™	Non-ionic	0	N	Y(4)	0
VDISTILL DV53™	SMA 2000™	Non-ionic	0	Y	N	1, 3
VDISTILL DV63™	SMA 2000™	Non-ionic	0	Y	N	1, 3
1 octane sulfonate	SMA 2021™	Anionic	-1	Y	N	0
Amin GCK 30H	SMA 2021™	Anionic	-1	Y	N	1
Amphosol 2CSF™	SMA 2021™	amphoteric	1	N	Y	0
Anthraquinone-2-sulfonic acid	SMA 2021™	Anionic	-1	Y	Y	0
Apo 10	SMA 2021™	Non-ionic	0	Y	N	0
Apo 12	SMA 2021™	Non-ionic	0	Y	N	0
Cedepal TD-403 MFLD™	SMA 2021™	Anionic	-1	Y	N	0
Coco Glucoside	SMA 2021™	Non-ionic	0	N C6 ring	Y	0
Dodecane-1-sulfonic acid	SMA 2021™	Anionic	-1	Y	N	0
PFDMP O310F	SMA 2021™	Non-ionic	0	N	Y	0
Genapol X080	SMA 2021™	Non-ionic	0	Y	N	0
HEGMDDE H370	SMA 2021™	Non-ionic	0	Y	N	0
HEGMDE H360	SMA 2021™	Non-ionic	0	Y	N	0
HEGMOE H350	SMA 2021™	Non-ionic	0	Y	N	0
HXEGMDDE H375	SMA 2021™	Non-ionic	0	Y	N	0
Lakeland PAE 136™	SMA 2021™	Anionic	-1	Y	N	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
Stepan Lathanol LAL™	SMA 2021™	Anionic	-1	Y	N	0
MNA C13	SMA 2021™	Non-ionic	0	N	Y	0
MNA C14	SMA 2021™	Non-ionic	0	N	Y	0
n-Decanoyl-L-Methionine	SMA 2021™	Anionic	-1	N	Y	0
n-Decanoyl-L-Threonine	SMA 2021™	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Methionine	SMA 2021™	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Threonine	SMA 2021™	Anionic	-1	N	Y	0
n-Octanoyl-L-Methionine	SMA 2021™	Anionic	-1	N	Y	0
Sodium dodecylbenzene sulfonate	SMA 2021™	Anionic	-1	N	Y	0
nDDTP D342	SMA 2021™	Non-ionic	0	N	Y	0
nDDTGP D323	SMA 2021™	Non-ionic	0	N	Y	0
nDDTP D335	SMA 2021™	Non-ionic	0	N	Y	0
Ninate 411™	SMA 2021™	Anionic	-1	N	Y	0
nNDTP N335	SMA 2021™	Non-ionic	0	N	N	0
OEGMDE O330	SMA 2021™	Non-ionic	0	Y	N	0
Poly-10-laurate	SMA 2021™	Non-ionic	0	N	Y	0
Poly-10-oleate	SMA 2021™	Non-ionic	0	N	Y	1
Poly-10-palmitate	SMA 2021™	Non-ionic	0	N	Y	0
Poly-10-stearate	SMA 2021™	Non-ionic	0	N	Y	0
Span 20™	SMA 2021™	Non-ionic	0	N	N	0
Stepan Mild L3™	SMA 2021™	Anionic	-1	Y	N	0
Stepan SLL FB™	SMA 2021™	Anionic	-1	N	Y	0
Stepan sulfonic acid™	SMA 2021™	Anionic	-1	Y	N	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
TC Lab 35	SMA 2021™	Amphoteric	1	N	Y	0
Triton X-100™	SMA 2021™	Non-ionic	0	N	Y	0
Triton X-305™	SMA 2021™	Non-ionic	0	N	Y	0
Triton X-45™	SMA 2021™	Non-ionic	0	N	Y	0
Tween 40™	SMA 2021™	Non-ionic	0	N	Y(4)	0
Tween 60™	SMA 2021™	Non-ionic	0	N	Y(4)	0
Tween 80™	SMA 2021™	Non-ionic	0	N	Y(4)	1
Tween 85™	SMA 2021™	Non-ionic	0	N	Y(4)	0
Amphosol 2CSF™	SMA 3000™	amphoteric	1	N	Y	0
Anthraquinone-2-sulfonic acid	SMA 3000™	Anionic	-1	Y	Y	0
Apo 10	SMA 3000™	Non-ionic	0	Y	N	0
Apo 12	SMA 3000™	Non-ionic	0	Y	N	0
Coco Glucoside	SMA 3000™	Non-ionic	0	N C6 ring	Y	0
PFDMP O310F	SMA 3000™	Non-ionic	0	N	Y	0
Genapol X080	SMA 3000™	Non-ionic	0	Y	N	0
HEGMDDE H370	SMA 3000™	Non-ionic	0	Y	N	0
HEGMDE H360	SMA 3000™	Non-ionic	0	Y	N	0
HEGMOE H350	SMA 3000™	Non-ionic	0	Y	N	0
HXEGMDDE H375	SMA 3000™	Non-ionic	0	Y	N	0
Lakeland PAE 136™	SMA 3000™	Anionic	-1	Y	N	0
Stepan Lathanol LAL™	SMA 3000™	Anionic	-1	Y	N	0
MNA C12	SMA 3000™	Non-ionic	0	N	Y	0
MNA C13	SMA 3000™	Non-ionic	0	N	Y	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
MNA C14	SMA 3000™	Non-ionic	0	N	Y	0
n-Decanoyl-L-Methionine	SMA 3000™	Anionic	-1	N	Y	0
n-Decanoyl-L-Threonine	SMA 3000™	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Histidine	SMA 3000™	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Methionine	SMA 3000™	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Threonine	SMA 3000™	Anionic	-1	N	Y	0
n-Octanoyl-L-Methionine	SMA 3000™	Anionic	-1	N	Y	0
Sodium dodecylbenzene sulfonate	SMA 3000™	Anionic	-1	N	Y	0
nDDDTp D342	SMA 3000™	Non-ionic	0	N	Y	0
nDDTGP D323	SMA 3000™	Non-ionic	0	N	Y	0
nDDTP D335	SMA 3000™	Non-ionic	0	N	Y	0
Ninate 411™	SMA 3000™	Anionic	-1	N	Y	0
nNDTP N335	SMA 3000™	Non-ionic	0	N	N	0
nODTTP O320	SMA 3000™	Non-ionic	0	N	Y	0
OEGMDE O330	SMA 3000™	Non-ionic	0	Y	N	0
Poly-10-laurate	SMA 3000™	Non-ionic	0	N	Y	0
Poly-10-oleate	SMA 3000™	Non-ionic	0	N	Y	1
Poly-10-palmitate	SMA 3000™	Non-ionic	0	N	Y	0
Poly-10-stearate	SMA 3000™	Non-ionic	0	N	Y	0
Span 20™	SMA 3000™	Non-ionic	0	N	N	0
Stepan Mild L3™	SMA 3000™	Anionic	-1	Y	N	0
Stepan SLL FB™	SMA 3000™	Anionic	-1	N	Y	0
Stepan sulfonic acid™	SMA 3000™	Anionic	-1	Y	N	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
TC Lab 35	SMA 3000™	Amphoteric	1	N	Y	0
Triton X-100™	SMA 3000™	Non-ionic	0	N	Y	0
Triton X-305™	SMA 3000™	Non-ionic	0	N	Y	0
Triton X-45™	SMA 3000™	Non-ionic	0	N	Y	0
Tween 65™	SMA 3000™	Non-ionic	0	N	Y(4)	0
Tween 85™	SMA 3000™	Non-ionic	0	N	Y (4)	0
Amin GCK 30H	SMA-IBE	Anionic	-1	Y	N	1
Amphosol 2CSF™	SMA-IBE	amphoteric	1	N	Y	0
Apo 10	SMA-IBE	Non-ionic	0	Y	N	0
Apo 12	SMA-IBE	Non-ionic	0	Y	N	0
Stepan Bio Soft D-40™	SMA-IBE	Anionic	-1	Y	Y	0
Coco Glucoside	SMA-IBE	Non-ionic	0	N C6 ring	Y	0
Fluoro octyl maltoside O310F	SMA-IBE	Non-ionic	0	N	Y	0
HEGMDE H370	SMA-IBE	Non-ionic	0	Y	N	0
HEGMDE H360	SMA-IBE	Non-ionic	0	Y	N	0
HXEGMDE H375	SMA-IBE	Non-ionic	0	Y	N	0
Lakeland PAE 136™	SMA-IBE	Anionic	-1	Y	N	0
MNA C14	SMA-IBE	Non-ionic	0	N	Y	0
n-Decanoyl-L-Methionine	SMA-IBE	Anionic	-1	N	Y	0
n-Decanoyl-L-Threonine	SMA-IBE	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Histidine	SMA-IBE	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Methionine	SMA-IBE	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Threonine	SMA-IBE	Anionic	-1	N	Y	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
n-Octanoyl-L-Methionine	SMA-IBE	Anionic	-1	N	Y	0
nDDTP D342	SMA-IBE	Non-ionic	0	N	Y	0
nDDTGP D323	SMA-IBE	Non-ionic	0	N	Y	0
nDDTP D335	SMA-IBE	Non-ionic	0	N	Y	0
OEGMIDE O330	SMA-IBE	Non-ionic	0	Y	N	0
Poly-10-laurate	SMA-IBE	Non-ionic	0	N	Y	0
Poly-10-oleate	SMA-IBE	Non-ionic	0	N	Y	1
Span 20™	SMA-IBE	Non-ionic	0	N	N	0
Stepan Mild L3™	SMA-IBE	Anionic	-1	Y	N	0
Stepan SLL FB™	SMA-IBE	Anionic	-1	N	Y	0
Stepan sulfonic acid™	SMA-IBE	Anionic	-1	Y	N	0
Triton X-305™	SMA-IBE	Non-ionic	0	N	Y	0
Triton X-45™	SMA-IBE	Non-ionic	0	N	Y	0
Tween 60™	SMA-IBE	Non-ionic	0	N	Y(4)	0
Tween 65™	SMA-IBE	Non-ionic	0	N	Y(4)	0
Tween 85™	SMA-IBE	Non-ionic	0	N	Y(4)	0
1,2-Hexanediol	SMA 725™	Anionic	-1	Y	N	0
Amin GCK 30H	SMA 725™	Anionic	-1	Y	N	1
Amphosol 2CSF™	SMA 725™	amphoteric	1	N	Y	0
Apo 10	SMA 725™	Non-ionic	0	Y	N	0
Apo 12	SMA 725™	Non-ionic	0	Y	N	0
Stepan Bio Soft D-40™	SMA 725™	Anionic	-1	Y	Y	0
Stepan Bio Terge AS-40™	SMA 725™	Anionic	-1	Y	N	1

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
Dodecane-1-sulfonic acid	SMA 725 TM	Anionic	-1	Y	N	0
PFDMP-O310F	SMA 725 TM	Non-ionic	0	N	Y	0
Genapol X080	SMA 725 TM	Non-ionic	0	Y	N	0
HEGMDE H370	SMA 725 TM	Non-ionic	0	Y	N	0
HEGMDE H360	SMA 725 TM	Non-ionic	0	Y	N	0
HXEGMDDE H375	SMA 725 TM	Non-ionic	0	Y	N	0
Lakeland PAE 136	SMA 725 TM	Anionic	-1	Y	N	0
Lathanol LAL TM	SMA 725 TM	Anionic	-1	Y	N	0
MNA C12	SMA 725 TM	Non-ionic	0	N	Y	0
MNA C14	SMA 725 TM	Non-ionic	0	N	Y	0
n-Decanoyl-L-Methionine	SMA 725 TM	Anionic	-1	N	Y	0
n-Decanoyl-L-Threonine	SMA 725 TM	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Histidine	SMA 725 TM	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Methionine	SMA 725 TM	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Threonine	SMA 725 TM	Anionic	-1	N	Y	0
n-Octanoyl-L-Methionine	SMA 725 TM	Anionic	-1	N	Y	0
nDDTP D342	SMA 725 TM	Non-ionic	0	N	Y	0
nDDTGP D323	SMA 725 TM	Non-ionic	0	N	Y	0
nDDTP D335	SMA 725 TM	Non-ionic	0	N	Y	0
Ninate 411 TM	SMA 725 TM	Anionic	-1	N	Y	0
nNDTP N335	SMA 725 TM	Non-ionic	0	N	N	0
OEGMDE O330	SMA 725 TM	Non-ionic	0	Y	N	0
Poly-10-laurate	SMA 725 TM	Non-ionic	0	N	Y	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
Poly10-oleate	SMA 725™	Non-ionic	0	N	Y	1
Poly-10-palmitate	SMA 725™	Non-ionic	0	N	Y	0
Poly-10-stearate	SMA 725™	Non-ionic	0	N	Y	0
Span 20™	SMA 725™	Non-ionic	0	N	N	0
Stepan Mild L3™	SMA 725™	Anionic	-1	Y	N	0
Stepan SLL FB™	SMA 725™	Anionic	-1	N	Y	0
Triton X-100™	SMA 725™	Non-ionic	0	N	Y	0
Triton X-305™	SMA 725™	Non-ionic	0	N	Y	0
Triton X-45™	SMA 725™	Non-ionic	0	N	Y	0
Tween 65™	SMA 725™	Non-ionic	0	N	Y(4)	0
Tween 80™	SMA 725™	Non-ionic	0	N	Y (4)	1
VDISTILL DV53™	SMA 725™	Non-ionic	0	Y	N	1, 3
Amin GCK 30H	SMA Cumene	Anionic	-1	Y	N	1
Amphosol 2CSF™	SMA Cumene	amphoteric	1	N	Y	0
Apo 10	SMA Cumene	Non-ionic	0	Y	N	0
Apo 12	SMA Cumene	Non-ionic	0	Y	N	0
Coco Glucoside	SMA Cumene	Non-ionic	0	N C6 ring	Y	0
PFDMP-O310F	SMA Cumene	Non-ionic	0	N	Y	0
Genapol X080	SMA Cumene	Non-ionic	0	Y	N	0
HEGMDE H370	SMA Cumene	Non-ionic	0	Y	N	0
HEGMDE H360	SMA Cumene	Non-ionic	0	Y	N	0
HEGMOE H350	SMA Cumene	Non-ionic	0	Y	N	0
HXEGMDDE H375	SMA Cumene	Non-ionic	0	Y	N	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
Lakeland PAE 136™	SMA Cumene	Anionic	-1	Y	N	0
MNA C12	SMA Cumene	Non-ionic	0	N	Y	0
MNA C13	SMA Cumene	Non-ionic	0	N	Y	0
MNA C14	SMA Cumene	Non-ionic	0	N	Y	0
n-Decanoyl-L-Methionine	SMA Cumene	Anionic	-1	N	Y	0
n-Decanoyl-L-Threonine	SMA Cumene	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Histidine	SMA Cumene	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Methionine	SMA Cumene	Anionic	-1	N	Y	0
n-Dodecanoyl-L-Threonine	SMA Cumene	Anionic	-1	N	Y	0
n-Octanoyl-L-Methionine	SMA Cumene	Anionic	-1	N	Y	0
Sodium dodecylbenzene sulfonate	SMA Cumene	Anionic	-1	N	Y	0
nDDTTP D342	SMA Cumene	Non-ionic	0	N	Y	0
nDDTGP D323	SMA Cumene	Non-ionic	0	N	Y	0
nDDTP D335	SMA Cumene	Non-ionic	0	N	Y	0
nNDTP N335	SMA Cumene	Non-ionic	0	N	N	0
OEGMIDE O330	SMA Cumene	Non-ionic	0	Y	N	0
Poly-10-laurate	SMA Cumene	Non-ionic	0	N	Y	0
Poly-10-oleate	SMA Cumene	Non-ionic	0	N	Y	1
Poly-10-stearate	SMA Cumene	Non-ionic	0	N	Y	0
Span 20™	SMA Cumene	Non-ionic	0	N	N	0
Stepan Mild L3™	SMA Cumene	Anionic	-1	Y	N	0
Stepan SLL FB™	SMA Cumene	Anionic	-1	N	Y	0
Stepan sulfonic acid™	SMA Cumene	Anionic	-1	Y	N	0

Table 12C. Positively Scoring Nanonets						
Surfactant	Polymer	Surfactant class	Score	Linear head group	Branched head group	DB
TC Lab 35	SMA Cumene	Amphoteric	1	N	Y	0
Triton X-100™	SMA Cumene	Non-ionic	0	N	Y	0
Triton X-305™	SMA Cumene	Non-ionic	0	N	Y	0
Triton X-45™	SMA Cumene	Non-ionic	0	N	Y	0
Tween 40™	SMA Cumene	Non-ionic	0	N	Y(4)	0
Tween 65™	SMA Cumene	Non-ionic	0	N	Y(4)	0
Tween 80™	SMA Cumene	Non-ionic	0	N	Y (4)	1
Tween 85™	SMA Cumene	Non-ionic	0	N	Y (4)	0

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
1,2-Hexanediol	DIBMA	1,2-glycol	NA	NA	109.7	0.232
Amphosol 2CSF™	DIBMA	Dipropionate	NA	NA	7.808	0.315
Apo 10	DIBMA	Phosphine oxide	NA	NA	5.835	0.259
Apo 12	DIBMA	Phosphine oxide	NA	NA	82.41	0.272
Stapan Bio Soft D-40™	DIBMA	Sulfonate	NA	NA	5.13	0.322
Cedepal TD-403 MFLD™	DIBMA	Ether sulfate	NA	NA	6.421	0.235
Coco Glucoside	DIBMA	Gglucopyranoside	13.1	NA	27.32	0.154
Dodecane-1-sulfonic acid	DIBMA	Sulfonate	NA	NA	5.108	0.227
PFDMP O310F	DIBMA	Maltoside-sugar	NA	NA	23.83	0.172
Genapol X080	DIBMA	Poly ethylene glycol	12	0.06-0.15	10.58	0.147
HEGMDE H370	DIBMA	Ethylene glycol	NA	0.069	8.218	0.14
HEGMDE H360	DIBMA	Ethylene glycol	NA	0.9	6.033	0.158

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
HEGMOEH350	DIBMA	Ethylene glycol	NA	10	5.505	0.25
HXEGMDDE H375	DIBMA	Ethylene glycol	NA	NA	9.351	0.079
Lakeland PAE 136™	DIBMA	Phosphate ester	NA	NA	7.924	0.161
Lathanol LAL™	DIBMA	Sulfoacetate	NA	NA	6.814	0.151
MNA C12	DIBMA	Glucopyranoside	NA	0.004	6.122	0.068
MNA C13	DIBMA	Glucopyranoside	NA	0.002	6.423	0.046
MNA C14	DIBMA	Mannitol	NA	0.001	7.548	0.217
n-Decanoyl-L-Methionine	DIBMA	Amino acid	NA	NA	85.47	0.172
n-Decanoyl-L-Threonine	DIBMA	Amino acid	NA	NA	90.68	0.213
n-Dodecanoyl-L-Histidine	DIBMA	Amino acid	NA	NA	99.96	0.249
n-Dodecanoyl-L-Methionine	DIBMA	Amino acid	NA	NA	87.09	0.138
n-Dodecanoyl-L-Threonine	DIBMA	Amino acid	NA	NA	99.77	0.203
n-Octanoyl-L-Methionine	DIBMA	Amino acid	NA	NA	116.6	0.239
Sodium dodecylbenzene sulfonate	DIBMA	Sulfonate	NA	NA	4.235	0.53
nDDTTP D342	DIBMA	Thiomaltopyranoside	NA	0.05	11.67	0.131
nDDTGP D323	DIBMA	Thioglucopyranoside	NA	0.9	262.2	0.176
nDDTP D335	DIBMA	Thiomaltopyranoside	NA	0.9	6.881	0.078
Ninate 411™	DIBMA	Sulfonate	NA	1,214 mg/l	216.2	0.288
nNDTP N335	DIBMA	Thioglucopyranoside	NA	2.9	137	0.357
OEGMIDE O330	DIBMA	Ethylene glycol	NA	0.09	8.099	0.165
Poly-10- laurate	DIBMA	Polyglycerol fatty acid ester	12	NA	114.8	0.226
Poly-10- oleate	DIBMA	Polyglycerol fatty acid ester	10.5	NA	358	0.254
Poly-10-stearate	DIBMA	Polyglycerol fatty acid ester	10.5	NA	112.3	0.172

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
Span 20™	DIBMA	Sorbitan	NA	NA	1061	0.472
Stepan Mild L3™	DIBMA	Lactylate	NA	NA	372.9	0.067
Stepan SLL FB™	DIBMA	Lactylate	NA	NA	136.3	0.144
Stepan sulfonic acid™	DIBMA	Sulfonate	NA	NA	2.821	0.186
TC Lab 35	DIBMA	Acetate	13	NA	23.27	0.328
Triton X-100™	DIBMA	Ethylene glycol	13.4	0.24	10.07	0.089
Triton X-305™	DIBMA	Ethylene glycol	17.3	0.65	10.66	0.181
Triton X-45™	DIBMA	Ethylene glycol	9.8	0.1	156.3	0.325
Tween 20™	DIBMA	Polysorbate	16.7	0.05	9.173	0.072
Tween 40™	DIBMA	Polysorbate	15.6	NA	9.691	0.079
Tween 60™	DIBMA	Polysorbate	14.9	0.022	134.2	0.637
tween 65™	DIBMA	Polysorbate	15.9	NA	114.5	0.286
Tween 80™	DIBMA	Polysorbate	15	0.012	10.42	0.026
Tween 85™	DIBMA	Polysorbate	NA	NA	189.8	0.252
1-Octane sulfonate	SMA 2000™	Sulfonate	NA	NA	1.562	0.367
Amin GCK 30H	SMA 2000™	Acyl glycinate	NA	NA	7.286	0.3
Amphosol 2CSF™	SMA 2000™	Dipropionate	NA	NA	7.808	0.315
Apo 10	SMA 2000™	Phosphine oxide	NA	NA	5.835	0.259
Apo 12	SMA 2000™	Phosphine oxide	NA	NA	82.41	0.272
Stepan Bio Soft D-40™	SMA 2000™	Sulfonate	NA	NA	5.13	0.322
Stepan Bio Terge AS-40™	SMA 2000™	Sulfonate	NA	NA	6.434	0.181
Coco Glucoside	SMA 2000™	Glucopyranoside	13.1	NA	27.32	0.154
Dodecane-1-sulfonic acid	SMA 2000™	Sulfonate	NA	NA	5.108	0.227

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
PFDMP-O310F	SMA 2000™	Maltoside-sugar	NA	NA	23.83	0.172
Genapol X080	SMA 2000™	Poly ethylene glycol	12	0.06-0.15	10.58	0.147
HEGMDE H370	SMA 2000™	Ethylene glycol	NA	0.069	8.218	0.14
HEGMDE H360	SMA 2000™	Ethylene glycol	NA	0.9	6.033	0.158
HEGMOEH350	SMA 2000™	Ethylene glycol	NA	10	5.505	0.25
HXEGMDDE H375	SMA 2000™	Ethylene glycol	NA	NA	9.351	0.079
Lakeland PAE 136™	SMA 2000™	Phosphate ester	NA	NA	7.924	0.161
Stepan Lathanol LAL™	SMA 2000™	Sulfoacetate	NA	NA	6.814	0.151
MNA C13	SMA 2000™	Glucopyranoside	NA	0.002	6.423	0.046
MNA C14	SMA 2000™	Mannitol	NA	0.001	7.548	0.217
n-Decanoyl-L-Methionine	SMA 2000™	Amino acid	NA	NA	85.47	0.172
n-Decanoyl-L-Threonine	SMA 2000™	Amino acid	NA	NA	90.68	0.213
n-Dodecanoyl-L-Histidine	SMA 2000™	Amino acid	NA	NA	99.96	0.249
n-Dodecanoyl-L-Methionine	SMA 2000™	Amino acid	NA	NA	87.09	0.138
n-Dodecanoyl-L-Threonine	SMA 2000™	Amino acid	NA	NA	99.77	0.203
n-Octanoyl-L-Methionine	SMA 2000™	Amino acid	NA	NA	116.6	0.239
Sodium dodecylbenzene sulfonate	SMA 2000™	Sulfonate	NA	NA	4.235	0.53
nDDTP D342	SMA 2000™	Thiomaltopyranoside	NA	0.05	11.67	0.131
nDDTGP D323	SMA 2000™	Thioglucopyranoside	NA	0.9	262.2	0.176
nDDTP D335	SMA 2000™	Thiomaltopyranoside	NA	0.9	6.881	0.078
Ninate 411™	SMA 2000™	Sulfonate	NA	1,214 mg/l	216.2	0.288
nNDTP N335	SMA 2000™	Thioglucopyranoside	NA	2.9	137	0.357
OEGMDE O330	SMA 2000™	Ethylene glycol	NA	0.09	8.099	0.165

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
Poly-10-laurate	SMA 2000™	Polyglycerol fatty acid ester	12	NA	114.8	0.226
Poly-10-oleate	SMA 2000™	Polyglycerol fatty acid ester	10.5	NA	358	0.254
Poly-10-stearate	SMA 2000™	Polyglycerol fatty acid ester	10.5	NA	112.3	0.172
Span 20™	SMA 2000™	Sorbitan	NA	NA	1061	0.472
Stepan Mild L3™	SMA 2000™	Lactylate	NA	NA	372.9	0.067
Stepan SLL FB™	SMA 2000™	Lactylate	NA	NA	136.3	0.144
Stepan sulfonic acid™	SMA 2000™	Sulfonate	NA	NA	2.821	0.186
TC Lab 35	SMA 2000™	Acetate	13	NA	23.27	0.328
Triton X-100™	SMA 2000™	Ethylene glycol	13.4	0.24	10.07	0.089
Triton X-305™	SMA 2000™	Ethylene glycol	17.3	0.65	10.66	0.181
Triton X-45™	SMA 2000™	Ethylene glycol	9.8	0.1	156.3	0.325
Tween 20™	SMA 2000™	Polysorbate	16.7	0.05	9.173	0.072
Tween 40™	SMA 2000™	Polysorbate	15.6	NA	9.691	0.079
Tween 60™	SMA 2000™	Polysorbate	14.9	0.022	134.2	0.637
Tween 65™	SMA 2000™	Polysorbate	10.5	NA	114.5	0.286
Tween 80™	SMA 2000™	Polysorbate	15	0.012	10.42	0.026
Tween 85™	SMA 2000™	Polysorbate	11	NA	189.8	0.252
VDISTILL DV53™	SMA 2000™	Fatty acid	NA	NA	NA	NA
VDISTILL DV63™	SMA 2000™	Fatty acid	NA	NA	NA	NA
1 octane sulfonate	SMA 2021™	Sulfonate	NA	NA	1.562	0.367
Amin GCK 30H	SMA 2021™	Acyl glycinate	NA	NA	7.286	0.3
Amphosol 2CSF	SMA 2021™	Dipropionate	NA	NA	7.808	0.315
Anthraquinone-2-sulfonic acid	SMA 2021™	Sulfonate	NA	NA	1.008	0.343

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
Apo 10	SMA 2021™	Phosphine oxide	NA	NA	5.835	0.259
Apo 12	SMA 2021™	Phosphine oxide	NA	NA	82.41	0.272
Cedepal TD-403 MFLD™	SMA 2021™	Ether sulfate	NA	NA	6.421	0.235
Coco Glucoside	SMA 2021™	Glucopyranoside	13.1	NA	27.32	0.154
Dodecane-1-sulfonic acid	SMA 2021™	Sulfonate	NA	NA	5.108	0.227
PFDMP-O310F	SMA 2021™	Maltoside-sugar	NA	NA	23.83	0.172
Genapol X080	SMA 2021™	Poly ethylene glycol	12	0.06-0.15	10.58	0.147
HEGMDE H370	SMA 2021™	Ethylene glycol	NA	0.069	8.218	0.14
HEGMDE H360	SMA 2021™	Ethylene glycol	NA	0.9	6.033	0.158
HEGMOE H350	SMA 2021™	Ethylene glycol	NA	10	5.505	0.25
HXEGMDDE H375	SMA 2021™	Ethylene glycol	NA	NA	9.351	0.079
Lakeland PAE 136™	SMA 2021™	Phosphate ester	NA	NA	7.924	0.161
Lathanol LAL™	SMA 2021™	Sulfoacetate	NA	NA	6.814	0.151
MNA C13	SMA 2021™	Glucopyranoside	NA	0.002	6.423	0.046
MNA C14	SMA 2021™	Mannitol	NA	0.001	7.548	0.217
n-Decanoyl-L-Methionine	SMA 2021™	Amino acid	NA	NA	85.47	0.172
n-Decanoyl-L-Threonine	SMA 2021™	Amino acid	NA	NA	90.68	0.213
n-Dodecanoyl-L-Methionine	SMA 2021™	Amino acid	NA	NA	87.09	0.138
n-Dodecanoyl-L-Threonine	SMA 2021™	Amino acid	NA	NA	99.77	0.203
n-Octanoyl-L-Methionine	SMA 2021™	Amino acid	NA	NA	116.6	0.239
Sodium dodecylbenzene sulfonate	SMA 2021™	Sulfonate	NA	NA	4.235	0.53
nDDDTp D342	SMA 2021™	Thiomaltopyranoside	NA	0.05	11.67	0.131
nDDTGP D323	SMA 2021™	Thioglucopyranoside	NA	0.9	262.2	0.176

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
nDDTP D335	SMA 2021™	Thiomaltopyranoside	NA	0.9	6.881	0.078
Ninate 411™	SMA 2021™	Sulfonate	NA	1,214 mg/l	216.2	0.288
nNDTP N335	SMA 2021™	Thioglucopyranoside	NA	2.9	137	0.357
OEGMDE O330	SMA 2021™	Ethylene glycol	NA	0.09	8.099	0.165
Poly-10-laurate	SMA 2021™	Polyglycerol fatty acid ester	12	NA	114.8	0.226
Poly-10-oleate	SMA 2021™	Polyglycerol fatty acid ester	10.5	NA	358	0.254
Poly-10-palmitate	SMA 2021™	Polyglycerol fatty acid ester	10.5	NA	111.8	0.226
Poly-10-stearate	SMA 2021™	Polyglycerol fatty acid ester	NA	NA	112.3	0.172
Span 20™	SMA 2021™	Sorbitan	NA	NA	1061	0.472
Stepan Mild L3™	SMA 2021™	Lactylate	NA	NA	372.9	0.067
Stepan SLL FB™	SMA 2021™	Lactylate	NA	NA	136.3	0.144
Stepan sulfonic acid™	SMA 2021™	Sulfonate	NA	NA	2.821	0.186
TC Lab 35	SMA 2021™	Acetate	13	NA	23.27	0.328
Triton X-100™	SMA 2021™	Ethylene glycol	13.4	0.24	10.07	0.089
Triton X-305™	SMA 2021™	Ethylene glycol	17.3	0.65	10.66	0.181
Triton X-45™	SMA 2021™	Ethylene glycol	9.8	0.1	156.3	0.325
Tween 40™	SMA 2021™	Polysorbate	15.6	NA	9.691	0.079
Tween 60™	SMA 2021™	Polysorbate	14.9	0.022	134.2	0.637
Tween 80™	SMA 2021™	Polysorbate	15	0.012	10.42	0.026
Tween 85™	SMA 2021™	Polysorbate	11	NA	189.8	0.252
Amphosol 2CSF™	SMA 3000™	Dipropionate	NA	NA	7.808	0.315
Anthraquinone-2-sulfonic acid	SMA 3000™	Sulfonate	NA	NA	1.008	0.343
Apo 10	SMA 3000™	Phosphine oxide	NA	NA	5.835	0.259

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
Apo 12	SMA 3000™	Phosphine oxide	NA	NA	82.41	0.272
Coco Glucoside	SMA 3000™	Glucopyranoside	13.1	NA	27.32	0.154
PFDMP-O310F	SMA 3000™	Maltoside-sugar	NA	NA	23.83	0.172
Genapol X080	SMA 3000™	Poly ethylene glycol	12	0.06-0.15	10.58	0.147
HEGMDE H370	SMA 3000™	Ethylene glycol	NA	0.069	8.218	0.14
HEGMDE H360	SMA 3000™	Ethylene glycol	NA	0.9	6.033	0.158
HEGMOE H350	SMA 3000™	Ethylene glycol	NA	10	5.505	0.25
HXEGMDDE H375	SMA 3000™	Ethylene glycol	NA	NA	9.351	0.079
Lakeland PAE 136™	SMA 3000™	Phosphate ester	NA	NA	7.924	0.161
Lathanol LAL™	SMA 3000™	Sulfoacetate	NA	NA	6.814	0.151
MNA C12	SMA 3000™	Glucopyranoside	NA	0.004	6.122	0.068
MNA C13	SMA 3000™	Glucopyranoside	NA	0.002	6.423	0.046
MNA C14	SMA 3000™	Mannitol	NA	0.001	7.548	0.217
n-Decanoyl-L-Methionine	SMA 3000™	Amino acid	NA	NA	85.47	0.172
n-Decanoyl-L-Threonine	SMA 3000™	Amino acid	NA	NA	90.68	0.213
n-Dodecanoyl-L-Histidine	SMA 3000™	Amino acid	NA	NA	99.96	0.249
n-Dodecanoyl-L-Methionine	SMA 3000™	Amino acid	NA	NA	87.09	0.138
n-Dodecanoyl-L-Threonine	SMA 3000™	Amino acid	NA	NA	99.77	0.203
n-Octanoyl-L-Methionine	SMA 3000™	Amino acid	NA	NA	116.6	0.239
Sodium dodecylbenzene sulfonate	SMA 3000™	Sulfonate	NA	NA	4.235	0.53
nDDDT D342	SMA 3000™	Thiomaltopyranoside	NA	0.05	11.67	0.131
nDDTGP D323	SMA 3000™	Thioglucopyranoside	NA	0.9	262.2	0.176
nDDTP D335	SMA 3000™	Thiomaltopyranoside	NA	0.9	6.881	0.078

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
Ninate 411™	SMA 3000™	Sulfonate	NA	1,214 mg/l	216.2	0.288
nNDTP N335	SMA 3000™	Thioglucopyranoside	NA	2.9	137	0.357
nODTP O320	SMA 3000™	Thiomaltopyranoside	NA	8.5	6.022	0.324
OEGMDE O330	SMA 3000™	ethylene glycol	NA	0.09	8.099	0.165
Poly-10-laurate	SMA 3000™	polyglycerol fatty acid ester	12	NA	114.8	0.226
Poly-10-oleate	SMA 3000™	polyglycerol fatty acid ester	10.5	NA	358	0.254
Poly-10-palmitate	SMA 3000™	polyglycerol fatty acid ester	10.5	NA	111.8	0.226
Poly-10-stearate	SMA 3000™	polyglycerol fatty acid ester	11	NA	112.3	0.172
Span 20™	SMA 3000™	Sorbitan	NA	NA	1061	0.472
Stepan Mild L3™	SMA 3000™	Lactylate	NA	NA	372.9	0.067
Stepan SLL FB™	SMA 3000™	Lactylate	NA	NA	136.3	0.144
Stepan sulfonic acid™	SMA 3000™	Sulfonate	NA	NA	2.821	0.186
TC Lab 35	SMA 3000™	Acetate	13	NA	23.27	0.328
Triton X-100™	SMA 3000™	Ethylene glycol	13.4	0.24	10.07	0.089
Triton X-305™	SMA 3000™	Ethylene glycol	17.3	0.65	10.66	0.181
Triton X-45™	SMA 3000™	Ethylene glycol	9.8	0.1	156.3	0.325
Tween 65™	SMA 3000™	Polysorbate	10.5	NA	114.5	0.286
Tween 85™	SMA 3000™	Polysorbate	11	NA	189.8	0.252
Amin GCK 30H	SMA-IBE	Acyl glycinat	NA	NA	7.286	0.3
Amphosol 2CSF™	SMA-IBE	Dipropionate	NA	NA	7.808	0.315
Apo 10	SMA-IBE	Phosphine oxide	NA	NA	5.835	0.259
Apo 12	SMA-IBE	Phosphine oxide	NA	NA	82.41	0.272
Stepan Bio Soft D-40™	SMA-IBE	Sulfonate	NA	NA	5.13	0.322

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
Coco Glucoside	SMA-IBE	Glucopyranoside	13.1	NA	27.32	0.154
Fluoro octyl maltoside O310F	SMA-IBE	Maltoside-sugar	NA	NA	23.83	0.172
HEGMDE H370	SMA-IBE	Ethylene glycol	NA	0.069	8.218	0.14
HEGMDE H360	SMA-IBE	Ethylene glycol	NA	0.9	6.033	0.158
HXEGMDE H375	SMA-IBE	Ethylene glycol	NA	NA	9.351	0.079
Lakeland PAE 136™	SMA-IBE	Phosphate ester	NA	NA	7.924	0.161
MNA C14	SMA-IBE	Mannitol	NA	0.001	7.548	0.217
n-Decanoyl-L-Methionine	SMA-IBE	Amino acid	NA	NA	85.47	0.172
n-Decanoyl-L-Threonine	SMA-IBE	Amino acid	NA	NA	90.68	0.213
n-Dodecanoyl-L-Histidine	SMA-IBE	Amino acid	NA	NA	99.96	0.249
n-Dodecanoyl-L-Methionine	SMA-IBE	Amino acid	NA	NA	87.09	0.138
n-Dodecanoyl-L-Threonine	SMA-IBE	Amino acid	NA	NA	99.77	0.203
n-Octanoyl-L-Methionine	SMA-IBE	Amino acid	NA	NA	116.6	0.239
nDDTP D342	SMA-IBE	Thiomaltopyranoside	NA	0.05	11.67	0.131
nDDTGP D323	SMA-IBE	Thioglucofuranoside	NA	0.9	262.2	0.176
nDDTP D335	SMA-IBE	Thiomaltopyranoside	NA	0.9	6.881	0.078
OEGMDE O330	SMA-IBE	Ethylene glycol	NA	0.09	8.099	0.165
Poly-10-laurate	SMA-IBE	Polyglycerol fatty acid ester	12	NA	114.8	0.226
Poly-10-oleate	SMA-IBE	Polyglycerol fatty acid ester	10.5	NA	358	0.254
Span 20™	SMA-IBE	Sorbitan	NA	NA	1061	0.472
Stepan Mild L3™	SMA-IBE	Lactylate	NA	NA	372.9	0.067
Stepan SLL FB™	SMA-IBE	Lactylate	NA	NA	136.3	0.144
Stepan sulfonic acid™	SMA-IBE	Sulfonate	NA	NA	2.821	0.186

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
Triton X-305™	SMA-IBE	Ethylene glycol	17.3	0.65	10.66	0.181
Triton X-45™	SMA-IBE	Ethylene glycol	9.8	0.1	156.3	0.325
Tween 60™	SMA-IBE	Polysorbate	14.9	0.022	134.2	0.637
Tween 65™	SMA-IBE	Polysorbate	10.5	NA	114.5	0.286
Tween 85™	SMA-IBE	Polysorbate	11	NA	189.8	0.252
1,2-Hexanediol	SMA 725™	1,2-glycol	NA	NA	109.7	0.232
Amin GCK 30H	SMA 725™	Acyl glycinate	NA	NA	7.286	0.3
Amphosol 2CSF™	SMA 725™	Dipropionate	NA	NA	7.808	0.315
Apo 10	SMA 725™	Phosphine oxide	NA	NA	5.835	0.259
Apo 12	SMA 725™	Phosphine oxide	NA	NA	82.41	0.272
Stepan Bio Soft D-40™	SMA 725™	Sulfonate	NA	NA	5.13	0.322
Stepan Bio Terge AS-40™	SMA 725™	Sulfonate	NA	NA	6.434	0.181
Dodecane-1-sulfonic acid	SMA 725™	Sulfonate	NA	NA	5.108	0.227
PFDMP-O310F	SMA 725™	Maltoside-sugar	NA	NA	23.83	0.172
Genapol X080	SMA 725™	Poly ethylene glycol	12	0.06-0.15	10.58	0.147
HEGMDE H370	SMA 725™	Ethylene glycol	NA	0.069	8.218	0.14
HEGMDE H360	SMA 725™	Ethylene glycol	NA	0.9	6.033	0.158
HXEGMDDE H375	SMA 725™	Ethylene glycol	NA	NA	9.351	0.079
Lakeland PAE 136	SMA 725™	Phosphate ester	NA	NA	7.924	0.161
Stepan Lathanol LAL™	SMA 725™	Sulfoacetate	NA	NA	6.814	0.151
MNA C12	SMA 725™	Glucopyranoside	NA	0.004	6.122	0.068
MNA C14	SMA 725™	Mannitol	NA	0.001	7.548	0.217
n-Decanoyl-L-Methionine	SMA 725™	Amino acid	NA	NA	85.47	0.172

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
n-Decanoyl-L-Threonine	SMA 725™	Amino acid	NA	NA	90.68	0.213
n-Dodecanoyl-L-Histidine	SMA 725™	Amino acid	NA	NA	99.96	0.249
n-Dodecanoyl-L-Methionine	SMA 725™	Amino acid	NA	NA	87.09	0.138
n-Dodecanoyl-L-Threonine	SMA 725™	Amino acid	NA	NA	99.77	0.203
n-Octanoyl-L-Methionine	SMA 725™	Amino acid	NA	NA	116.6	0.239
nDDTP D342	SMA 725™	Thiomaltopyranoside	NA	0.05	11.67	0.131
nDDTGP D323	SMA 725™	Thioglucopyranoside	NA	0.9	262.2	0.176
nDDTP D335	SMA 725™	Thiomaltopyranoside	NA	0.9	6.881	0.078
Ninate 411™	SMA 725™	Sulfonate	NA	1,214 mg/l	216.2	0.288
nNDTP N335	SMA 725™	Thioglucopyranoside	NA	2.9	137	0.357
OEGMDE O330	SMA 725™	Ethylene glycol	NA	0.09	8.099	0.165
Poly-10-laurate	SMA 725™	Polyglycerol fatty acid ester	12	NA	114.8	0.226
Poly-10-oleate	SMA 725™	Polyglycerol fatty acid ester	10.5	NA	358	0.254
Poly-10-palmitate	SMA 725™	Polyglycerol fatty acid ester	10.5	NA	111.8	0.226
Poly-10-stearate	SMA 725™	Polyglycerol fatty acid ester	11	NA	112.3	0.172
Span 20™	SMA 725™	Sorbitan	8.6	NA	1061	0.472
Stepan Mild L3™	SMA 725™	Lactylate	NA	NA	372.9	0.067
Stepan SLL FB™	SMA 725™	Lactylate	NA	NA	136.3	0.144
Triton X-100™	SMA 725™	Ethylene glycol	13.4	0.24	10.07	0.089
Triton X-305™	SMA 725™	Ethylene glycol	17.3	0.65	10.66	0.181
Triton X-45™	SMA 725™	Ethylene glycol	9.8	0.1	156.3	0.325
Tween 65™	SMA 725™	Polysorbate	10	NA	114.5	0.286
Tween 80™	SMA 725™	Polysorbate	15	0.012	10.42	0.026

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
VDISTILL DV53™	SMA 725™	Fatty acid	NA	NA	NA	NA
Amin GCK 30H	SMA Cumene	Acyl glycinate	NA	NA	7.286	0.3
Amphosol 2CSF™	SMA Cumene	Dipropionate	NA	NA	7.808	0.315
Apo 10	SMA Cumene	Phosphine oxide	NA	NA	5.835	0.259
Apo 12	SMA Cumene	Phosphine oxide	NA	NA	82.41	0.272
Coco Glucoside	SMA Cumene	Glucopyranoside	13.1	NA	27.32	0.154
PFDMP-O310F	SMA Cumene	Maltoside-sugar	NA	NA	23.83	0.172
Genapol X080	SMA Cumene	Poly ethylene glycol	12	0.06-0.15	10.58	0.147
HEGMDDE H370	SMA Cumene	Ethylene glycol	NA	0.069	8.218	0.14
HEGMDE H360	SMA Cumene	Ethylene glycol	NA	0.9	6.033	0.158
HEGMOE H350	SMA Cumene	Ethylene glycol	NA	10	5.505	0.25
HXEGMDDE H375	SMA Cumene	Ethylene glycol	NA	NA	9.351	0.079
Lakeland PAE 136™	SMA Cumene	Phosphate ester	NA	NA	7.924	0.161
MNA C12	SMA Cumene	Glucopyranoside	NA	0.004	6.122	0.068
MNA C13	SMA Cumene	Glucopyranoside	NA	0.002	6.423	0.046
MNA C14	SMA Cumene	Mannitol	NA	0.001	7.548	0.217
n-Decanoyl-L-Methionine	SMA Cumene	Amino acid	NA	NA	85.47	0.172
n-Decanoyl-L-Threonine	SMA Cumene	Amino acid	NA	NA	90.68	0.213
n-Dodecanoyl-L-Histidine	SMA Cumene	Amino acid	NA	NA	99.96	0.249
n-Dodecanoyl-L-Methionine	SMA Cumene	Amino acid	NA	NA	87.09	0.138
n-Dodecanoyl-L-Threonine	SMA Cumene	Amino acid	NA	NA	99.77	0.203
n-Octanoyl-L-Methionine	SMA Cumene	Amino acid	NA	NA	116.6	0.239
Sodium dodecylbenzene sulfonate	SMA Cumene	Sulfonate	NA	NA	4.235	0.53

Table 12D. Positively Scoring Nanonets						
Surfactant	Polymer	Head group	HLB	CMC [mM]	Micelle size	PDI
nDDTP D342	SMA Cumene	Thiomaltopyranoside	NA	0.05	11.67	0.131
nDDTGP D323	SMA Cumene	Thioglucopyranoside	NA	0.9	262.2	0.176
nDDTP D335	SMA Cumene	Thiomaltopyranoside	NA	0.9	6.881	0.078
nNDTP N335	SMA Cumene	Thioglucopyranoside	NA	2.9	137	0.357
OEGMIDE O330	SMA Cumene	Ethylene glycol	NA	0.09	8.099	0.165
Poly-10-laurate	SMA Cumene	Polyglycerol fatty acid ester	12	NA	114.8	0.226
Poly-10-oleate	SMA Cumene	polyglycerol fatty acid ester	10.5	NA	358	0.254
Poly-10-stearate	SMA Cumene	polyglycerol fatty acid ester	11	NA	112.3	0.172
Span 20™	SMA Cumene	Sorbitan	NA	NA	1061	0.472
Stepan Mild L3™	SMA Cumene	Lactylate	NA	NA	372.9	0.067
Stepan SLL FB™	SMA Cumene	Lactylate	NA	NA	136.3	0.144
Stepan sulfonic acid™	SMA Cumene	Sulfonate	NA	NA	2.821	0.186
TC Lab 35	SMA Cumene	Acetate	13	NA	23.27	0.328
Triton X-100™	SMA Cumene	Ethylene glycol	13.4	0.24	10.07	0.089
Triton X-305™	SMA Cumene	Ethylene glycol	17.3	0.65	10.66	0.181
Triton X-45™	SMA Cumene	Ethylene glycol	9.8	0.1	156.3	0.325
Tween 40™	SMA Cumene	Polysorbate	15.6	NA	9.691	0.079
Tween 65™	SMA Cumene	Polysorbate	15.9	NA	114.5	0.286
Tween 80™	SMA Cumene	Polysorbate	10.5	0.012	10.42	0.026
Tween 85™	SMA Cumene	Polysorbate	11	NA	189.8	0.252

Table 13. Non-ionic Surfactants and Polymer Combinations That Did Not Form a Nanonet Response in Titration Assays (No Sigmoid Detected)							
Surfactant	Alkyl Chain Carbon no.	Polymer	Average Polymer Diameter [nm]	Theoretical Polymer Diameter	Head Group	Surfactant Aggregate Diameter [nm]	Poly dispersity index (PDI)
Coco Glucoside	18	SMA 725™	15.03	6.8	Glucopyranoside	27.32	0.154
HEGMOE H350	8	SMA 725™	15.03	6.8	Ethylene glycol	5.505	0.25
MNA C13	13	SMA 725™	15.03	6.8	Glucopyranoside	6.423	0.046
nHDTP H301	7	SMA 725™	15.03	6.8	Thioglucopyranoside	8.935	0.011
nODTTP O320	8	SMA 725™	15.03	6.8	Thiomaltopyranoside	6.022	0.324
Tween 20™	12	SMA 725™	15.03	6.8	Polysorbate	9.173	0.072
Tween 40™	16	SMA 725™	15.03	6.8	Polysorbate	9.691	0.079
Tween 60™	18	SMA 725™	15.03	6.8	Polysorbate	134.2	0.637
Tween 85™	16	SMA 725™	15.03	6.8	Polysorbate	189.8	0.252
Vdistill DV63™	8, 18	SMA 725™	15.03	6.8	Fatty acid	NA	NA
MNA C12	12	SMA 2000™	5.242	2.6	Glucopyranoside	6.122	0.068
nHDTP H301	7	SMA 2000™	5.242	2.6	Thioglucopyranoside	8.935	0.011
nODTTP O320	8	SMA 2000™	5.242	2.6	Thiomaltopyranoside	6.022	0.324
Poly-10-palmitate	16	SMA 2000™	5.242	2.6	Polyglycerol fatty acid ester	111.8	0.226
nHDTP H301	7	DIBMA	92.7	3.2	Thioglucopyranoside	8.935	0.011
nODTTP O320	8	DIBMA	92.7	3.2	Thiomaltopyranoside	6.022	0.324
Poly-10-palmitate	16	DIBMA	92.7	3.2	Polyglycerol fatty acid ester	111.8	0.226
Vdistill DV53™	18,18	DIBMA	92.7	3.2	Fatty acid	NA	NA
Vdistill DV63™	18,18	DIBMA	92.7	3.2	Fatty acid	NA	NA
MNA C12	12	SMA 2021™	6.627	3.7	Glucopyranoside	6.122	0.068
nHDTP H301	7	SMA 2021™	6.627	3.7	Thioglucopyranoside	8.935	0.011

Table 13. Non-ionic Surfactants and Polymer Combinations That Did Not Form a Nanonet Response in Titration Assays (No Sigmoid Detected)							
Surfactant	Alkyl Chain Carbon no.	Polymer	Average Polymer Diameter [nm]	Theoretical Polymer Diameter	Head Group	Surfactant Aggregate Diameter [nm]	Poly dispersity index (PDI)
nODTTP O320	8	SMA 2021™	6.627	3.7	Thiomaltopyranoside	6.022	0.324
Tween 20™	12	SMA 2021™	6.627	3.7	Polysorbate	9.173	0.072
Tween 65™	18	SMA 2021™	6.627	3.7	Polysorbate	114.5	0.286
Vdistill DV53™	18, 18	SMA 2021™	6.627	3.7	Fatty acid	NA	NA
Vdistill DV63™	18,18	SMA 2021™	6.627	3.7	Fatty acid	NA	NA
nHDTP H301	7	SMA 3000™	3.713	2.8	Thioglucopyranoside	8.935	0.011
Tween 20™	12	SMA 3000™	3.713	2.8	Polysorbate	9.173	0.072
Tween 40™	16	SMA 3000™	3.713	2.8	Polysorbate	9.691	0.079
Tween 60™	18	SMA 3000™	3.713	2.8	Polysorbate	134.2	0.637
Tween 80™	18	SMA 3000™	3.713	2.8	Polysorbate	10.42	0.026
Vdistill DV53™	18,18	SMA 3000™	3.713	2.8	Fatty acid	NA	NA
Vdistill DV63™	18,18	SMA 3000™	3.713	2.8	Fatty acid	NA	NA
Genapol X-080	13	SMA IBE	9.759	5.4	Poly ethylene glycol	10.58	0.147
hegmoe H350	8	SMA IBE	9.759	5.4	Ethylene glycol	5.505	0.25
MNA C12	12	SMA IBE	9.759	5.4	Glucopyranoside	6.122	0.068
MNA C13	13	SMA IBE	9.759	5.4	Glucopyranoside	6.423	0.046
nHDTP H301	7	SMA IBE	9.759	5.4	Thioglucopyranoside	8.935	0.011
nNDTP N335	9	SMA IBE	9.759	5.4	Thioglucopyranoside	137	0.357
nODTTP O320	8	SMA IBE	9.759	5.4	Thiomaltopyranoside	6.022	0.324
Poly-10- palmitate	16	SMA IBE	9.759	5.4	Polyglycerol fatty acid ester	111.8	0.226
Poly-10-stearate	18	SMA IBE	9.759	5.4	Polyglycerol fatty acid ester	112.3	0.172

Surfactant	Alkyl Chain Carbon no.	Polymer	Average Polymer Diameter [nm]	Theoretical Polymer Diameter	Head Group	Surfactant Aggregate Diameter [nm]	Poly dispersity index (PDI)
Triton X-100™	8	SMA IBE	9.759	5.4	Ethylene glycol	10.07	0.089
Tween 20™	12	SMA IBE	9.759	5.4	Polysorbate	9.173	0.072
Tween 40™	16	SMA IBE	9.759	5.4	Polysorbate	9.691	0.079
Tween 80™	18	SMA IBE	9.759	5.4	Polysorbate	10.42	0.026
Vdistill DV53™	18,18	SMA IBE	9.759	5.4	Fatty acid	NA	NA
Vdistill DV63™	18,18	SMA IBE	9.759	5.4	Fatty acid	NA	NA
nHDTP H301	7	SMA Cumene	7.123	1.5	Thioglucopyranoside	8.935	0.011
nODTTP O320	8	SMA Cumene	7.123	1.5	Thiomaltopyranoside	6.022	0.324
Poly-10- palmitate	16	SMA Cumene	7.123	1.5	Polyglycerol fatty acid ester	111.8	0.226
Tween 20™	12	SMA Cumene	7.123	1.5	Polysorbate	9.173	0.072
Tween 60™	18	SMA Cumene	7.123	1.5	Polysorbate	134.2	0.637
Vdistill DV53™	18,18	SMA Cumene	7.123	1.5	Fatty acid	NA	NA
Vdistill DV63™	18,18	SMA Cumene	7.123	1.5	Fatty acid	NA	NA

Surfactant	Alkyl Chain Carbon No.	Polymer	Average Polymer Diameter [nm]	Theoretical Polymer	Head Group	Surfactant Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
1-Octane sulfonate	8	SMA 725™	15.03	6.8	Sulfonate	1.562	0.367

Table 14. Anionic Surfactants and Polymer Combinations That Did Not Form a Nanonet Response in Titrations Assays (No Sigmoid Detected)							
Surfactant	Alkyl Chain Carbon No.	Polymer	Average Polymer Diameter [nm]	Theoretical Polymer	Head Group	Surfactant Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
Anthraquinone-2-sulfonic	0	SMA 725™	15.03	6.8	Sulfonate	1.008	0.343
Cedepal TD-403 MFLD™	13	SMA 725™	15.03	6.8	Ether sulfate	6.421	0.235
n-Octanoyl-L-threonine	8	SMA 725™	15.03	6.8	Amino acid	119	0.372
Sodium dodecylbenzene sulfonate	12	SMA 725™	15.03	6.8	Sulfonate	4.235	0.53
p-Toluene sulfonic acid	1	SMA 725™	15.03	6.8	Sulfonate	0.8236	0.32
Potassium ethyl xanthogenate	2	SMA 725™	15.03	6.8	Xanthogenate	190.7	0.281
Sodium-1-pentasilfonate	5	SMA 725™	15.03	6.8	Sulfonate	0.8036	0.537
Stapan sulfonic acid™	12	SMA 725™	15.03	6.8	Sulfonate	2.821	0.186
Anthraquinone-2 sulfonic acid sodium salt	0	SMA 2000™	5.242	2.6	Sulfonate	1.008	0.343
Cedepal TD 403 MFLD™	13	SMA 2000™	5.242	2.6	Ether sulfate	6.421	0.235
n-Octanoyl-L-Threonine	8	SMA 2000™	5.242	2.6	Amino acid	119	0.372
p-Toluene Sulfonic acid	1	SMA 2000™	5.242	2.6	Sulfonate	0.8236	0.32
Potassium ethyl xanthogenate	2	SMA 2000™	5.242	2.6	Xanthogenate	190.7	0.281
Sodium-1-pentasilfonate	5	SMA 2000™	5.242	2.6	Sulfonate	0.8036	0.537
1-Octane Sulfonate	8	DIBMA	92.7	3.2	Sulfonate	1.562	0.367
Amin GCK	18, 8	DIBMA	92.7	3.2	Acyl glycinate	7.286	0.3

Table 14. Anionic Surfactants and Polymer Combinations That Did Not Form a Nanonet Response in Titrations Assays (No Sigmoid Detected)							
Surfactant	Alkyl Chain Carbon No.	Polymer	Average Polymer Diameter [nm]	Theoretical Polymer	Head Group	Surfactant Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
Anthraquinone-2 sulfonic acid sodium salt	0	DIBMA	92.7	3.2	Sulfonate	1.008	0.343
Stepan Bio Terge AS-40™	14	DIBMA	92.7	3.2	Sulfonate	6.434	0.181
n-Octanoyl-L-Threonine	8	DIBMA	92.7	3.2	Amino acid	119	0.372
p-Toluene Sulfonic Acid	1	DIBMA	92.7	3.2	Sulfonate	0.8236	0.32
Potassium ethyl xanthogenate	2	DIBMA	92.7	3.2	Xanthogenate	190.7	0.281
Sodium 1 pentasulfonate	5	DIBMA	92.7	3.2	Sulfonate	0.8036	0.537
Stepan Bio Soft D-40™	0	SMA 2021™	6.627	3.7	Sulfonate	5.13	0.322
Stepan Bio Terge AS-40™	14	SMA 2021™	6.627	3.7	Sulfonate	6.434	0.181
n-Octanoyl-L-Threonine	8	SMA 2021™	6.627	3.7	Amino acid	119	0.372
p-Toluene sulfonic acid	1	SMA 2021™	6.627	3.7	Sulfonate	0.8236	0.32
Potassium ethyl xanthogenate	2	SMA 2021™	6.627	3.7	Xanthogenate	190.7	0.281
Sodium-1-pentasulfonate	5	SMA 2021™	6.627	3.7	Sulfonate	0.8036	0.537
1-Octane sulfonate	8	SMA 3000™	3.713	2.8	Sulfonate	1.562	0.367
Amin GCK	18, 8	SMA 3000™	3.713	2.8	Acyl glycinate	7.286	0.3
Stepan Bio Soft D-40™	0	SMA 3000™	3.713	2.8	Sulfonate	5.13	0.322
Stepan Bio Terge AS-40™	14	SMA 3000™	3.713	2.8	Sulfonate	6.434	0.181
Cedepal TD-403 MFLD™	13	SMA 3000™	3.713	2.8	Ether sulfate	6.421	0.235
Dodecane-1- sulfonic acid	12	SMA 3000™	3.713	2.8	Sulfonate	5.108	0.227

Table 14. Anionic Surfactants and Polymer Combinations That Did Not Form a Nanonet Response in Titrations Assays (No Sigmoid Detected)							
Surfactant	Alkyl Chain Carbon No.	Polymer	Average Polymer Diameter [nm]	Theoretical Polymer	Head Group	Surfactant Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
n-Octanoyl-L-Threonine	8	SMA 3000™	3.713	2.8	Amino acid	119	0.372
p-Toluene sulfonic acid ethyl xanthogenate	1	SMA 3000™	3.713	2.8	Sulfonate	0.8236	0.32
Potassium xanthogenate	2	SMA 3000™	3.713	2.8	Xanthogenate	190.7	0.281
Sodium-1- pentasulfonate	5	SMA 3000™	3.713	2.8	Sulfonate	0.8036	0.537
1 Octane sulfonate	8	SMA IBE	9.759	5.4	Sulfonate	1.562	0.367
Anthraquinone-2 sulfonic	0	SMA IBE	9.759	5.4	Sulfonate	1.008	0.343
Stepan Bio Terge AS-40™	14	SMA IBE	9.759	5.4	Sulfonate	6.434	0.181
Cedepal TD 403 MFLD™	13	SMA IBE	9.759	5.4	Ether sulfate	6.421	0.235
Dodecane-1- sulfonic acid	12	SMA IBE	9.759	5.4	Sulfonate	5.108	0.227
Stepan Lathanol LAL™	12	SMA IBE	9.759	5.4	Sulfoacetate	6.814	0.151
n-Octanoyl-L- threonine	8	SMA IBE	9.759	5.4	Amino acid	119	0.372
Sodium dodecylbenzene sulfonate	12	SMA IBE	9.759	5.4	Sulfonate	4.235	0.53
Ninate 411™	12	SMA IBE	9.759	5.4	Sulfonate	216.2	0.288
p-Toluene sulfonic acid ethyl xanthogenate	1	SMA IBE	9.759	5.4	Sulfonate	0.8236	0.32
Potassium xanthogenate	2	SMA IBE	9.759	5.4	Xanthogenate	190.7	0.281
Sodium-1- pentasulfonate	5	SMA IBE	9.759	5.4	Sulfonate	0.8036	0.537
1 -Octane sulfonate	8	SMA Cumene	7.123	1.5	Sulfonate	1.562	0.367
Anthraquinone-2-sulfonic	0	SMA Cumene	7.123	1.5	Sulfonate	1.008	0.343

Table 14. Anionic Surfactants and Polymer Combinations That Did Not Form a Nanonet Response in Titrations Assays (No Sigmoid Detected)							
Surfactant	Alkyl Chain Carbon No.	Polymer	Average Polymer Diameter [nm]	Theoretical Polymer	Head Group	Surfactant Aggregate Diameter [nm]	Poly Dispersity Index (PDI)
Stepan Bio Soft D-40™	0	SMA Cumene	7.123	1.5	Sulfonate	5.13	0.322
Stepan Bio Terge AS-40™	14	SMA Cumene	7.123	1.5	Sulfonate	6.434	0.181
Cedepal TD 403 MFLD™	13	SMA Cumene	7.123	1.5	ether sulfate	6.421	0.235
Dodecane-1- sulfonic acid	12	SMA Cumene	7.123	1.5	Sulfonate	5.108	0.227
Stepan Lathanol LAL™	12	SMA Cumene	7.123	1.5	Sulfoacetate	6.814	0.151
n-Octanoyl-L-threonine	8	SMA Cumene	7.123	1.5	Amino acid	119	0.372
Ninate 411™	12	SMA Cumene	7.123	1.5	Sulfonate	216.2	0.288
p-Toluene sulfonic acid	1	SMA Cumene	7.123	1.5	Sulfonate	0.8236	0.32
Potassium xanthogenate ethyl	2	SMA Cumene	7.123	1.5	Xanthogenate	190.7	0.281
Sodium-1-penta sulfonate	5	SMA Cumene	7.123	1.5	Sulfonate	0.8036	0.537

Example 18

Nanonet Formation Prefers an Alkyl chain Length of > C7 in a Pure Surfactant Aggregate System

In this section, a study of the influence of surfactant alkyl chain length on nanonet formation was undertaken. A wide range of surfactant aggregate sizes was employed. The surfactants are listed in Table 15, including surfactant aggregate size. Despite the small alkyl chains, light scattering measurements of this class of surfactant aggregates displayed a broad range of surfactant aggregate size. Yet, in all five surfactant cases, with the exception of 1,2 hexane diol and SMA 725, nanonet formation did not occur. This was so with even the relatively smallest polymer particle size tested.

The results in Table 15 therefore suggest that, in some cases, factors other than relative size ratios may influence nanonet formation. Here, the results suggest that for surfactant aggregates composed purely of surfactants that contain under eight carbons in their linear, hydrophobic alkyl chain lengths nanonet formation is not likely to occur within the time scales studied.

Table 15. Surfactants With a Linear Alkyl Chain of Less Than Eight Carbon Atoms						
Surfactant	Surfactant Class	Head Group	Alkyl Chain Carbon no.	Conc.	Surfactant Aggregate diameter [nm]	PDI
<i>p</i> -Toluene Sulfonic Acid	Anionic	Sulfonate	6	10%	0.8236	0.32
Potassium Ethyl Xanthogenate	Anionic	Xanthogenate	2	10%	190.7	0.281
Sodium-1-pentasulfonate	Anionic	Sulfonate	5	0.25%	0.8036	0.573
1,2 -Hexanediol	Anionic	Polyol	6	1%	109.7	0.232
n-Heptyl- β -D-Thioglucopyranoside (nHDTP-H310)	Non-Ionic	Thioglucopyranoside	7	3%	8.935	0.11

Example 19

Nanonet formation prefers a soluble or emulsified surfactant aggregate with diameter greater than the theoretical compacted diameter of a SMA polymer

From earlier results, it became apparent that fatty acid-containing surfactants such as VDISTILL DV53 and VDISTILL DV63 form oils and therefore cannot be mixed with at water. As a result no surfactant aggregate diameter was measured by dynamic light scattering as the surfactant did not form a stable emulsion in water. Conversion to a soap or solubilization in other surfactants may be utilized to convert an insoluble fatty acid oil into a soluble surfactant aggregate.

Non-ionic surfactants with branched hydrophilic headgroups dominate non-ionic surfactants which do not form nanonets but satisfy nanonet formation rules. Despite having a larger measured surfactant-aggregate diameter than the tested polymer, some non-ionic surfactants were still unable to form nanonets. Of these surfactants, a disproportionate number had either a polysorbate (TWEEN™) or polyglyceryol (PGL) headgroup. It is possible that these large head groups inhibit polymer interactions with the alkyl chain of the surfactant. Another possible explanation is that the large headgroups create a surfactant aggregate shape that does not reflect a sphere, which would create a false surfactant aggregate diameter or prevent polymer association. An additional possible explanation is that these surfactants do associate with the SMA polymers, but they are not stable after precipitation and the surfactant can dissociate back into solution.

The effect of a branched hydrophilic headgroup may be visually illustrated in Figure 20. The large polyglycerol-10 headgroup class of surfactants displayed a unique trend compared to all the other surfactant classes. Instead of nanonet score increasing with decreasing HLB values, nanonet scores were higher with the more hydrophilic surfactants. However, as all of the polyglycerol-10 surfactants formed large surfactant aggregates, they still formed nanonets with at least one of all the polymers tested.

Some false positive results may be explained, in part, by error in the sizing method and concentration dependence of aggregate formation common to

surfactants. As an example, for many surfactants, increase in concentration lead some surfactants to form larger structures such as worm-like micelles, disordered aggregates, gels, and even bilayers (Gao, J., Ge, W. & Li, J. *Sc. China Ser. B-Chem.* (2005) 48: 470. <https://doi.org/10.1360/042004-71>). When DLS measurements on surfactants were made in this work, surfactant solutions were serially diluted and measured until the best PDI was obtained. This means that surfactant concentration during sizing could lie anywhere from 1 to 0.01% wt/vol surfactant concentration depending on the surfactant. However, in the nanonet formulation experiments described and employed using method #1 herein, polymer and surfactant were mixed together in a concentrated stock (1%) before dilution and precipitation in the nanonet formulation titrations. This ensured that the surfactant aggregate remained above its critical micelle concentration; and therefore was present to interact with the polymer before dilution and precipitation. As a result, the effective concentration and diameter of the surfactant aggregate in the nanonet formation experiments could be significantly higher than the concentration and recorded size at which the surfactant sizing was measured.

The measured surfactant aggregate diameter for several commercially marketed surfactants, particularly Biosoft, Bioterge, Cedepal, and Ninate 411, was larger than the theoretical polymer diameter for each tested polymer (Table 15). Thus, nanonet formation was expected to occur using these surfactants with all the tested polymers. However, while each surfactant could be formulated into a nanonet with at least one polymer, other factors lead to not all combinations resulting in the formation of nanonets. For example, only select polymers formed nanonets with the following surfactants: Stepan Bio-Soft D-40™, Stepan Bio-Terge AS-40™, Cedepal TD-403 MFLD™, and Ninate 411™ surfactants (Table 15). Further, the Ninate 411™ surfactant showed no sign of the formation of nanonets with the following polymers: SMA-IBE and SMA cumene (Table 15). For these two polymers, a significant difference between DLS measured and theoretical particle diameter was observed (5.8 nm by DLS and 2.8 nm theoretical particle diameter). Additionally, these two polymers also contain

different ratios of hydrophobic to hydrophilic functional groups in their repeating block units.

It is therefore unlikely that trends in polymer particle size or block hydrophobicity are preventing the formation of nanonets with Ninate 411™. What is more likely is that the tested commercially-available surfactants are not pure chemicals, and therefore may have secondary components that alter polymer behavior or that complicate sizing measurements. For example, two different Bio-Terge formulations are available for commercial purchase, Bio-Terge 40S or Bio-Terge 40K, both of which contain sodium C₁₄-C₁₆ olefin sulfonate as the primary active ingredient. However, while the 40S formulation is suitable for applications in preparing stable microemulsions, the 40k formulation is only recommended as a general washing detergent (Manufacturers product sheet).

It is likely that the undisclosed additives, solvents or remnants of the surfactant manufacturing process in these formulations can negatively affect nanonet formation. It may also be the case that there is inaccurate surfactant sizing data. This may explain some of the inconsistencies between predicted and actual observation in nanonet formation experiments between the different polymers. For example, if a given polymer surfactant combination did not result in the formation of nanonets with high efficiency, a stabilizing ligand or co-surfactant on the outside of the surfactant micelle (such as isopropyl amine in the case of Ninate 411™), may shield the surfactant aggregate and inhibit the formation of nanonets.

The size differential between a surfactant aggregate and an amphipathic polymer particle, driven by the hydrophobic effect has been tested using 7 different polymers and over 50 unique surfactants. The data strongly support the proposition that the formation of nanonets requires a surfactant aggregate with a larger diameter than the diameter of the minimum monomeric particle diameter with which the surfactant aggregate is associating. In addition, the formation of nanonets using the tested polymers was strongly favored to occur with the following: (1) greater than 7 carbons in the hydrophobic alkyl chain on the

surfactant, (2) the surfactant is water miscible, (3) surfactants with relatively smaller head groups. Taken together, the results suggest that nanonet design may be enhanced by selecting an amphipathic block co-polymer, the length of which can be based on the surfactant aggregate size. The vast majority of formed nanonets displayed an increase in diameter relative to the starting polymer (Figure 18).

Example 20

Polymer composition influences surfactant capture into Nanonets. A study of surfactants and the influence of parameters including degree of water miscibility was undertaken with respect to nanonet formation. In the study, factors found influential in forming desirable nanonets included the following: A water soluble, amphipathic block co-polymer with a block stoichiometry of X_nY_m . X is an anionic or non-ionic hydrophilic functional group. Y is a hydrophobic functional group. In the polymer formula, $n = 1$ and $m = 1, 2$ or 3 .

The nanonet library was expanded by testing polymers with different functional groups. Factors explored included, polymer length, ratios of polymer functional groups, two distinct hydrophobic chemical groups. The tested polymers are outlined in Table 16. For all tested polymers, nanonets could be formed with surfactant aggregates larger than the compact polymer particle diameter (Figure 19). The maximum sigmoid AUC scores for nanonet formation were observed with polymers that had a 2:1 styrene to maleic acid ratio, followed by 3:1, followed by the 1:1. Relatively shorter polymers generally had higher scores for nanonet formation.

Table 16. Dynamic Light Scattering Characterization using Intensity and Number Values of Hydrolyzed Polymer Aggregates

Polymer	[St] to [MA] ratio	Calculated Aggregate Diameter [nm]	Conc.	Aggregate Diameter [nm] (Intensity Mean d)	PDI	Conc.	Aggregate Diameter [nm] (Number Mean d)	PDI
SMA-2000™	2:1	2.6	0.10%	5.242	0.113	0.10%	4.633	0.292
SMA-2021™	2:1	3.7	0.25%	6.627	0.274	0.10%	8.383	0.169

Table 16. Dynamic Light Scattering Characterization using Intensity and Number Values of Hydrolyzed Polymer Aggregates								
Polymer	[St] to [MA] ratio	Calculated Aggregate Diameter [nm]	Conc.	Aggregate Diameter [nm] (Intensity Mean d)	PDI	Conc.	Aggregate Diameter [nm] (Number Mean d)	PDI
SMA-3000™	3:1	2.8	0.25%	3.713	0.213	0.01%	4.68	0.355
SMA-725™	3:1	6.8	0.10%	15.03	0.219	0.10%	10.16	0.283
SMA- cumene terminated	3:1	1.5	0.10%	7.123	0.164	0.50%	3.325	0.297
SMA-IBE	1:1	5.4	0.25%	9.759	0.209	0.01%	4.256	0.869
DIBMA	NA	3.2	0.50%	76.74*	0.135	0.10%	5.704	0.271

Not all possible combinations of surfactant and polymer are likely to form nanonets within the time frame of the screening methods employed (see Example 15). 33/116 combinations of polymer plus surfactant did not result in a score indicating nanonet formation by screening method #1 (see Reagents and Materials Examples 15-21). These results (no nanonet formation) are able to be predicted using the teachings described herein. However, 5.4% (14/306) or of the formulations screened in method #2 (see Reagents and Materials Examples 15-21) resulted in a score indicating the formation of nanonets. Yet, based upon relative size alone, nanonet formation would not have been predicted. The surfactant aggregate diameter was smaller than the polymer particle diameter. Thus, nanonet formation would not have been predicted to occur using the teaching herein. These false positive results may be attributable to: In screening method #1, a relatively small rise in absorbance may have been construed by the developed computer model as sigmoid curve formation. The result would have been a low, but nonetheless positive score indicating nanonet formation; even when no nanonets were formed. False negative results may be attributable to: insufficient titration points; in some traces Nanonet formation occurred over only one tested ratio, meaning the window of association is very small. In other traces the titration curves only began to increase at the very highest concentrations of the titration. These signals would be missed in the current experiments.

ROUT Method of Statistical Analysis for Outlier Datapoint Identification

A rational basis for removing some data points initially identified by the model was also developed. For example, the results were further screened to remove those cases with low formation factors, N_f and absorbance values less than 0.2 (Table 9 and 10). The dataset for the surfactant polymer combinations analyzed by screening method #4 (see Reagents and Materials Examples 15-21) for nanonet formation were analyzed for outliers. The method employed was similar that described by Motulsky, H.J., Brown, and R.E. Detecting in *Outliers When Fitting Data With Nonlinear Regression – A New Method Based On Robust Nonlinear Regression And The False Discovery Rate*. BMC Bioinformatics 7, 123 (2006)). Using the ROUT method, outliers in surfactant size in both the positively scoring nanonets and negatively scoring nanonet datasets were identified.

Thus, screening plus ROUT analysis reduced the number of false positive results to 9/192 results. This means in only 9/192 cases, a false positive score for nanonet formation was obtained.

Factors other than size relationship may also explain the results. These factors may include: (1) surfactant alkyl chain carbon number, (2) surfactant water solubility, and (3) surfactant hydrophobic lipid balance. Additionally, there may be some sources of error in surfactant aggregate and polymer particle size measurement. Other reasons for the disparity between expected and observed results may include: (1) surfactant composition, (2) surfactant purity, (3) polymer composition, (4) molecular weight distribution, (5) surfactant headgroup size, (6) surfactant packing parameters, and (7) co-monomer distribution. For example, in several cases, our measures of polymer contour length and molecular weight varied from the specifications provided with commercially available samples. What follows is a more detailed discussion of these other factors.

Surfactant Alkyl Chain Carbon Number.

five of the 62 tested surfactants contained small hydrophobic segments (linear alkyl chain with under 7 carbons). Interestingly, five of these surfactants

did not result in a score indicating nanonet formation using method #1 (titration assay, Table 15). This was the case with all tested polymers with the exception of surfactant 1,2 hexane diol with SMA-725

DLS measurements of this class of low carbon number surfactant aggregates displayed a broad range of surfactant aggregate sizes. This latter result suggests that for aggregates composed purely of surfactants that contain under eight carbons in their linear, hydrophobic alkyl chains, nanonet formation will not likely occur. Thus, the more general conclusion is that nanonet formation is more likely to occur when the surfactant has an alkyl chain carbon number of greater than seven when measured as a pure surfactant aggregate. The effect of alkyl chain number can be further visualized by examining nanonet formation with select surfactant head groups and varying alkyl chains (Figure 21). In almost all cases, an increase in alkyl chain number lead to an increase in nanonet score, and increased likelihood of forming a nanonet with the tested polymers (Figs. 21A-D). Similarly, nanonet scores generally displayed an inverse effect with hydrophilic headgroups: as HLB values increased due to an increase in number of hydrophilic headgroups, nanonet scores decreased (Figs. 21E and F). This effect of alkyl chain length and HLB in the surfactant on nanonet scores, and compatibility with different polymers suggests that nanonet formation occurs best with the 2:1 block ratio. As the polymer composition shifts from 2:1 to either 3:1 or 1:1, the number of possible nanonets to be formed generally decreases. Furthermore, as polymer length decreases, nanonets can be formed with smaller and more hydrophilic surfactant aggregates (high HLB values). It is likely that the inverse is also true: the more hydrophobic the surfactant, the larger the polymer and more hydrophobic block is preferred. For example, the 1,2-Hexanediol or VDISTILL 53™, with calculated HLB values of 7.9 (Using the Davies method, J.T. Davies, E.K. Rideal, Interfacial Phenomena, Academic Press, New York, 1961, p. 371.) and 1, respectively, were only found to associate with the most hydrophobic polymer; SMA-725™.

Example 21

Formation of Optimized Nanonet Library:

For chemical treatments of water, little to no contamination of downstream effluents by the introduced chemical is required. Therefore in a nanonet formulation the majority of surfactant must be bound to the polymer, to minimize any residual surfactant downstream in the treatment process. To identify the optimum amount of surfactant, the titration data for a given surfactant can be used. The midway point on each titration curve, also referred to as the N_f , represents the half maximal surfactant binding. The midway point was calculated according to method #4 (see Reagents and Materials Examples 15-21). At this concentration it is expected that polymer is in excess, so there is minimal surfactant in solution, and at least half of all polymer is expected be in a nanonet complex. Surfactants with high midway points (>500ppm) for a given polymer, were considered to have low affinity and therefore these combinations were not identified to form stable nanonets.

In order to better identify stable nanonets, the height of absorption peak of each surfactant and polymer combination was filtered to 0.2 and higher to obtain an optimized nanonet library. In the titration data, a low peak absorbance is representative of little surfactant aggregate being captured by the polymer, and is therefore an inefficient nanonet combination and likely to lead to high concentrations of residual surfactant post-removal of any formed nanonet. From these results, it should be understood that these filtered combinations may still form small numbers of nanonet complexes, but they are non-optimal and that a different polymer may be more appropriate.

Using this approach, a nanonet library containing approximately 199 stable, optimized nanonets was identified. The nanonet library contains a broad range of anionic and non-ionic surfactants with linear alkyl chain lengths varying from C₈ to C₁₈. Hydrophilic headgroups contained within the nanonet library are diverse in nature, including sulfonates, phosphate esters, phosphine oxides, monosaccharides, disaccharides, thioesters, amino acids, glycols, and polyglycerols. The full nanonet library is documented in Tables 9 and 10.

Page furnished blank

Table 17. Copper Removal (40uL NanoNet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Cu ²⁺ Added [ppm]	Measured [ppm]	% Difference
A1	1	Apo 10 / DIBMA	10.03	9.9	-1.9
A2	2	Apo 12 / DIBMA	10.03	6.8	-32.4
A3	3	Coco glucoside / DIBMA	10.03	10.4	3.3
A4	4	Genapol X080 / DIBMA	10.03	10.1	0.3
A5	5	HEGMDDE-H370 / DIBMA	10.03	8	-20.7
A6	6	HEGMDE-H360 / DIBMA	10.03	10.4	3.5
A7	7	HXEGMDDE-H375 / DIBMA	10.03	10.1	-0.2
A8	8	DDDM-MNA-C12 / DIBMA	10.03	7.8	-22.2
A9	9	pGBTM-MNA-C13 / DIBMA	10.03	9.3	-8
A10	10	pGBTEM-MNA-C14 / DIBMA	10.03	8	-20.9
A11	11	nDDTTP-D342 / DIBMA	10.03	10	-1.2
A12	12	nDDTGP-D323 / DIBMA	10.03	8.9	-11.4
B1	13	OEGMDE-O330 / DIBMA	10.03	7.1	-29.4
B2	14	Polyglycerol-10-mono-laurate / DIBMA	10.03	9.5	-6.1
B3	15	Polyglycerol-10-mono-oleate / DIBMA	10.03	7.1	-29.6
B4	16	Span 20 / DIBMA	10.03	10.4	3.6
B5	17	Triton X-100™ / DIBMA	10.03	9.7	-3.2
B6	18	Triton X-45™ / DIBMA	10.03	9.9	12.3
B7	19	Tween 20™ / DIBMA	10.03	7.4	-26.9
B8	20	Tween 65™ / DIBMA	10.03	7.8	-23
B9	21	Tween 85™ / DIBMA	10.03	9.5	-5.2
B10	22	Apo 10 / SMA 2000™	10.03	0.9	-91.1
B11	23	Apo 12 / SMA 2000™	10.03	0.5	-95.4
B12	24	Coco glucoside / SMA 2000™	10.03	1	-90.1
C1	25	Genapol X080 / SMA 2000™	10.03	1	-90.2
C2	26	HEGMDDE-H370 / SMA 2000™	10.03	1.5	-85.2
C3	27	HEGMDE-H360 / SMA 2000™	10.03	0.9	-90.6
C4	28	HEGMOE-H350 / SMA 2000™	10.03	1.4	-85.7
C5	29	HXEGMDDE-H375 / SMA 2000™	10.03	1.1	-89.4
C6	30	pGBTM-MNA-C13 / SMA 2000™	10.03	0.8	-91.6

Table 17. Copper Removal (40uL NanoNet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Cu2+ Added [ppm]	Measured [ppm]	% Difference
C7	31	pGBTEM-MNA-C14 / SMA 2000™	10.03	1.2	-88.1
C8	32	nDDDTG-D342 / SMA 2000™	10.03	1	-90
C9	33	nDDTGP-D323 / SMA 2000™	10.03	0.9	-90.9
C10	34	nDDTP-D335 / SMA 2000™	10.03	1.3	-87.5
C11	35	OEGMDE-O330 / SMA 2000™	10.03	0.9	-91
C12	36	Polyglycerol-10-mono-laurate / SMA 2000™	10.03	1.2	-87.8
D1	37	Polyglycerol-10-mono-oleate / SMA 2000™	10.03	1	-90.1
D2	38	Span 20 / SMA 2000™	10.03	1	-89.8
D3	39	Triton X-100™ / SMA 2000™	10.03	1.1	-89.4
D4	40	Triton X-305™ / SMA 2000™	10.03	1.1	-88.7
D5	41	Triton X-45™ / SMA 2000™	10.03	1	-89.9
D6	42	Tween 20™ / SMA 2000™	10.03	1.2	-88.5
D7	43	Tween 40™ / SMA 2000™	10.03	0.6	-93.6
D8	44	Tween 60™ / SMA 2000™	10.03	1.1	-89.1
D9	45	Tween 65™ / SMA 2000™	10.03	1.3	-86.6
D10	46	Tween 80™ / SMA 2000™	10.03	1.1	-89.2
D11	47	Tween 85™ / SMA 2000™	10.03	0.8	-91.6
D12	48	Apo 10 / SMA 2021™	10.03	1.4	-86
E1	49	Apo 12 / SMA 2021™	10.03	1.2	-88.3
E2	50	Coco glucoside / SMA 2021™	10.03	1.1	-88.9
E3	51	HEGMDDE-H370 / SMA 2021™	10.03	1	-89.6
E4	52	HEGMDE-H360 / SMA 2021™	10.03	1.1	-89.5
E5	53	HXEGMDDE-H375 / SMA 2021™	10.03	1	-89.8
E6	54	pGBTM-MNA-C13 / SMA 2021™	10.03	1.2	-87.7
E7	55	pGBTEM-MNA-C14 / SMA 2021™	10.03	0.9	-90.8
E8	56	nDDTGP-D323 / SMA 2021™	10.03	1	-90.1
E9	57	nDDDTG-D342 / SMA 2021™	10.03	1.2	-88.2
E10	58	OEGMDE-O330 / SMA 2021™	10.03	1.2	-88.3
E11	59	Polyglycerol-10-mono-laurate / SMA 2021™	10.03	0.9	-90.6

Table 17. Copper Removal (40uL NanoNet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Cu ²⁺ Added [ppm]	Measured [ppm]	% Difference
E12	60	Polyglycerol-10-mono-oleate / SMA 2021™	10.03	0.8	-92.5
F1	61	Triton X-100™ / SMA 2021™	10.03	1.9	-81.5
F2	62	Triton X-45™ / SMA 2021™	10.03	1.3	-87.2
F3	63	Tween 40™ / SMA 2021™	10.03	1.2	-87.8
F4	64	Tween 60™ / SMA 2021™	10.03	1.2	-88.3
F5	65	Tween 80™ / SMA 2021™	10.03	2.2	-78.1
F6	66	Tween 85™ / SMA 2021™	10.03	1.2	-88.1
F7	67	Apo 10 / SMA 3000™	10.03	3.9	-61.5
F8	68	Apo 12 / SMA 3000™	10.03	4	-59.9
F9	69	Coco glucoside / SMA 3000™	10.03	5.4	-46.4
F10	70	Genapol X080 / SMA 3000™	10.03	0.8	-91.8
F11	71	HEGMDDE-H370 / SMA 3000™	10.03	3.8	-61.9
F12	72	HEGMDE-H360 / SMA 3000™	10.03	4.1	-59.6
G1	73	HXEGMDDE-H375 / SMA 3000™	10.03	1.2	-88.2
G2	74	DDDM-MNA-C12 / SMA 3000™	10.03	4.6	-54
G3	75	pGBTM-MNA-C13 / SMA 3000™	10.03	4.3	-57.4
G4	76	pGBTEM-MNA-C14 / SMA 3000™	10.03	4.8	-52.5
G5	77	nDDDT-D342 / SMA 3000™	10.03	5.6	-44.3
G6	78	nDDTG-D323 / SMA 3000™	10.03	1.6	-84.4
G7	79	OEGMDE-O330 / SMA 3000™	10.03	3.5	-65.6
G8	80	Polyglycerol-10-mono-laurate / SMA 3000™	10.03	0.8	-92.2
G9	81	Polyglycerol-10-mono-oleate / SMA 3000™	10.03	0.5	-94.7
G10	82	Polyglycerol-10-mono-palmitate / SMA 3000™	10.03	1.2	-87.9
G11	83	Span 20 / SMA 3000™	10.03	0.6	-93.6
G12	84	Triton X-100 / SMA 3000™	10.03	1.8	-81.7
H1	85	Triton X-45™ / SMA 3000™	10.03	1.1	-89.2
H2	86	Tween 65™ / SMA 3000™	10.03	1.1	-89.2
H3	87	Tween 85™ / SMA 3000™	10.03	4	-60.6
H4	88	Apo 12 / SMA-IBE	10.03	1.4	-86
H5	89	Coco glucoside / SMA-IBE	10.03	1.6	-83.9

Table 17. Copper Removal (40uL NanoNet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Cu2+ Added [ppm]	Measured [ppm]	% Difference
H6	90	HEGMDDE-H370 / SMA-IBE	10.03	1.1	-92.8
H7	91	HEGMDE-H360 / SMA-IBE	10.03	1.6	-84
H8	92	HXEGMDDE-H375 / SMA-IBE	10.03	1.5	-85
H9	93	nDDDTTP-D342 / SMA-IBE	10.03	1.5	-85.1
H10	94	nDDTP-D335 / SMA-IBE	10.03	1.2	-88.1
H11	95	Polyglycerol-10-mono-oleate / SMA-IBE	10.03	1.2	-88.3
H12	96	Triton X-45™ / SMA-IBE	10.03	1	-89.7
A1	97	Tween 85™/ SMA-IBE	10.03	1.5	-85.1
A2	98	Genapol X080 / SMA 725™	10.03	2	-79.8
A3	99	nDDDTTP-D342 / SMA 725™	10.03	0.9	-91.1
A4	100	nDDTGP-D323/ SMA 725™	10.03	1	-89.6
A5	101	Polyglycerol-10-mono-oleate / SMA 725™	10.03	0.5	-94.7
A6	102	Span 20 / SMA 725™	10.03	1.1	-88.6
A7	103	HEGMDDE-H370 / SMA 725™	10.03	1.5	-85.3
A8	104	HEGMDE-H360 / SMA 725™	10.03	1.9	-81.4
A9	105	HXEGMDDE-H375 / SMA 725™	10.03	1.3	-86.7
A10	106	pGBTEM-MNA-C14 / SMA 725™	10.03	1.7	-83.5
A11	107	Polyglycerol-10-mono-palmitate / SMA 725™	10.03	1	-90.1
A12	108	Triton X-100™ / SMA 725™	10.03	2.4	-76.6
B1	109	Triton X-45™ / SMA 725™	10.03	1.4	-85.6
B2	110	Tween 65™ / SMA 725™	10.03	1.3	-86.7
B3	111	Tween 80™ / SMA 725	10.03	1.6	-83.7
B4	112	Apo 10 / SMA Cumene	10.03	0.8	-91.7
B5	113	Apo 12 / SMA Cumene	10.03	1	-90.4
B6	114	PFDMP-O310F / SMA Cumene	10.03	1.1	-89.4
B7	115	Genapol X080 / SMA Cumene	10.03	0.9	-91
B8	116	HEGMDDE-H370 / SMA Cumene	10.03	0.9	-90.8
B9	117	HEGMDE-H360 / SMA Cumene	10.03	0.9	-90.8
B10	118	HXEGMDDE-H375 / SMA Cumene	10.03	0.8	-92.1
B11	119	DDDM-MNA-C12 / SMA Cumene	10.03	0.7	-92.9

Table 17. Copper Removal (40uL NanoNet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Cu ²⁺ Added [ppm]	Measured [ppm]	% Difference
B12	120	pGBTM-MNA-C13/ SMA Cumene	10.03	0.6	-93.6
C1	121	nDDDTp-D342 / SMA Cumene	10.03	1.3	-87.4
C2	122	nDDTGP-D323 / SMA Cumene	10.03	0.8	-92
C3	123	nDDTP-D335 / SMA Cumene	10.03	0.8	-92.1
C4	124	OEGMDE-O330 / SMA Cumene	10.03	1.3	-87.4
C5	125	Polyglycerol-10-mono-laurate / SMA Cumene	10.03	1.4	-86.5
C6	126	Polyglycerol-10-mono-oleate / SMA Cumene	10.03	1.2	-88.4
C7	127	Span 20 / SMA Cumene	10.03	0.7	-93.3
C8	128	Triton X-100™ / SMA Cumene	10.03	1	-90.4
C9	129	Triton X-45™ / SMA Cumene	10.03	1.2	-88.3
C10	130	Tween 65™ / SMA Cumene	10.03	1.4	-85.9
C11	131	Tween 85™ / SMA Cumene	10.03	1.2	-87.9

Table 18. Copper Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Anionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Cu ²⁺ Added [ppm]	Measured [ppm]	% Difference
A1	1	Stepan Bio-Soft D-40™ / DIBMA	10.03	9.2	-8.2
A2	2	Lakeland PAE136™ / DIBMA	10.03	8.9	-11.6
A3	3	Stepan Lathanol LAL™ / DIBMA	10.03	9.6	-4.5
A4	4	n-Decanoyl-L-threonine / DIBMA	10.03	9.4	-6
A5	5	n-Dodecanoyl-L-threonine / DIBMA	10.03	9.5	-5.2
A6	6	n-Decanoyl-L-methionine / DIBMA	10.03	9	-10.7
A7	7	n-Dodecanoyl-L-methionine / DIBMA	10.03	8.8	-12.4
A8	8	Ninate 411™ / DIBMA	10.03	9.8	-2.3
A9	9	n-Octanoyl-L-methionine / DIBMA	10.03	9.3	-7.5
A10	10	Stepan Mild L3™ / DIBMA	10.03	8.5	-15.1
A11	11	Stepan Bio-Terge AS-40™ / SMA 2000™	10.03	1.3	-87.2
A12	12	Stepan Bio-Soft D-40™ / SMA 2000™	10.03	1.2	-88.2
B1	13	Dodecane-1-sulfonic acid sodium salt / SMA 2000™	10.03	1.5	-85.1

Table 18. Copper Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Anionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Cu ²⁺ Added [ppm]	Measured [ppm]	% Difference
B2	14	Lakeland PAE 136™ / SMA 2000™	10.03	0.8	-91.7
B3	15	Stepan Lathanol LAL™ / SMA 2000™	10.03	1.2	-88.4
B4	16	n-Decanoyl-L-methionine / SMA 2000™	10.03	1.4	-86.3
B5	17	n-Dodecanoyl-L-methionine / SMA 2000™	10.03	1.6	-83.6
B6	18	n-Decanoyl-L-threonine / SMA 2000™	10.03	1.5	-84.9
B7	19	n-Dodecanoyl-L-histidine / SMA 2000™	10.03	0.7	-93.4
B8	20	Stepan Mild L3™ / SMA 2000™	10.03	1.4	-86.3
B9	21	n-Dodecanoyl-L-threonine / SMA 2000™	10.03	1.4	-86
B10	22	Sodium dodecylbenzene sulfonate / SMA 2000™	10.03	1.4	-86.3
B11	23	Ninate 411™ / SMA 2000™	10.03	1.5	-85.1
B12	24	Stepan SLL-FB™ / SMA 2000™	10.03	0.9	-90.9
C1	25	Stepan Sulfonic Acid™ / SMA 2000™	10.03	2.5	-75.5
C2	26	Dodecane-1-sulfonic acid sodium salt / SMA 2021™	10.03	1.1	-88.9
C3	27	Lakeland PAE 136™ / SMA 2021™	10.03	1.3	-87.2
C4	28	Stepan Lathanol LAL / SMA 2021™	10.03	1.4	-86.3
C5	29	n-Decanoyl-L-methionine / SMA 2021™	10.03	1.1	-89
C6	30	Cedepal TD-403 MFLD™ / SMA 2021™	10.03	1.2	-88.1
C7	31	Stepan Mild L3™ / SMA 2021™	10.03	9.2	-8.7
C8	32	n-Dodecanoyl-L-methionine / SMA 2021™	10.03	1.6	-84.3
C9	33	n-Decanoyl-L-methionine / SMA 3000™	10.03	4.6	-54.6
C10	34	n-Dodecanoyl-L-methionine / SMA 3000™	10.03	4.6	-54.3
C11	35	n-Decanoyl-L-threonine / SMA 2021™	10.03	1.9	-81.2
C12	36	n-Dodecanoyl-L-threonine / SMA 2021™	10.03	1.6	-84.3

Table 18. Copper Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Anionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Cu ²⁺ Added [ppm]	Measured [ppm]	% Difference
D1	37	Sodium dodecylbenzene sulfonate / SMA 2021™	10.03	1.5	-84.7
D2	38	Ninate 411™ / SMA 2021™	10.03	1.5	-85.4
D3	39	Stepan SLL-FB™ / SMA 2021™	10.03	2.1	-79.1
D4	40	Stepan Sulfonic Acid™ / SMA 2021™	10.03	1.7	-82.7
D5	41	Lakeland PAE 136™ / SMA 3000™	10.03	4.1	-59.6
D6	42	n-Dodecanoyl-L-threonine / SMA 3000™	10.03	4.5	-54.9
D7	43	Ninate 411™ / SMA 3000™	10.03	3.2	-68.3
D8	44	Stepan Mild L3™ / SMA 3000™	10.03	4.6	-54.2
D9	45	Stepan SLL-FB™ / SMA 3000™	10.03	4.2	-58
D10	46	Stepan Sulfonic Acid™ / SMA 3000™	10.03	3.4	-66.5
D11	47	n-Decanoyl-L-methionine / SMA-IBE	10.03	2	-79.8
D12	48	n-Dodecanoyl-L-methionine/ SMA-IBE	10.03	1.4	-85.8
E1	49	Stepan Mild L3™ / SMA-IBE	10.03	1.7	-83.5
E2	50	Stepan SLL-FB™ / SMA-IBE	10.03	1.6	-84
E3	51	Amin GCK30H / SMA 725™	10.03	2	-79.8
E4	52	Stepan Bio-Terge AS-40™ / SMA 725™	10.03	1.5	-85.3
E5	53	n-Dodecanoyl-L-histidine / SMA 725™	10.03	1.1	-88.7
E6	54	Stepan Lathanol LAL™ / SMA 725™	10.03	1.3	-86.9
E7	55	n-Decanoyl-L-methionine / SMA 725™	10.03	1.7	-83.5
E8	56	n-Dodecanoyl-L-methionine / SMA 725™	10.03	1.6	-84.2
E9	57	Dodecane-1-sulfonic acid sodium salt / SMA 725™	10.03	1.4	-86.1
E10	58	Lakeland PAE 136™ / SMA 725™	10.03	1.6	-84.2
E11	59	n-dodecanoyl-L-threonine / SMA 725™	10.03	1.3	-87.4
E12	60	Stepan Mild L3™ / SMA 725™	10.03	1.5	-84.8
F1	61	Stepan SLL-FB™ / SMA 725™	10.03	1.3	-86.6
F2	62	Lakeland PA E136™ / SMA Cumene	10.03	1.3	-87
F3	63	n-Dodecanoyl-L-threonine / SMA Cumene	10.03	2	-80

Table 18. Copper Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Anionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Cu ²⁺ Added [ppm]	Measured [ppm]	% Difference
F4	64	Stepan Mild L3™ / SMA Cumene	10.03	1.4	-85.7
F5	65	Stepan SLL-FB™ / SMA Cumene	10.03	1.3	-86.6
F6	66	n-Decanoyl-L-methionine / SMA Cumene	10.03	1.4	-85.6
F7	67	n-Dodecanoyl-L-methionine / SMA Cumene	10.03	3.1	-68.9
F8	68	n-Dodecanoyl-L-histidine / SMA Cumene	10.03	1.4	-85.6

Example 22

Proof of Concept Screen of Nanonet Library For Sequestering Divalent Metal Ions Zinc and Copper Found In Seawater

Metals contained in seawater are a threat to aquatic systems and organisms. Removal of heavy metals from seawater or wastewater is often approached by precipitation of dissolved metal ion followed by filtration. However, these methods can be challenging as they require control over pH of seawater.

The nanonet libraries with anionic and non-ionic surfactants were screened for zinc and copper metal ion removal from seawater spiked with zinc and copper metal ions (see Reagents and Materials Examples 15-21, method #3). All hydrolyzed SMA-containing polymers showed excellent Cu²⁺ metal ion removal from seawater especially the polymers SMA-2000™, SMA-2021™, and SMA-725™ with surfactants with sulfonate and amino acid-containing head groups (Figure 22). In contrast, nanonets with DIBMA showed remaining Cu²⁺ metal ion (< 10ppm) in seawater. Figure 23 illustrates excellent Cu²⁺ metal ion removal from seawater with nanonets with non-ionic surfactants and SMA-2000™, SMA-2021™, SMA-Cumene terminated™, and SMA-725™. Contrary, nanonets with SMA-3000™ and DIBMA™ were less efficient in metal ion removal. Overall, nanonets with anionic surfactants were less efficient for zinc metal ion removal from seawater, specifically in combination with SMA-3000™

and DIBMATM (Figure 24). Interestingly, the surfactant *N*-Dodecanoyl-L-Histidine in combination with SMA-2000TM, SMA-725TM, and SMA-Cumene terminatedTM was most efficient in Zn²⁺ metal ion removal along with SMA-725TM and Dodecane-1-sulfonic acid (Figure 24). Nanonets with non-ionic surfactants and SMA-2000TM and SMA-Cumene terminatedTM removed zinc metal ion with less than 5 ppm remaining Zn²⁺ in solution (Figure 25). Nanonets with SMA-3000TM and DIBMATM were less efficient in metal ion removal. This work demonstrates that new water treatment formulations that are specific for single metal species can be rapidly identified through screening of nanonet libraries.

Table 19. Zinc Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Zn ²⁺ Added [ppm]	Measured [ppm]	% Difference
A1	1	Apo 10 / DIBMA	10.07	11.6	15.3
A2	2	Apo 12 / DIBMA	10.07	10.1	0.6
A3	3	Coco glucoside / DIBMA	10.07	11.5	13.7
A4	4	Genapol X080 / DIBMA	10.07	11.2	10.8
A5	5	HEGMDDE-H370 / DIBMA	10.07	11.7	16.1
A6	6	HEGMDE-H360 / DIBMA	10.07	11.6	15.5
A7	7	HXEGMDDE-H375 / DIBMA	10.07	11.7	16
A8	8	DDDM-MNA-C12 / DIBMA	10.07	11.2	11.6
A9	9	pGBTM-MNA-C13 / DIBMA	10.07	11.3	12.4
A10	10	pGBTEM-MNA-C14 / DIBMA	10.07	11.3	12.4
A11	11	nDDDTp-D342 / DIBMA	10.07	11	9.1
A12	12	nDDTGP-D323 / DIBMA	10.07	11.3	12.6
B1	13	OEGMDE-O330 / DIBMA	10.07	10.5	4.7
B2	14	Polyglycerol-10-mono-laurate / DIBMA	10.07	11.3	12.4
B3	15	Polyglycerol-10-mono-oleate / DIBMA	10.07	10.9	8.6
B4	16	Span 20 TM / DIBMA	10.07	10.4	3.8
B5	17	Triton X-100 TM / DIBMA	10.07	11	9.5
B6	18	Triton X-45 TM / DIBMA	10.07	11.9	18.3
B7	19	Tween 20 TM / DIBMA	10.07	10.7	6.2
B8	20	Tween 65 TM / DIBMA	10.07	10.7	6.1

Table 19. Zinc Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Zn2+ Added [ppm]	Measured [ppm]	% Difference
B9	21	Tween 85™ / DIBMA	10.07	10.6	5.4
B10	22	Apo 10 / SMA 2000™	10.07	5.3	-47
B11	23	Apo 12 / SMA 2000™	10.07	5.5	-45.1
B12	24	Coco glucoside / SMA 2000™	10.07	5	-50.2
C1	25	Genapol X080 X080 / SMA 2000™	10.07	5.2	-48.8
C2	26	HEGMDDE-H370 / SMA 2000™	10.07	5.3	-47
C3	27	HEGMDE-H360 / SMA 2000™	10.07	5.7	-43.1
C4	28	HEGMOE-H350 / SMA 2000™	10.07	5.5	-45.8
C5	29	HXEGMDDE-H375 / SMA 2000™	10.07	4.6	-54.6
C6	30	pGBTM-MNA-C13 / SMA 2000™	10.07	5.5	-45.3
C7	31	pGBTEM-MNA-C14 / SMA 2000™	10.07	5	-50.5
C8	32	nDDDTTP-D342 / SMA 2000™	10.07	5.1	-49.7
C9	33	nDDTGP-D323 / SMA 2000™	10.07	5.2	-48
C10	34	nDDTP-D335 / SMA 2000™	10.07	5.4	-46.4
C11	35	OEGMDE-O330 / SMA 2000™	10.07	5.2	-48.4
C12	36	Polyglycerol-10-mono-laurate / SMA 2000™	10.07	4.9	-51.8
D1	37	Polyglycerol-10-mono-oleate / SMA 2000™	10.07	4.7	-53.3
D2	38	Span 20 / SMA 2000™	10.07	5.6	-44.7
D3	39	Triton X-100™ / SMA 2000™	10.07	5.9	-41.4
D4	40	Triton X-305™ / SMA 2000™	10.07	5	-50.4
D5	41	Triton X-45™ / SMA 2000™	10.07	5	-50.7
D6	42	Tween 20™ / SMA 2000™	10.07	5	-50.1
D7	43	Tween 40™ / SMA 2000™	10.07	4.6	-54.2

Table 19. Zinc Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Zn2+ Added [ppm]	Measured [ppm]	% Difference
D8	44	Tween 60™ / SMA 2000™	10.07	5.1	-49
D9	45	Tween 65™ / SMA 2000™	10.07	5.6	-44.3
D10	46	Tween 80™ / SMA 2000™	10.07	9	-10.2
D11	47	Tween 85™ / SMA 2000™	10.07	4.8	-52
D12	48	Apo 10 / SMA 2021™	10.07	6.7	-33.4
E1	49	Apo 12 / SMA 2021™	10.07	6.6	-34.1
E2	50	Coco glucoside / SMA 2021™	10.07	6.9	-31.9
E3	51	HEGMDDE-H370 / SMA 2021™	10.07	7.2	-28.5
E4	52	HEGMDE-H360 / SMA 2021™	10.07	7.2	-28.9
E5	53	HXEGMDDE-H375 / SMA 2021™	10.07	7.2	-28.6
E6	54	pGBTM-MNA-C13 / SMA 2021™	10.07	7.3	-27.3
E7	55	pGBTEM-MNA-C14 / SMA 2021™	10.07	7.5	-26
E8	56	nDDTGP-D323 / SMA 2021™	10.07	6.9	-31.6
E9	57	nDDDTP-D342 / SMA 2021™	10.07	7.2	-28.8
E10	58	OEGMDE-O330 / SMA 2021™	10.07	6.8	-32.5
E11	59	Polyglycerol-10-mono-laurate / SMA 2021™	10.07	8.2	-18.8
E12	60	Polyglycerol-10-mono-oleate / SMA 2021™	10.07	6.4	-36.2
F1	61	Triton X-100™ / SMA 2021™	10.07	7.2	-28.4
F2	62	Triton X-45™ / SMA 2021™	10.07	7	-30.7
F3	63	Tween 40™ / SMA 2021™	10.07	7.8	-23
F4	64	Tween 60™ / SMA 2021™	10.07	6.6	-34.4
F5	65	Tween 80™ / SMA 2021™	10.07	7	-30.9
F6	66	Tween 85™ / SMA 2021™	10.07	7.1	-29.3
F7	67	Apo 10 / SMA 3000™	10.07	10	-1
F8	68	Apo 12 / SMA 3000™	10.07	11	8.9

Table 19. Zinc Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Zn2+ Added [ppm]	Measured [ppm]	% Difference
F9	69	Coco glucoside / SMA 3000™	10.07	10.2	1
F10	70	Genapol X080 / SMA 3000™	10.07	10.6	5.5
F11	71	HEGMDDE-H370 / SMA 3000™	10.07	10.5	4.6
F12	72	HEGMDE-H360 / SMA 3000™	10.07	11.5	14.5
G1	73	HXEGMDDE-H375 / SMA 3000™	10.07	9.9	-1.2
G2	74	DDDM-MNA-C12 / SMA 3000™	10.07	11.1	10
G3	75	pGBTM-MNA-C13 / SMA 3000™	10.07	10.3	2.2
G4	76	pGBTEM-MNA-C14 / SMA 3000™	10.07	10.4	3
G5	77	nDDDT-D342 / SMA 3000™	10.07	9.6	-4.8
G6	78	nDDTGP-D323 / SMA 3000™	10.07	10.4	3.3
G7	79	OEGMDE-O330 / SMA 3000™	10.07	10.7	6.1
G8	80	Polyglycerol-10-mono-laurate / SMA 3000™	10.07	10.8	7
G9	81	Polyglycerol-10-mono-oleate / SMA 3000™	10.07	10.2	1.2
G10	82	Polyglycerol-10-mono-palmitate / SMA 3000™	10.07	6.4	-36.5
G11	83	Span 20™ / SMA 3000™	10.07	12.7	26.1
G12	84	Triton X-100™ / SMA 3000™	10.07	6.6	-34.7
H1	85	Triton X-45™ / SMA 3000™	10.07	11.2	10.8
H2	86	Tween 65™ / SMA 3000™	10.07	5.8	-42.6
H3	87	Tween 85™ / SMA 3000™	10.07	9	-10.2
H4	88	Apo 12 / SMA-IBE	10.07	6.9	-31.9
H5	89	Coco glucoside / SMA-IBE	10.07	7	-30.4
H6	90	HEGMDDE-H370 / SMA-IBE	10.07	7.2	-52.1

Table 19. Zinc Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Zn ²⁺ Added [ppm]	Measured [ppm]	% Difference
H7	91	HEGMDE-H360 / SMA-IBE	10.07	6.2	-38.1
H8	92	HXEGMDDE-H375 / SMA-IBE	10.07	7.5	-25.6
H9	93	nDDDT-D342 / SMA-IBE	10.07	7.4	-26.8
H10	94	nDDTP-D335 / SMA-IBE	10.07	6.5	-35.2
H11	95	Polyglycerol-10-mono-oleate / SMA-IBE	10.07	7.9	-21.3
H12	96	Triton X-45™ / SMA-IBE	10.07	8.5	-15.5
A1	97	Tween 85™ / SMA-IBE	10.07	7	-30.2
A2	98	Genapol X080 / SMA 725™	10.07	4.6	-53.9
A3	99	nDDDT-D342 / SMA 725™	10.07	6.9	-31.9
A4	100	nDDTGP-D323/ SMA 725™	10.07	4.2	-58.2
A5	101	Polyglycerol-10-mono-oleate / SMA 725™	10.07	5.1	-49.3
A6	102	Span 20™ / SMA 725™	10.07	4.8	-52.8
A7	103	HEGMDDE-H370 / SMA 725™	10.07	7.1	-29.3
A8	104	HEGMDE-H360 / SMA 725™	10.07	7.1	-29.5
A9	105	HXEGMDDE-H375 / SMA 725™	10.07	8.2	-18.3
A10	106	pGBTEM-MNA-C14 / SMA 725™	10.07	7.7	-23.5
A11	107	Polyglycerol-10-mono-palmitate / SMA 725™	10.07	7.3	-27.2
A12	108	Triton X-100™/ SMA 725™	10.07	6.7	-33.4
B1	109	Triton X-45™ / SMA 725™	10.07	6.8	-32.5
B2	110	Tween 65™ / SMA 725™	10.07	6.8	-32.5
B3	111	Tween 80™ / SMA 725™	10.07	7.2	-28.1
B4	112	Apo 10 / SMA Cumene	10.07	5.4	-46.7
B5	113	Apo 12 / SMA Cumene	10.07	6	-40.8
B6	114	PFDMP-O310F / SMA Cumene	10.07	6.1	-39.1
B7	115	Genapol X080 / SMA Cumene	10.07	5.3	-47.7
B8	116	HEGMDDE-H370 / SMA Cumene	10.07	5.8	-42.1

Table 19. Zinc Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Non-ionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Zn2+ Added [ppm]	Measured [ppm]	% Difference
B9	117	HEGMDE-H360 / SMA Cumene	10.07	5.4	-46.1
B10	118	HXEGMDDE-H375 / SMA Cumene	10.07	4.9	-51.8
B11	119	DDDM-MNA-C12 / SMA Cumene	10.07	5.6	-44
B12	120	pGBTM-MNA-C13/ SMA Cumene	10.07	6.1	-39.2
C1	121	nDDDTTP-D342 / SMA Cumene	10.07	5.6	-44.6
C2	122	nDDTGP-D323 / SMA Cumene	10.07	5.2	-48
C3	123	nDDTP-D335 / SMA Cumene	10.07	5.9	-41.8
C4	124	OEGMDE-O330 / SMA Cumene	10.07	7.1	-29.7
C5	125	Polyglycerol-10-mono-laurate / SMA Cumene	10.07	6.4	-36.8
C6	126	Polyglycerol-10-mono-oleate / SMA Cumene	10.07	5.7	-43.7
C7	127	Span 20™ / SMA Cumene	10.07	4.6	-54.2
C8	128	Triton X-100™ / SMA Cumene	10.07	5.4	-46.2
C9	129	Triton X-45™ / SMA Cumene	10.07	8.4	-16.7
C10	130	Tween 65™ / SMA Cumene	10.07	7.3	-27.4
C11	131	Tween 85™ / SMA Cumene	10.07	6.8	-32.4

Table 20. Zinc Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Anionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Zn2+ Added [ppm]	Measured [ppm]	% Difference
A1	1	Stepan Bio-Soft D-40™ / DIBMA	10.07	10.3	2.2
A2	2	Lakeland PAE 136™ / DIBMA	10.07	10.5	4.1
A3	3	Stepan Lathanol LAL™ / DIBMA	10.07	10.2	1.2
A4	4	n-Decanoyl-L-threonine / DIBMA	10.07	10.9	8.1

Table 20. Zinc Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Anionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Zn2+ Added [ppm]	Measured [ppm]	% Difference
A5	5	n-Dodecanoyl-L-threonine / DIBMA	10.07	10.5	3.8
A6	6	n-Decanoyl-L-methionine / DIBMA	10.07	9.6	-4.6
A7	7	n-Dodecanoyl-L-methionine / DIBMA	10.07	10.6	5.5
A8	8	Ninate 411™ / DIBMA	10.07	10.7	6
A9	9	n-Octanoyl-L-methionine / DIBMA	10.07	10.8	7.1
A10	10	Stepan Mild L3™ / DIBMA	10.07	11	9.5
A11	11	Stepan Bio-Terge AS-40™ / SMA 2000™	10.07	7.8	-22.7
A12	12	Stepan Bio-Soft D-40™ / SMA 2000™	10.07	8	-20.5
B1	13	Dodecane-1-sulfonic acid sodium salt / SMA 2000™	10.07	7.7	-23.2
B2	14	Lakeland PAE 136™ / SMA 2000™	10.07	7.9	-21.4
B3	15	Stepan Lathanol LAL™ / SMA 2000™	10.07	7.3	-27.1
B4	16	n-Decanoyl-L-methionine / SMA 2000™	10.07	7.6	-24.1
B5	17	n-Dodecanoyl-L-methionine / SMA 2000™	10.07	8	-20.8
B6	18	n-Decanoyl-L-threonine / SMA 2000™	10.07	7.2	-28.7
B7	19	n-Dodecanoyl-L-histidine / SMA 2000™	10.07	3.6	-64.1
B8	20	Stepan Mild L3™ / SMA 2000™	10.07	8.2	-18.8
B9	21	n-Dodecanoyl-L-threonine / SMA 2000™	10.07	8.2	-18.9
B10	22	Sodium dodecylbenzene sulfonate / SMA 2000™	10.07	7.3	-27.6
B11	23	Ninate 411™ / SMA 2000™	10.07	8.2	-18.2
B12	24	Stepan SLL-FB™ / SMA 2000™	10.07	7.5	-25.9
C1	25	Stepan Sulfonic Acid™ / SMA 2000™	10.07	8.1	-20
C2	26	Dodecane-1-sulfonic acid sodium salt / SMA 2021™	10.07	7.3	-27.9

Table 20. Zinc Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Anionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Zn ²⁺ Added [ppm]	Measured [ppm]	% Difference
C3	27	Lakeland PAE 136™ / SMA 2021™	10.07	6.4	-36.5
C4	28	Stepan Lathanol LAL™ / SMA 2021™	10.07	7.3	-27.6
C5	29	n-Decanoyl-L-methionine / SMA 2021™	10.07	7.2	-28.4
C6	30	Cedepal TD-403 MFLD™ / SMA 2021™	10.07	7.1	-29
C7	31	Stepan Mild L3™ / SMA 2021™	10.07	7.7	-23.6
C8	32	n-Dodecanoyl-L-methionine / SMA 2021™	10.07	11.3	12.6
C9	33	n-Decanoyl-L-methionine / SMA 3000™	10.07	11.8	16.9
C10	34	n-Dodecanoyl-L-methionine / SMA 3000™	10.07	10.9	8.5
C11	35	n-Decanoyl-L-threonine / SMA 2021™	10.07	7.6	-24.4
C12	36	n-Dodecanoyl-L-threonine / SMA 2021™	10.07	8.1	-19.7
D1	37	Sodium dodecylbenzene sulfonate / SMA 2021™	10.07	7.7	-23.8
D2	38	Ninate 411™ / SMA 2021™	10.07	8.3	-17.5
D3	39	Stepan SLL-FB™ / SMA 2021™	10.07	8.6	-14.1
D4	40	Stepan Sulfonic Acid™ / SMA 2021™	10.07	8.7	-13.9
D5	41	Lakeland PAE 136™/ SMA 3000™	10.07	11.1	9.8
D6	42	n-Dodecanoyl-L-threonine / SMA 3000™	10.07	10.5	4.5
D7	43	Ninate 411™ / SMA 3000™	10.07	11.4	12.8
D8	44	Stepan Mild L3™ / SMA 3000™	10.07	11.4	12.9
D9	45	Stepan SLL-FB™ / SMA 3000™	10.07	11	8.7
D10	46	Stepan Sulfonic Acid™ / SMA 3000™	10.07	11.9	17.8
D11	47	n-Decanoyl-L-methionine / SMA-IBE	10.07	9.2	-8.3
D12	48	n-Dodecanoyl-L-methionine/ SMA-IBE	10.07	8.2	-19

Table 20. Zinc Removal (40uL Nanonet Library Stock Solution Was Used) with Nanonets with Anionic Surfactants					
Well No.	Library No.	Nanonet Library Item	Zn2+ Added [ppm]	Measured [ppm]	% Difference
E1	49	Stepan Mild L3™ / SMA-IBE	10.07	7.1	-29.6
E2	50	Stepan SLL-FB™ / SMA-IBE	10.07	6.2	-37.9
E3	51	Amin GCK30H / SMA 725™	10.07	5.7	-43.8
E4	52	Stepan Bio-Terge AS-40™ / SMA 725™	10.07	6.7	-33.5
E5	53	n-Dodecanoyl-L-histidine / SMA 725™	10.07	3.4	-66.6
E6	54	Stepan Lathanol LAL™ / SMA 725™	10.07	6.6	-34.3
E7	55	n-Decanoyl-L-methionine / SMA 725™	10.07	6.8	-32.7
E8	56	n-Dodecanoyl-L-methionine / SMA 725™	10.07	5.8	-42.2
E9	57	Dodecane-1-sulfonic acid sodium salt / SMA 725™	10.07	5.7	-43.5
E10	58	Lakeland PAE 136™ / SMA 725™	10.07	5.9	-41.8
E11	59	n-Dodecanoyl-L-threonine / SMA 725™	10.07	6.4	-36.7
E12	60	Stepan Mild L3™ / SMA 725™	10.07	7.9	-22
F1	61	Stepan SLL-FB™ / SMA 725™	10.07	5.7	-43
F2	62	Lakeland PAE136 / SMA Cumene	10.07	5.9	-41.3
F3	63	n-Dodecanoyl-L-threonine / SMA Cumene	10.07	5.9	-41
F4	64	Stepan Mild L™3 / SMA Cumene	10.07	6.1	-39.1
F5	65	Stepan SLL-FB™ / SMA Cumene	10.07	5.5	-45
F6	66	n-Decanoyl-L-methionine / SMA Cumene	10.07	6.4	-36.2
F7	67	n-Dodecanoyl-L-methionine / SMA Cumene	10.07	7.2	-28.6
F8	68	n-Dodecanoyl-L-histidine / SMA Cumene	10.07	5.5	-45.4

Example 23

Nanonet formation utilizing a derivatized SMA polymer.

To demonstrate the alternative hydrophilic functional groups can be utilized to make nanonets, SMA-2000™ was derivatized by reacting the anhydride

with ethanolamine. The resulting polymer was able to complex with all three Triton series surfactants (TX-100™, TX-305™, TX-45™), confirming that derivatized polymers with alternative hydrophilic functional groups can also be utilized to form nanonets.

Triton™ X series were titrated at a concentration range of 1000-0ppm with Derivatized SMA 2000™ polymer at a concentration of 200ppm; nanonet activated with 10uL of HCl

Table 21. Nanonet Formation Efficiency Test with Triton™ Series Surfactants and SMA-725™, SMA-2000™				
Surfactant	Polymer [400PPM]	Concentration [PPM]	Nanonet Absorbance [280 nm]	Control Absorbance [280 nm]
Triton X100™	SMA 2000™	1000	0.2253	1.1656
Triton X100™	SMA 2000™	500	0.1728	0.5988
Triton X100™	SMA 2000™	250	0.1571	0.3206
Triton X100™	SMA 2000™	100	0.0768	0.1449
Triton X100™	SMA 2000™	64.8	0.0773	0.1257
Triton X100™	SMA 2000™	0	0.0619	0.0516
Triton X100™	SMA 725™	1000	1.2527	1.1345
Triton X100™	SMA 725™	500	1.549	0.5752
Triton X100™	SMA 725™	295.16	0.172	0.3518
Triton X100™	SMA 725™	250	0.1353	0.302
Triton X100™	SMA 725™	100	0.0698	0.1394
Triton X100™	SMA 725™	0	0.0511	0.1441
Triton X45™	SMA 2000™	1000	0.1186	1.4812
Triton X45™	SMA 2000™	616.18	0.0973	0.8388
Triton X45™	SMA 2000™	500	0.1049	0.7703
Triton X45™	SMA 2000™	250	0.0783	0.4892
Triton X45™	SMA 2000™	100	0.0654	0.1734
Triton X45™	SMA 2000™	0	0.0707	0.0518
Triton X45™	SMA 725™	1000	0.1837	1.6766
Triton X45™	SMA 725™	500	0.128	1.3428
Triton X45™	SMA 725™	250	0.1147	0.785
Triton X45™	SMA 725™	100	0.0703	0.3648
Triton X45™	SMA 725™	92.92	0.1003	0.3679
Triton X45™	SMA 725™	0	0.0552	0.0502

Table 22. Nanonet Formation with Triton™ Series Surfactant and Ethanolamine Derived SMA-2000™			
Surfactant	Concentration Surfactant [ppm]	Control (Absorbance at 540 nm)	Ethanolamine derived SMA-2000™ (Absorbance at 540 nm)
Triton X-100™	1000	0.0474	0.0689
Triton X-100™	500	0.0473	0.1056
Triton X-100™	250	0.0471	0.1909
Triton X-100™	125	0.048	0.1271
Triton X-100™	50	0.0474	0.0843
Triton X-100™	0	0.0406	0.0616
Triton X-305™	1000	0.0484	0.0493
Triton X-305™	500	0.0475	0.076
Triton X-305™	250	0.0467	0.0843
Triton X-305™	125	0.0462	0.0746
Triton X-305™	50	0.0444	0.0716
Triton X-305™	0	0.0408	0.0664
Triton X-45™	1000	0.6694	0.7939
Triton X-45™	500	0.309	0.8672
Triton X-45™	250	0.1338	0.9615
Triton X-45™	125	0.0951	1.0042
Triton X-45™	50	0.0517	0.116
Triton X-45™	0	0.0388	0.0636

Example 24

Effect of polymer on settling speed of nanonets floc.

Experimental protocol

3 mL of a 4000 ppm Ca^{2+} solution made from calcium chloride (83%-87%, Downflake) is added to an Eppendorf vial.

Table 23: Information on Polymers Used in This Set of Experiments.		
Polymer Name	Molecular Weight [kDa]	Stryene to Maleic Anhydride Ratio
SMA-725™	130	3:1
SMA-2000™	7.5	2:1
SMA-3000™	9.5	3:1

The polymer-surfactant nanonet complex is made by adding a 2% polymer solution to a 1% surfactant solution at a 2.5:1 polymer-to-surfactant ratio. To bind the nanonets to the free calcium ions, 100 uL of the polymer-surfactant mixture (97 ppm surfactant, 473 ppm polymer) is added to the calcium solution, after which clear white flocs can be seen in solution.

To test the effect of adding polymer alone, 71 uL of the 2% polymer solution (473 ppm polymer) is added to the calcium solution.

The mixture is shaken at 600 RPM (light shaking) for one minute. Light shaking ensures that the floc-break up is minimized during mixing, as this would increase the time required for batch settling.

The quartz cuvette use to contain the sample must be cleaned five times with deionized (DI) water and hand dried with a kimwipe until the outer faces are dry and optically clean.

After one minute of mixing, the flocculated suspension is immediately transferred to a clean quartz cuvette, after which the time lapse is started. Videos are taken at a frequency of 0.2 Hz for 25 minutes.

Calculation of batch settling parameters

A Python script was written to track the settling of the flocs. The region of interest (ROI) was taken as the area of the floc bed after 25 minutes of sedimentation. To do this, a box is manually selected spanning the inner walls of the cuvette and the bottom of the meniscus.

From here, an image processing routine automatically extracts the position of the interface between the clear liquid phase and the consolidated bed after sedimentation. The consolidated bed height can be calculated and a new ROI of the sedimentation region can be generated for subsequent analysis.

Within this new ROI, an average grayscale pixel intensity can be calculated over time; the flocs are white in color (in contrast to a dark black-colored background) so it is expected that this value will increase during sedimentation. This value can be interpreted as the amount of floc sediment at the bottom of the cuvette.

To output a single, quantifiable parameter of interest from a series of images tracking sedimentation, a sigmoid function is fitted to the apparent floc sediment data.

An example of this curve fitting of the apparent sedimentation is illustrated in Figure 26.

After curve fitting, a t_{50} value (center of sigmoid) can be reported which serves as an indicator of the settling speed. Depending on the nature of phase separation, this may also serve as an indicator of floc size (faster settling = lower t_{50}).

Results

Polymer selection directly influences settling speed

A graph of the t_{50} for the three polymers is illustrated in the Figure 27.

It is evident that the speed of sedimentation is directly impacted by the selection of polymer. SMA-725™ has the highest molecular weight and it is expected that this would settle the fastest by this rationale.

There is a difference in settling speed between the different polymers SMA-725™ and SMA-3000™. In general, the longer polymer settles significantly faster than the shorter polymers (SMA-725™ > SMA-2000™/3000™). However, SMA-2000™ settles faster than SMA-3000™ in these experiments, despite SMA-2000™ being a slightly shorter polymer. This lack of correlation between polymer length and settling speed suggests that there are additional polymer properties that can influence the phase separation behavior of the nanonets beyond polymer length. This assay suggests that efficient nanonet design requires more than just a large surfactant aggregate and small polymer; polymer size and chemical identity will also significantly effect nanonet removal in wastewater treatment processes.

Example 25

Preparation and characterization of Nanonet Fe:

100L of a 1.5M stock solution of KOH was heated to 95°C. To this solution, 10kg of pre-ground SMA 725 was added with vigorous mixing. The

solution was left for 3 hours to hydrolyze, at which point 4kg of oleic acid was pumped into the mixture. The solution was left to mix at 95°C. After 1 hour, 10L of a pre-heated 20% polyglycerol-10-stearate solution was added to the mixture. The solution was allowed to cool to 60°C while continuing mixing, at which point the solution was pumped through a 1 micron filter to remove any non-hydrolyzed polymer. Addition of distilled or deionized water was then added to the solution to compensate for any evaporative loss.

11 batches of nanonet Fe were prepared as described and each batch was measured for particle diameter and viscosity. The nanonet batches were recorded in Table 24 below. Particle diameter was measured by dynamic light scattering on a Malvern NanoZS and calculated from intensity values. Viscosity was measured on a Brookfield VII rheometer at 23°C.

Table 24: Characteristics of a 10% nanonet Fe solution		
Batch #	Size (nm)	Viscosity (Cp)
1	10.15	14
2	9.89	18
3	11.31	18
4	11.99	18
5	10.9	20
6	6.67	18
7	13.02	12
8	8.89	16
9	8.72	16
10	9.12	14
11	10.15	20

Example 26

Treatment of Produced water with NanoNet Fe in a water treatment process

In the following example, produced water sourced from the Permian Basin in Texas, United States was treated with nanonet Fe, Batch 7, from Example 25.

Incoming produced water was pumped at a 4GPM flow rate from 1000L totes through a 484 Mazzei venturi eductor. 50ppm nanonets were dosed into the eductor, along with concurrent addition of air at approximately 3 L/min. The water traveled through a reaction tank, then into a flocculation loop. The water then entered a dissolved air flotation unit, where nanonet-iron complexes floated to the surface and were removed from the water. The iron levels, turbidity, total dissolved solids, oxidative reductive potential, and alkalinity displayed in the Figures 28A-28E.

Example 27

Treatment of Produced water with NanoNet Fe in a water treatment process with addition of anionic flocculant.

Incoming produced water was pumped at an 8 GPM flow rate through a 784 Mazzei venturi eductor. 30ppm nanonet Fe, Batch 11, from Example 25 was actively dosed using a peristaltic pump into an venturi eductor. The treated fluid then flowed into a mixing tank, which was weakly aerated by a recycle sparge coming from the outflow of the treatment train. To aid capture of iron, the mixing tank was rapidly mixed using a dynamic, high shear mixer. The treated fluid exited the mixing tank, at which point it was dosed with 1 ppm of an anionic flocculant. The treated fluid then passed through a series of high speed and low speed mixing chambers to enhance unfolding of the flocculant and flocculation of the nanonet-iron complexes. The treated fluid subsequently entered a floatation chamber and flocs floated to the surface by addition of dissolved air. The final effluent exits the DAF through a series of weirs and is pumped through a final polishing filter with average pore size of 20 micron. The incoming and outgoing iron and bacterial levels from treating water with this process are displayed in the Tables 25 and 26 below.

Table 25: Iron concentrations (ppm) after treatment of produced water with nanonets.								
	Day 1		Day 2		Day 3		Day 4	
	Ave.	Std. Dev.	Ave.	Std. Dev.	Ave.	Std. Dev.	Ave.	Std. Dev.
Influent	14.4	0.53	18.2	2.70	33.48	10.38	36.08	12.07
Post-DAF	0.67	0.10	1.49	1.50	0.99	0.50	2.62	1.62
Post-Filter	0.24	0.42	0.1	0.17	0	0	0.42	0.67

Table 26: ATP levels (pg/mL) of produced water before and after treatment with nanonet Fe in a dissolved air flotation process treatment train						
	Trial 1			Trial 2		
Influent	2697			3724		
Post-DAF	111	137	449	330	100	86
Post-Filter	130	10	259	129	27	25

Example 28

Reduction in oxidative demand through addition of nanonet Fe.

The overall demand for oxidative chemicals in the treatment of water in oilfield operations is likely to be lowered by the removal of ferrous iron from solution. . To test this hypothesis, a solution of ferrous iron was treated both with and without nanonets, followed by sodium chlorite. The solutions were filtered and the filtrate then dosed with a fixed amount of chlorine dioxide. Increasing amounts of nanonets lead to a substantial increase in oxidant residual, demonstrating removal of oxidative demand in the system.

Figures 29A and 29B show side by side comparison of iron removal with Nanonets, repeated. Produced water supplemented with 50ppm Fe^{2+} , was first treated with NaClO_2 at the indicated concentration, then sequentially treated with the indicated dose of Nanonet Fe (0, 10, 20, 40 ppm as increasingly lighter bars). The solutions were mixed by inversion and allowed to sit for 5 minutes at room temperature (no aeration). The resulting solutions were gravity filtered (10

micron cellulose filter), and a sample of the resulting filtrate was analyzed for iron content by ferrozine assay (A). Subsequently the filtrate was treated with 25ppm ClO_2 and left to incubate for 5 minutes, after which time the residual ClO_2 was measured (B).

Although various embodiments of the invention are disclosed herein, many adaptations and modifications may be made within the scope of the invention in accordance with the common general knowledge of those skilled in this art. Such modifications include the substitution of known equivalents for any aspect of the invention in order to achieve the same result in substantially the same way. Numeric ranges are inclusive of the numbers defining the range. Furthermore, numeric ranges are provided so that the range of values is recited in addition to the individual values within the recited range being specifically recited in the absence of the range. The word "comprising" is used herein as an open-ended term, substantially equivalent to the phrase "including, but not limited to", and the word "comprises" has a corresponding meaning. As used herein, the singular forms "a", "an" and "the" include plural references unless the context clearly dictates otherwise. Thus, for example, reference to "a thing" includes more than one such thing. Citation of references herein is not an admission that such references are prior art to the present invention. Furthermore, material appearing in the background section of the specification is not an admission that such material is prior art to the invention. Any priority document(s) are incorporated herein by reference as if each individual priority document were specifically and individually indicated to be incorporated by reference herein and as though fully set forth herein. The invention includes all embodiments and variations substantially as hereinbefore described and with reference to the examples and drawings.

What is claimed is:

1. A kit for sequestering non-water moieties from an aqueous solution, the kit comprising:

a) a surfactant aggregate having an average aggregate diameter; and

b) a polymer having an average particle diameter which average particle diameter is the same or smaller than the average aggregate diameter; and

c) a precipitation agent,

provided that when the surfactant is selected from the group consisting of: Triton X-100TM, Triton X-305TM, N-dodecyl-beta-D-maltoside (DDM), sodium oleate, sodium cholate, sodium deoxycholate (DOC), beta-octyl glucoside (Beta-OG), lauryl dimethylamine n-oxide (LDAO), then the polymer is not selected from the group consisting of: SMA-2000TM, SMA-2021TM, and Di-isobutylene-co-maleic acid (DIBMA).

2. The kit of claim 1 wherein the precipitation agent is selected from the group consisting of: a cation generator, a cation, an acid, a divalent cation, a trivalent cation and mixtures thereof.

3. The kit of claim 1 wherein the precipitation agent is selected from the group consisting of: acetic acid, malic acid, citric acid, tartaric acid, fumaric acid, lactic acid, hydrochloric acid, sulfuric acid, and phosphoric acid.

4. The kit of any one of claims 1 to 3 wherein the surfactant is selected from the group consisting of: anionic surfactants, nonionic surfactants, sugar-derived surfactants, glycidyl-derived surfactants, fatty acid alcohol-derived surfactants, nonionic surfactants, saccharide polyethyleneoxide combination surfactants, saccharide ester surfactants, sulfonated sugar based surfactants, aldonamide based surfactants, amide sugar based surfactants, amino alcohol surfactants,

amino acid based surfactants, polyol surfactants, 1,2 glycol surfactants, and zwitterionic surfactants.

5. The kit of any one of claims 1 to 3 wherein the surfactant is a nonionic surfactant.

6. The kit of any one of claims 1 to 3 wherein the surfactant is an ionic surfactant.

7. The kit of any one of claims 1 to 3 wherein the surfactant is an anionic surfactant.

8. The kit of any one of claims 1 to 6 wherein the surfactant has a neutral charge.

9. The kit of any one of claims 1 to 6 wherein the surfactant is an uncharged surfactant.

10. The kit of any one of claims 1 to 9 wherein the surfactant is soluble in water.

11. The kit of any one of claims 1 to 9 wherein the surfactant is insoluble in water.

12. The kit of any one of claims 1 to 11 wherein the polymer is a long polymer.

13. The kit of any one of claims 1 to 11 wherein the polymer has a molecular weight of at least 1,500 Da.

14. The kit of any one of claims 1 to 11 wherein the polymer has a molecular weight of at least 26,000 Da.

15. The kit of any one of claims 1 to 11 wherein the polymer has a molecular weight of at least 100,000 Da.

16. The kit of any one of claims 1 to 11 wherein the polymer has a molecular weight of at least 125,000 Da.

17. The kit of any one of claims 1 to 16 wherein the polymer is an amphipathic polymer.

18. The kit of any one of claims 1 to 17 wherein the polymer comprises an alkyl chain having at least 8 carbon atoms in length.

19. The kit of any one of claims 1 to 18 wherein the polymer is a block co-polymer.

20. The kit of claim 19 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 4:1.

21. The kit of claim 20 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 3:1.

22. The kit of claim 20 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 2:1 to about 3:1.

23. The kit of claim 20 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 2:1.

24. The kit of claim 20 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 3:1.

25. The kit of claim 1 wherein the surfactant is selected from the group consisting of: octylphenol ethoxylates, nonylphenol ethoxylates, PGL1, PGL2, PGL3, PGL4, PGL5, Triton X-45TM, Triton X-100TM, Triton X-305TM, sodium oleate, LDAO, and mixtures thereof.
26. The kit of claim 1 wherein the surfactant is a mixture of PGL5 and sodium oleate.
27. The kit of claim 1 wherein the surfactant is a mixture of PGL5 and potassium oleate.
28. The kit of any one of claims 1, and 25 to 27 wherein the polymer is selected from the group consisting of: SMA-725TM, SMA-2000TM, SMA-3000TM, SMA-2021TM, polymethacrylate, DIBMA, styrene carbamate block co-polymers, and styrene maleimide block co-polymers.
29. The kit of any one of claims 1, and 25 to 27 wherein the polymer is SMA-725TM.
30. The kit of any one of claims 1 to 29 wherein at least a portion of the polymer is hydrolyzed.
31. A nanonet comprising:
- a) a surfactant aggregate having an average aggregate diameter; and
 - b) a polymer having an average particle diameter which average particle diameter is the same or smaller than the average aggregate diameter,
- provided that when the surfactant is selected from the group consisting of: Triton X-100TM, Triton X-305TM, N-dodecyl-beta-D-maltoside (DDM), sodium

oleate, sodium cholate, sodium deoxycholate (DOC), beta-octyl glucoside (Beta-OG), lauryl dimethylamine n-oxide (LDAO), then the polymer is not selected from the group consisting of: SMA-2000TM SMA-2021TM, and Di-isobutylene-co-maleic acid (DIBMA).

32. The nanonet of claim 31 wherein the nanonet has a diameter larger than the average polymer particle diameter.

33. The nanonet of claim 31 or 32 wherein the surfactant is selected from the group consisting of: anionic surfactants, nonionic surfactants, sugar-derived surfactants, glycidyl-derived surfactants, fatty acid alcohol-derived surfactants, nonionic surfactants, saccharide polyethyleneoxide combination surfactants, saccharide ester surfactants, sulfonated sugar based surfactants, aldonamide based surfactants, amide sugar based surfactants, amino alcohol surfactants, amino acid based surfactants, polyol surfactants, 1,2 Glycol surfactants, and zwitterionic surfactants.

34. The nanonet of claim 31 or 32 wherein the surfactant is a nonionic surfactant.

35. The nanonet of claim 31 or 32 wherein the surfactant is an ionic surfactant.

36. The nanonet of claim 31 or 32 wherein the surfactant is an anionic surfactant.

37. The nanonet of any one of claims 31 to 34 wherein the surfactant has a neutral charge.

38. The nanonet of any one of claims 31 to 34 wherein the surfactant is an uncharged surfactant.

39. The nanonet of any one of claims 31 to 38 wherein the surfactant is soluble in water.

40. The nanonet of any one of claims 31 to 38 wherein the surfactant is insoluble in water.

41. The nanonet of any one of claims 31 to 40 wherein the polymer is a long polymer.

42. The nanonet of any one of claims 31 to 40 wherein the polymer has a molecular weight of at least 1,500 Da.

43. The nanonet of any one of claims 31 to 40 wherein the polymer has a molecular weight of at least 26,000 Da.

44. The nanonet of any one of claims 31 to 40 wherein the polymer has a molecular weight of at least 100,000 Da.

45. The nanonet of any one of claims 31 to 40 wherein the polymer has a molecular weight of at least 125,000 Da.

46. The nanonet of any one of claims 31 to 45 wherein the polymer is an amphipathic polymer.

47. The nanonet of any one of claims 31 to 46 wherein the polymer comprises an alkyl chain having at least 8 carbon atoms in length.

48. The nanonet of any one of claims 31 to 47 wherein the polymer is a block co-polymer.

49. The nanonet of claim 48 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 4:1.
50. The nanonet of claim 48 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 3:1.
51. The nanonet of claim 48 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 2:1 to about 3:1.
52. The nanonet of claim 48 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 2:1.
53. The nanonet of claim 48 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 3:1.
54. The nanonet of claim 31 or 32 wherein the surfactant is selected from the group consisting of: octylphenol ethoxylates, nonylphenol ethoxylates, PGL1, PGL2, PGL3, PGL4, PGL5, Triton X-45TM, Triton X-100TM, Triton X-305TM, sodium oleate, LDAO, and mixtures thereof.
55. The nanonet of claim 31 or 32 wherein the surfactant is a mixture of PGL5 and sodium oleate.
56. The nanonet of claim 31 or 32 wherein the surfactant is a mixture of PGL5 and potassium oleate.
57. The nanonet of any one of claims 31, 32, and 54 to 56 wherein the polymer is selected from the group consisting of: SMA-725TM, SMA-2000TM, SMA-3000TM, SMA-2021TM, polymethacrylate, DIBMA, styrene carbamate block co-polymers, and styrene maleimide block co-polymers.

58. The nanonet of any one of claims 31, 32, and 54 to 56 wherein the polymer is SMA-725TM.

59. The nanonet of any one of claims 31 to 58 wherein at least a portion of the polymer is hydrolyzed.

60. The nanonet of any one of claims 31 to 59 further comprising a sequestered non-water moiety.

61. The nanonet of claim 60 wherein the sequestered non-water moiety is an iron moiety.

62. A method of sequestering a non-water moiety from an aqueous solution, the method comprising:

a) adding a surfactant aggregate having an average aggregate diameter to the aqueous solution comprising the non-water moiety; and

b) adding a polymer having an average particle diameter which average particle diameter is the same or smaller than the average aggregate diameter to the aqueous solution comprising the non-water moiety,

thereby forming a treated aqueous solution,

provided that when the surfactant is selected from the group consisting of: Triton X-100TM, Triton X-305TM, N-dodecyl-beta-D-maltoside (DDM), sodium oleate, sodium cholate, sodium deoxycholate (DOC), beta-octyl glucoside (Beta-OG), lauryl dimethylamine n-oxide (LDAO), then the polymer is not selected from the group consisting of: SMA-2000TM, SMA-2021TM, and Di-isobutylene-co-maleic acid (DIBMA)

63. The method of claim 62 further comprising adding a precipitation agent to the treated aqueous solution.

64. The method of claim 62 or 63 further comprising mixing.

65. The method of any one of claims 62 to 64 further comprising removal of solid particles from the treated aqueous solution.

66. The method of claim 65 wherein the removal of solid particles comprises at least one selected from the group consisting of: filtration, centrifugation, gravity separation, flotation, skimming, and electromagnetic attraction.

67. The method of any one of claims 62 to 66 wherein the surfactant is selected from the group consisting of: anionic surfactants, nonionic surfactants, sugar-derived surfactants, glycidyl-derived surfactants, fatty acid alcohol-derived surfactants, nonionic surfactants, saccharide polyethyleneoxide combination surfactant, saccharide ester surfactants, sulfonated sugar based surfactants, aldonamide based surfactants, amide sugar based surfactants, amino alcohol surfactants, amino acid based surfactants, polyol surfactants, 1,2 glycol surfactants, and zwitterionic surfactants.

68. The method of any one of claims 62 to 66 wherein the surfactant is a nonionic surfactant.

69. The method of any one of claims 62 to 66 wherein the surfactant is a anionic surfactant.

70. The method of any one of claims 62 to 66 wherein the surfactant has a neutral charge.

71. The method of any one of claims 62 to 68 wherein the surfactant is an uncharged surfactant.

72. The method of any one of claims 62 to 71 wherein the surfactant is soluble in water.

73. The method of any one of claims 62 to 71 wherein the surfactant is insoluble in water.

74. The method of any one of claims 62 to 73 wherein the polymer is a long polymer.

75. The method of any one of claims 62 to 73 wherein the polymer has a molecular weight of at least 1,500 Da.

76. The method of any one of claims 62 to 73 wherein the polymer has a molecular weight of at least 26,000 Da.

77. The method of any one of claims 62 to 73 wherein the polymer has a molecular weight of at least 100,000 Da.

78. The method of any one of claims 62 to 73 wherein the polymer has a molecular weight of at least 125,000 Da.

79. The method of any one of claims 62 to 78 wherein the polymer is an amphipathic polymer.

80. The method of any one of claims 62 to 78 wherein the polymer comprises an alkyl chain having at least 8 carbon atoms.

81. The method of any one of claims 62 to 78 wherein the polymer is a block co-polymer.

82. The method of claim 81 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 4:1.

83. The method of claim 81 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 1:1 to about 3:1.
84. The method of claim 81 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of from about 2:1 to about 3:1.
85. The method of claim 81 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 2:1.
86. The method of claim 81 wherein the polymer comprises a ratio of hydrophobic:hydrophilic groups of about 3:1.
87. The method of claim 62 wherein the surfactant is selected from the group consisting of: octylphenol ethoxylates, nonylphenol ethoxylates, PGL1, PGL2, PGL3, PGL4, PGL5, Triton X-45TM, Triton X-100TM, Triton X-305TM, sodium oleate, LDAO, and mixtures thereof.
88. The method of claim 62 wherein the surfactant is a mixture of PGL5 and sodium oleate.
89. The method of claim 62 wherein the surfactant is a mixture of PGL5 and potassium oleate.
90. The method of any one of claims 62, and 87 to 89 wherein the polymer is selected from the group consisting of: SMA-725TM, SMA-2000TM, SMA-3000TM, SMA-2021TM, polymethacrylate, DIBMA, styrene carbamate block co-polymers, and styrene maleimide block co-polymers.
91. The method of any one of claims 62, and 87 to 89 wherein the polymer is SMA-725TM.

92. The method of any one of claims 62 to 91 wherein at least a portion of the polymer is hydrolyzed.

93. A method of sequestering a non-water moiety from an aqueous solution, the method comprising adding the nanonet of any one of claims 31 to 61 to the aqueous solution comprising a non-water moiety, thereby forming a treated aqueous solution.

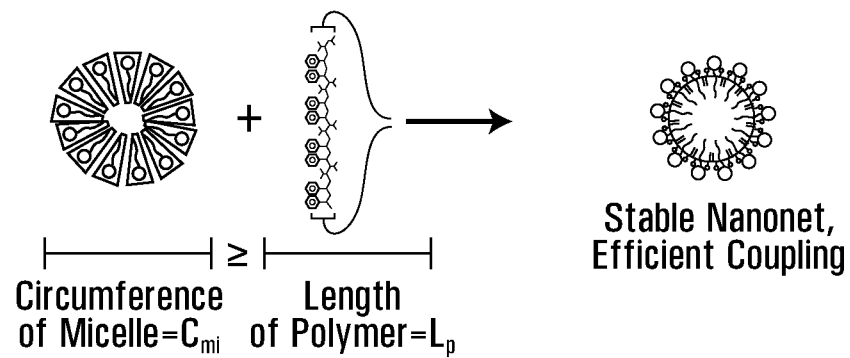
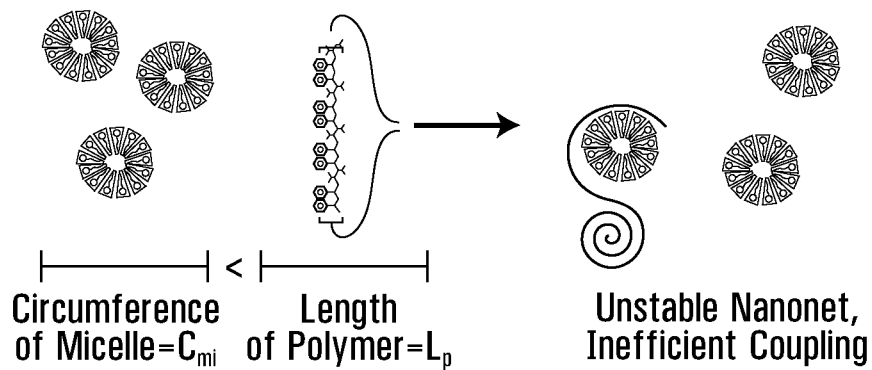
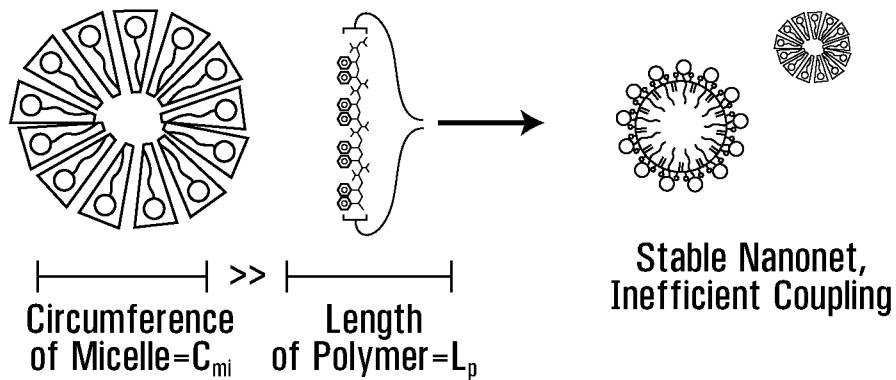
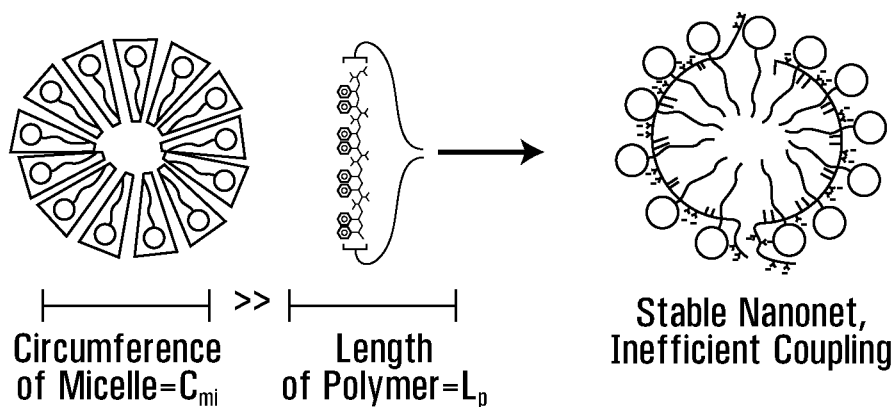
94. The method of claim 93 further comprising adding a precipitation agent to the treated aqueous solution.

95. The method of claims 93 or 94 further comprising mixing.

96. The method of any one of claims 93 to 95 further comprising removal of solid particles from the treated aqueous solution.

97. The method of claim 96 wherein the removal of solid particles comprises at least one selected from the group consisting of: filtration, centrifugation, gravity separation, flotation, skimming, and electromagnetic attraction.

1/29

**FIG. 1A****FIG. 1B****FIG. 1C****FIG. 1D**

2/29

TX-305

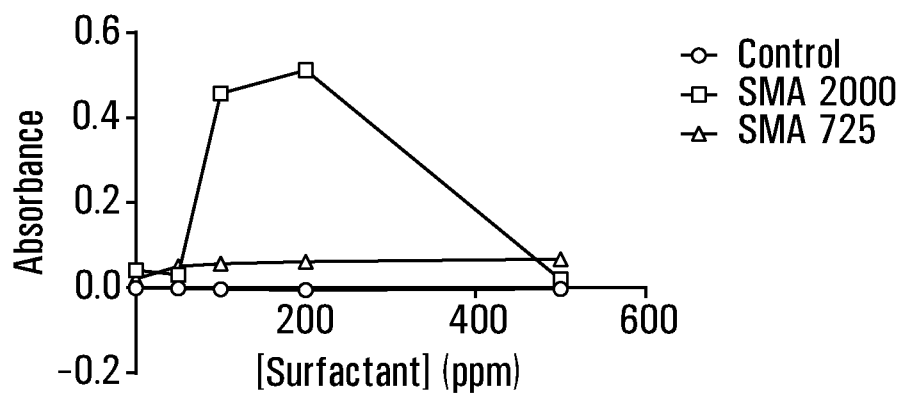


FIG. 2A

TX-100

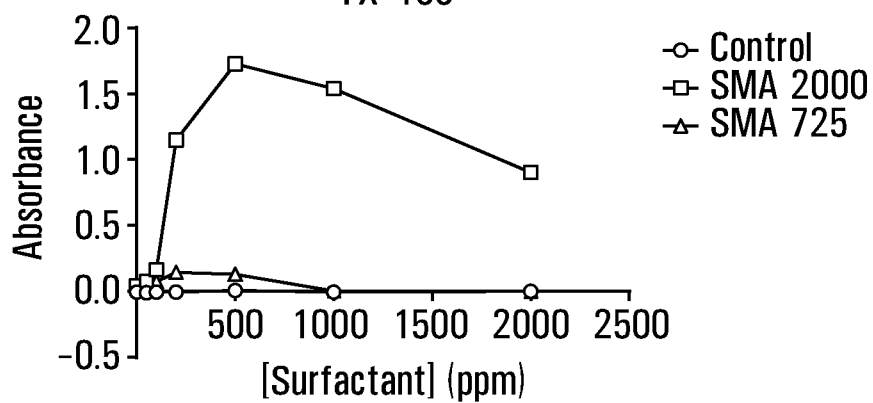


FIG. 2B

TX-45

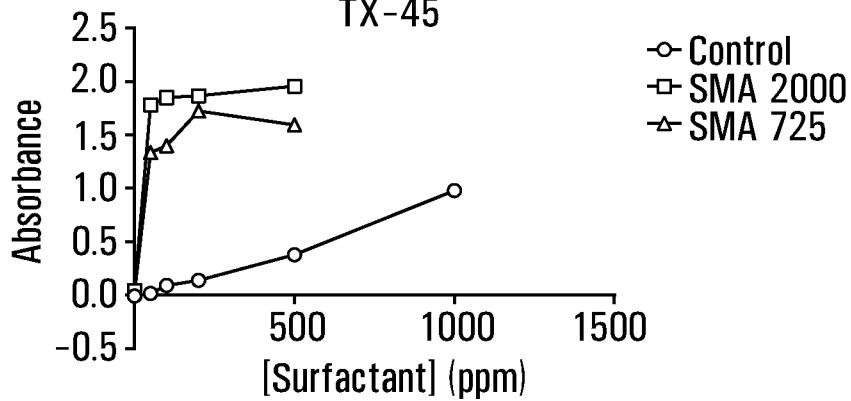


FIG. 2C

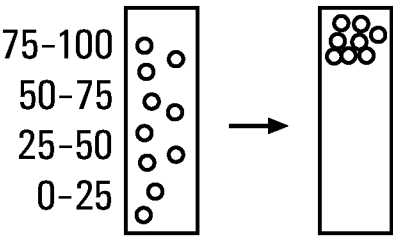


FIG. 3A

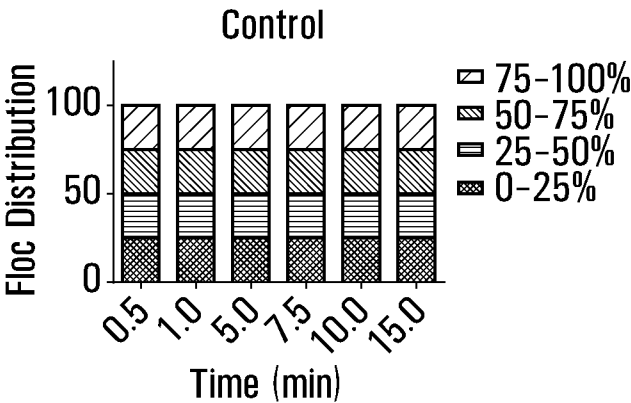


FIG. 3B

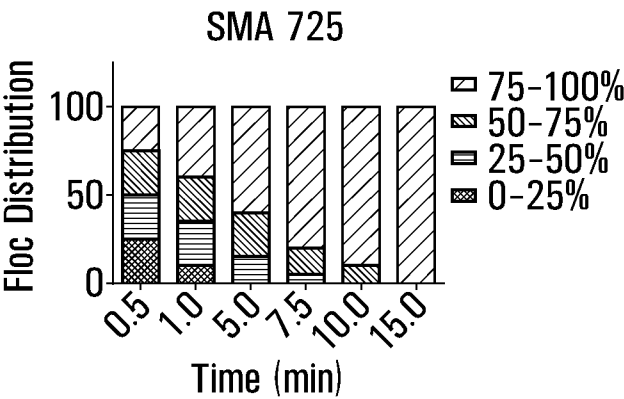


FIG. 3C

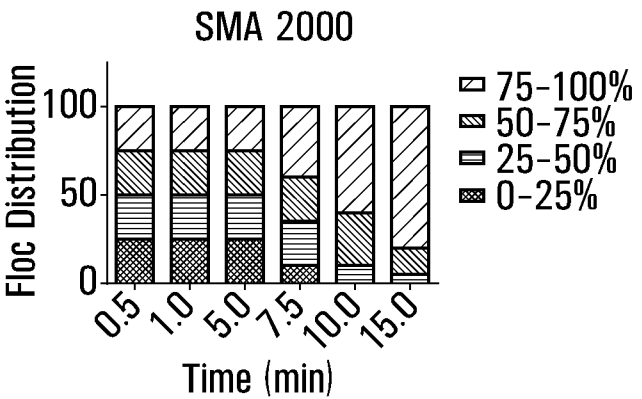


FIG. 3D

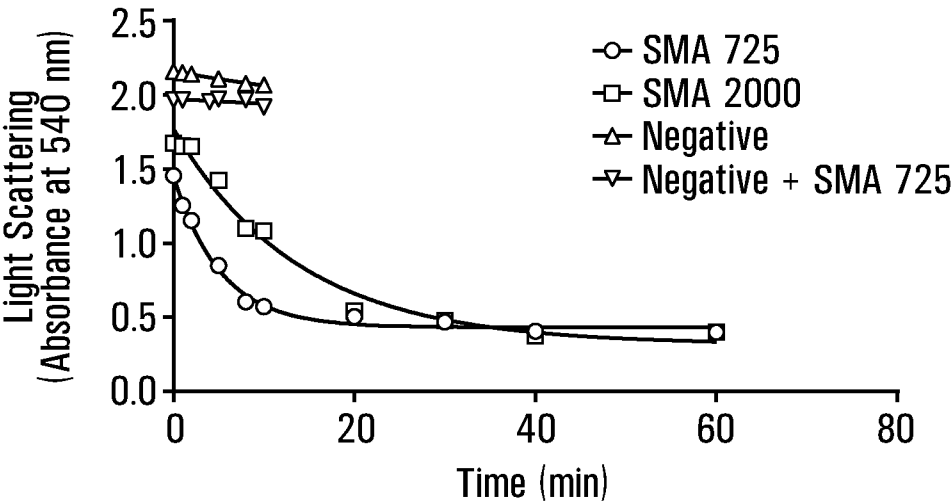
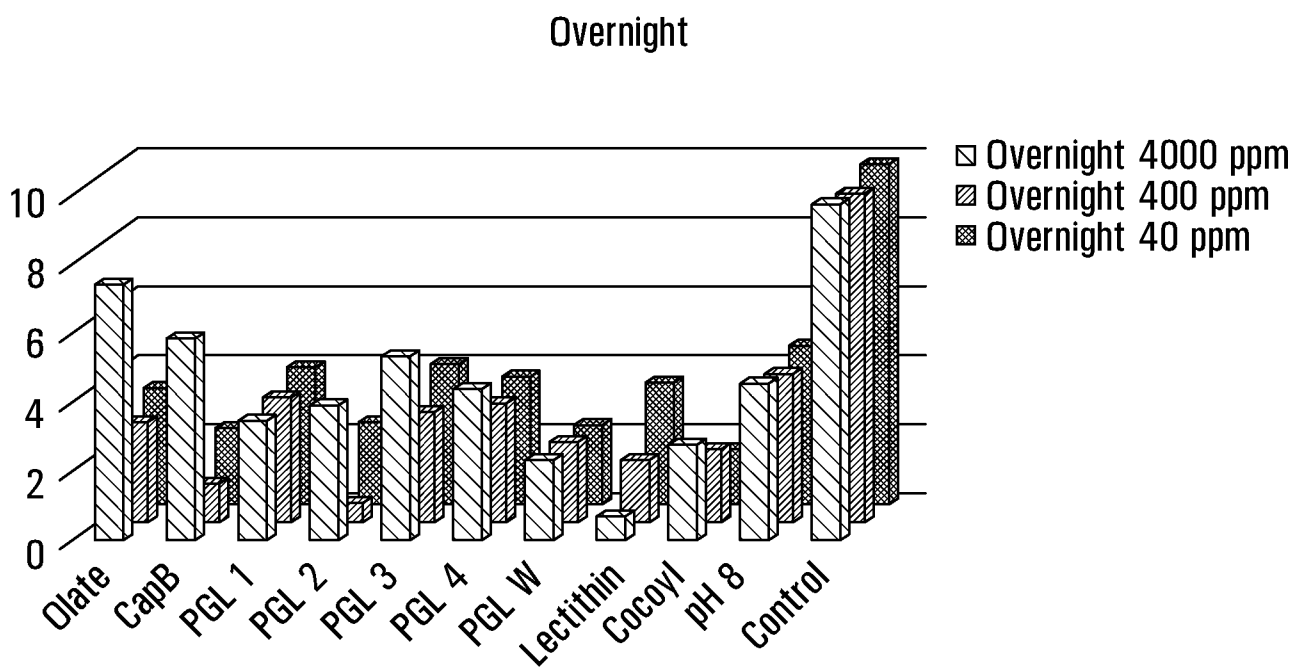
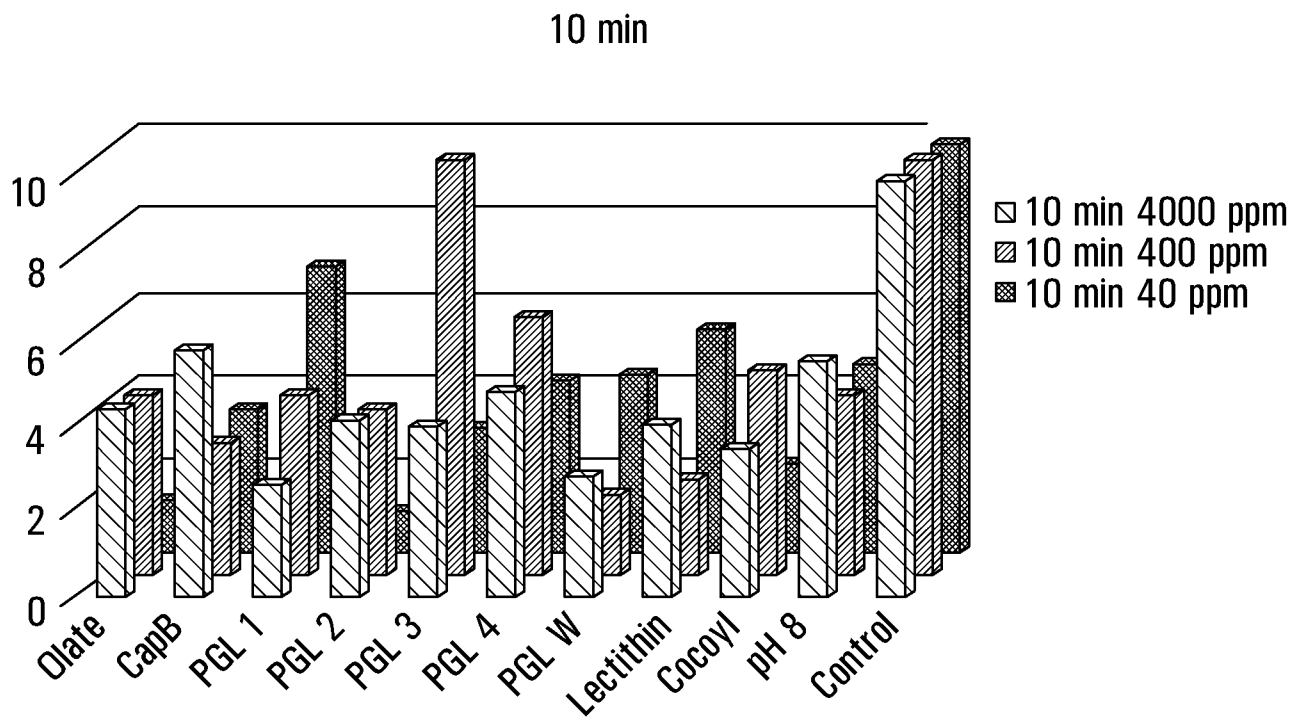


FIG. 3E

4/29

**FIG. 4**

5/29

**FIG. 5**

6/29

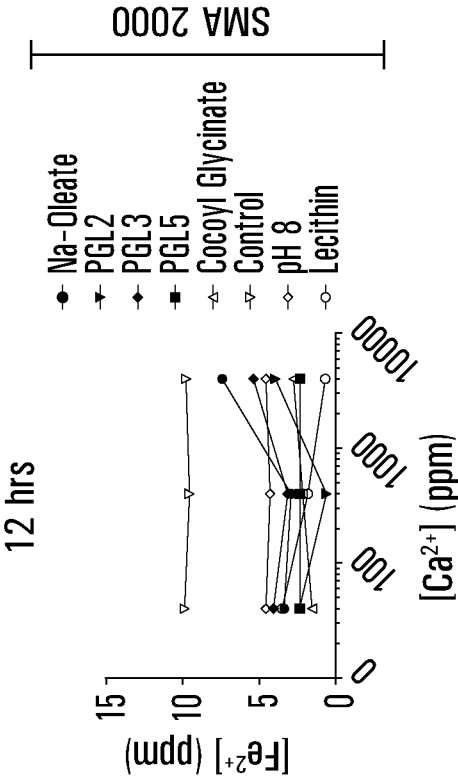


FIG. 6A

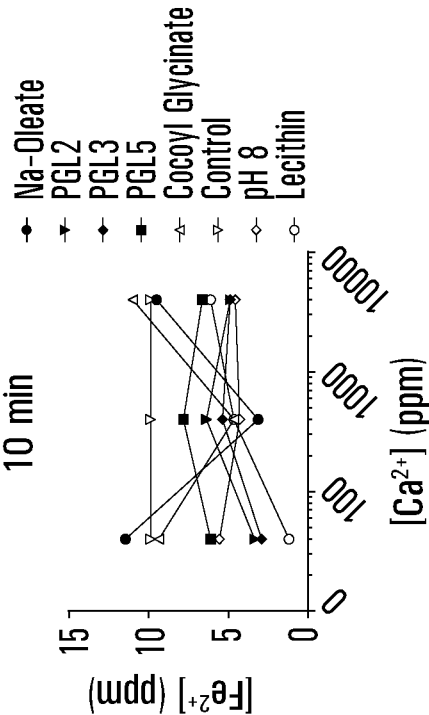


FIG. 6B

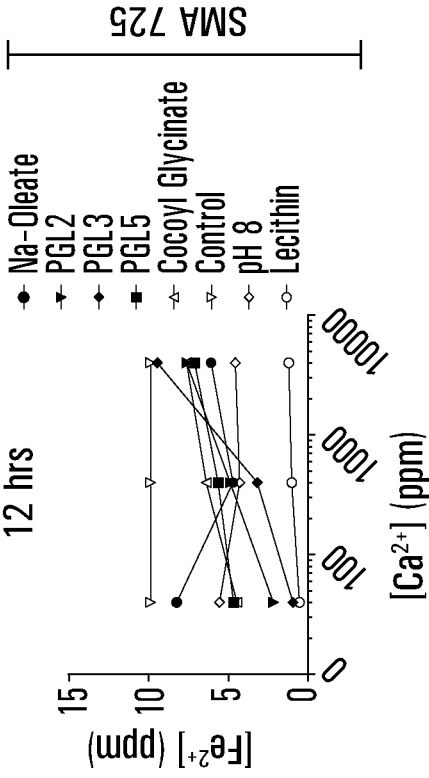


FIG. 6C

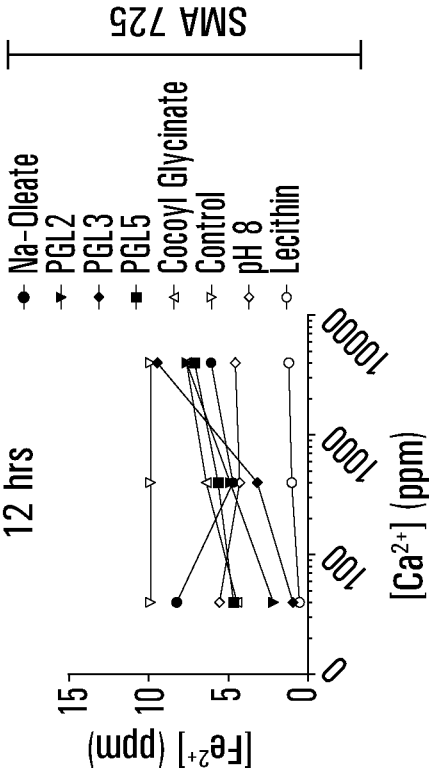


FIG. 6D

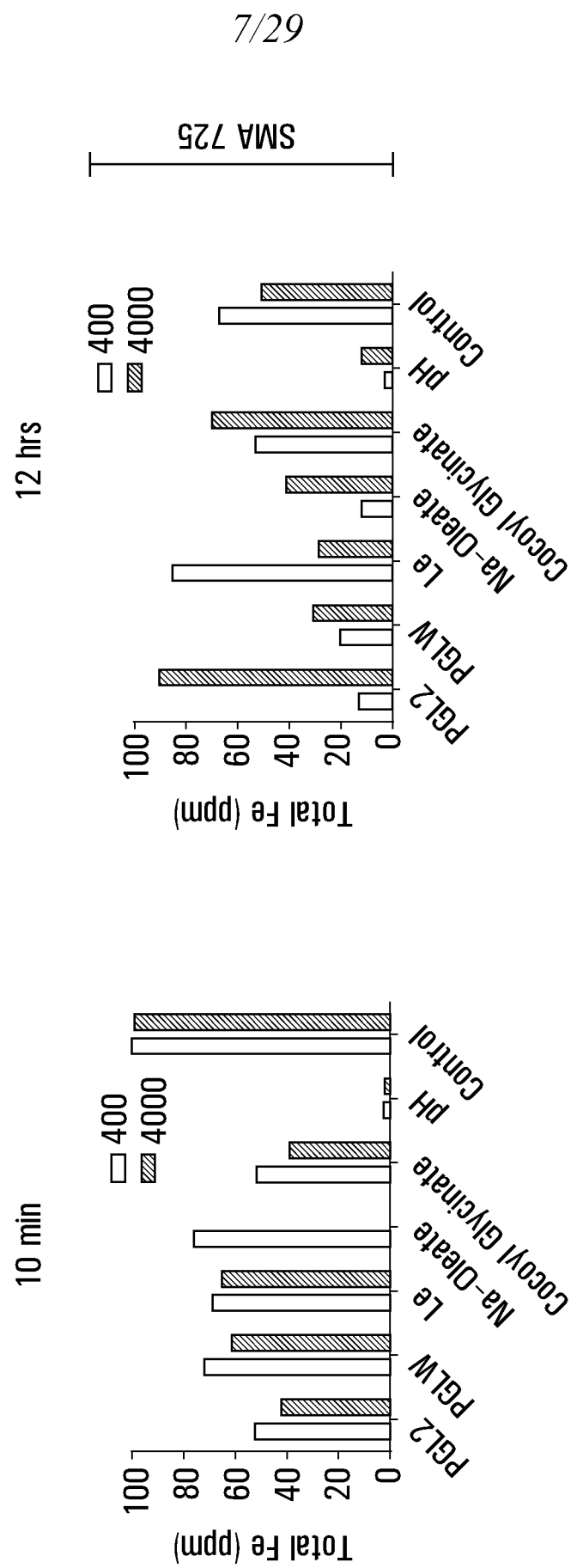
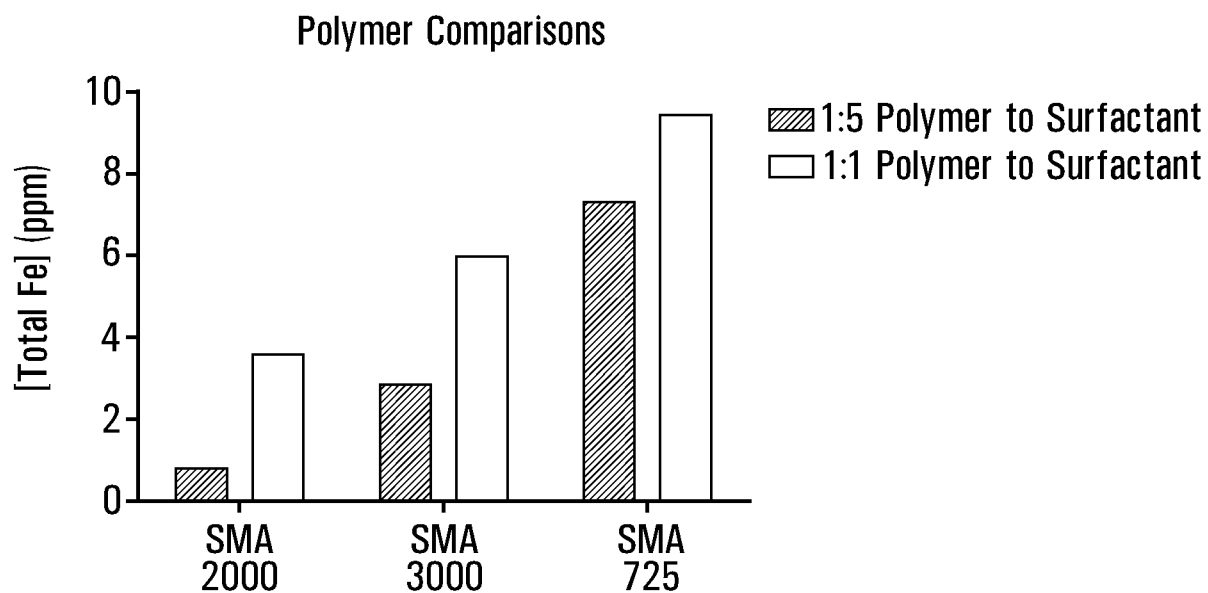
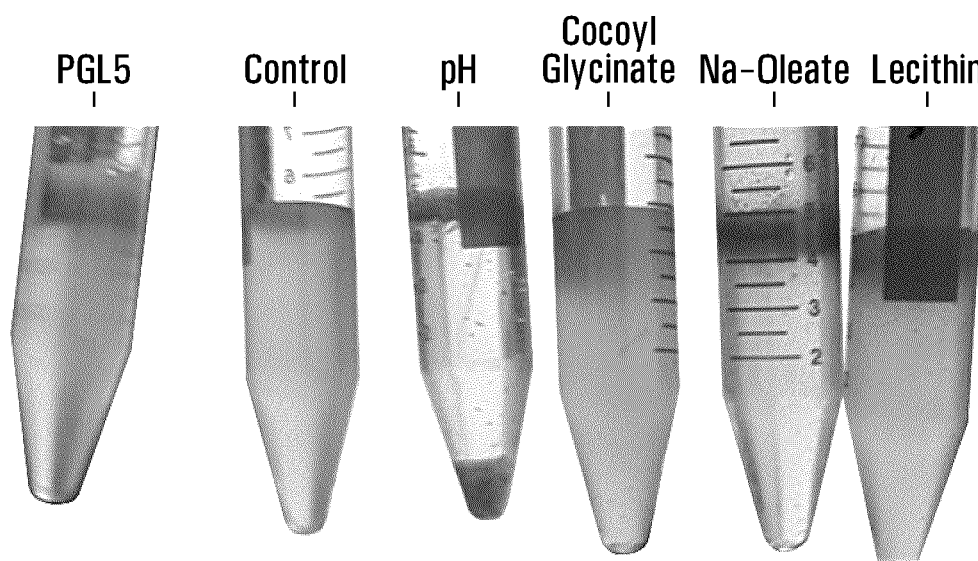


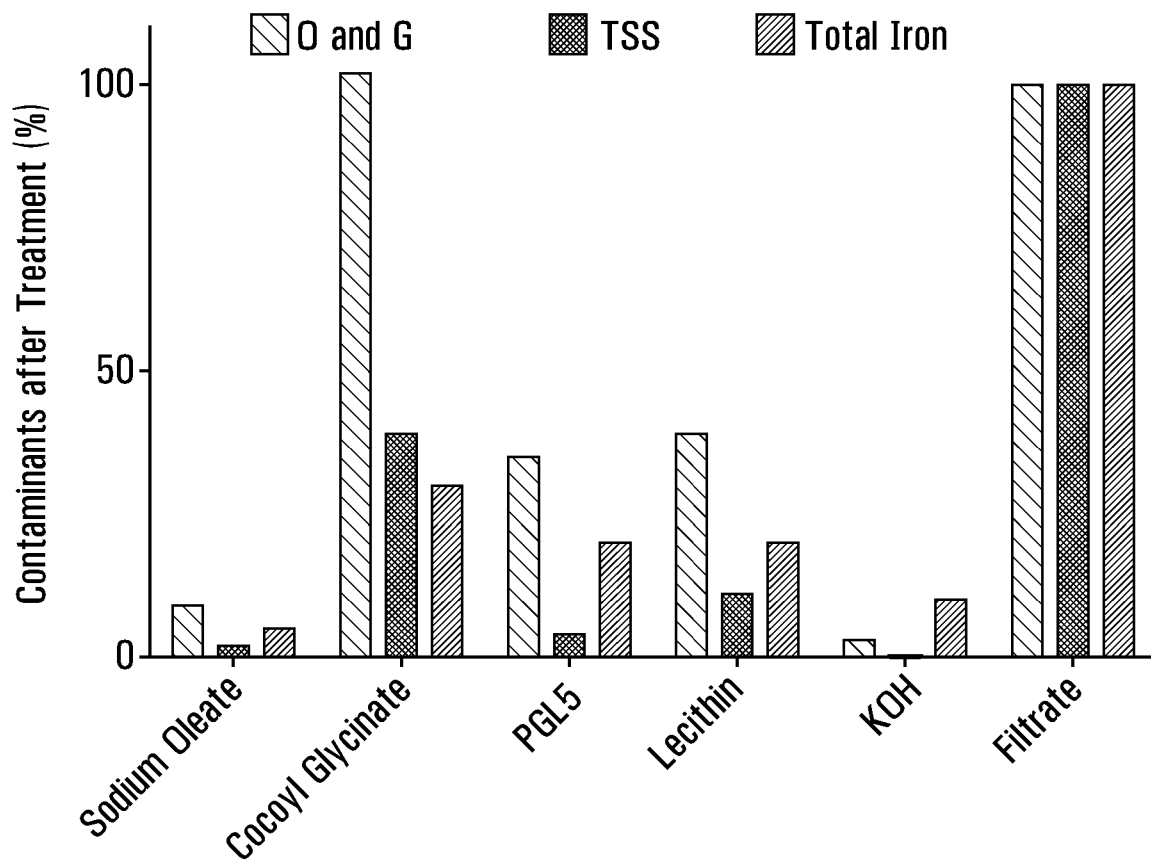
FIG. 7A

FIG. 7B

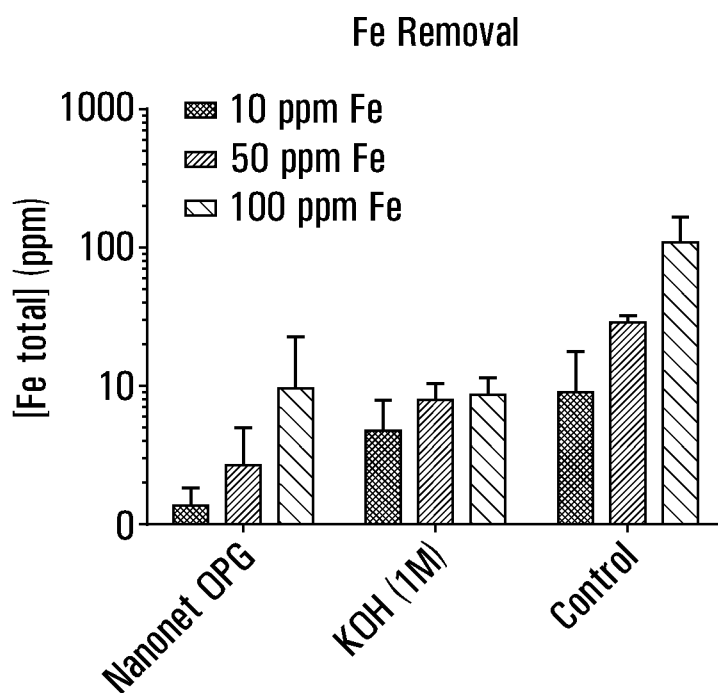
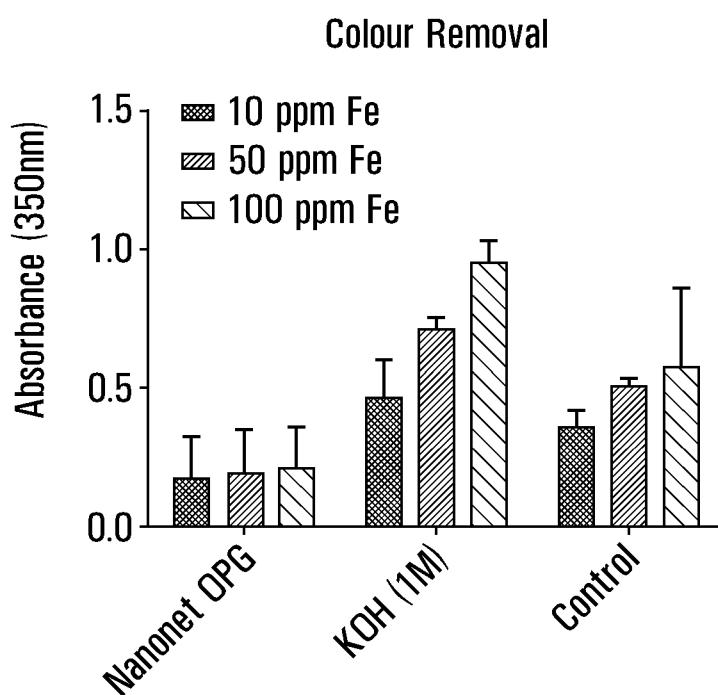
8/29

**FIG. 8****FIG. 9**

9/29

**FIG. 10**

10/29

**FIG. 11A****FIG. 11B**

11/29

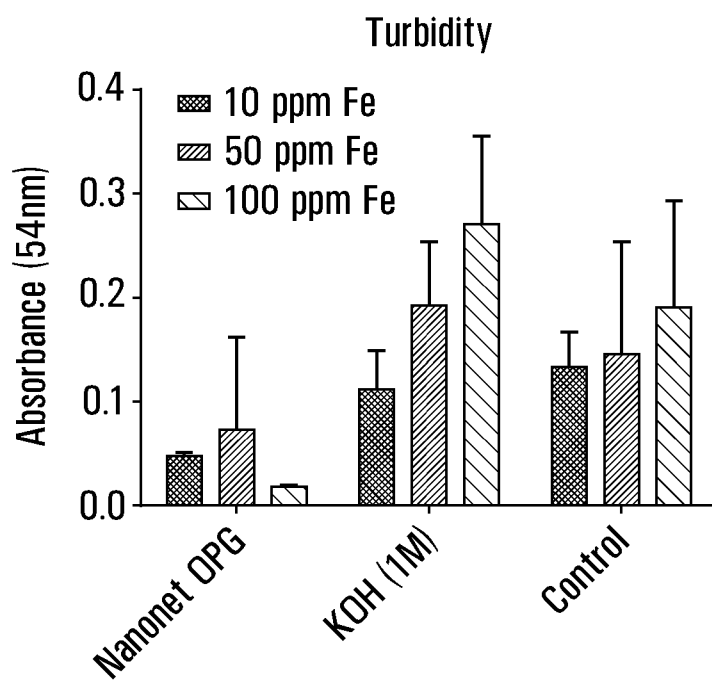
**FIG. 11C**

FIG. 12A

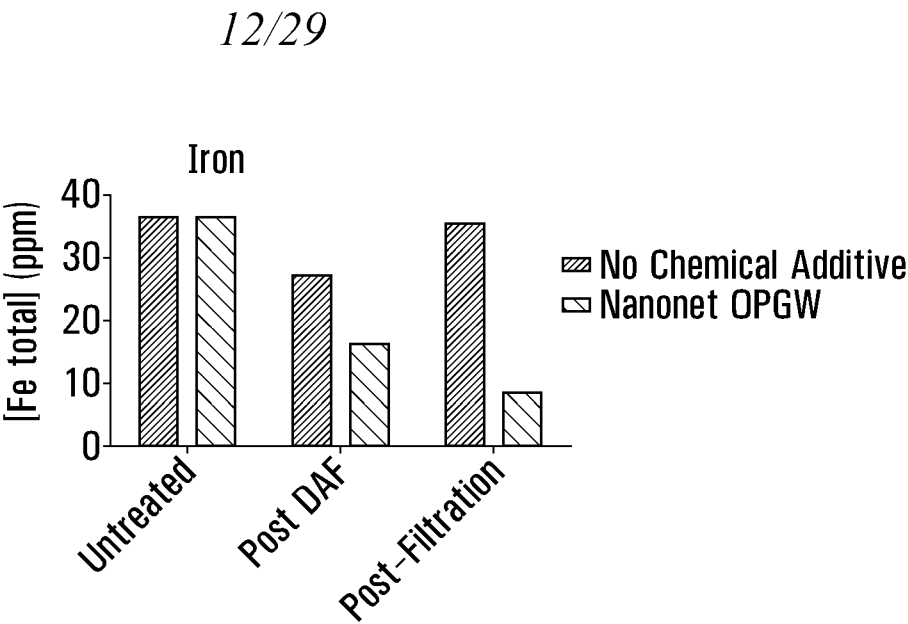


FIG. 12B

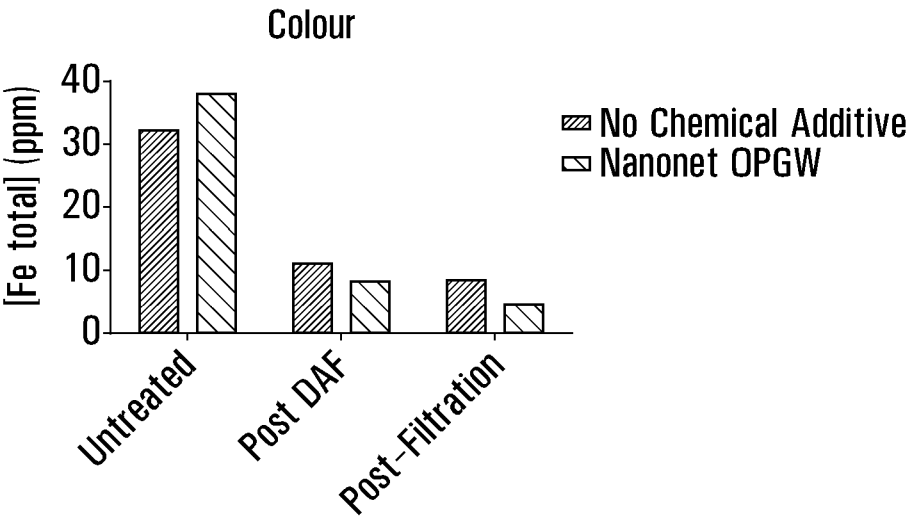
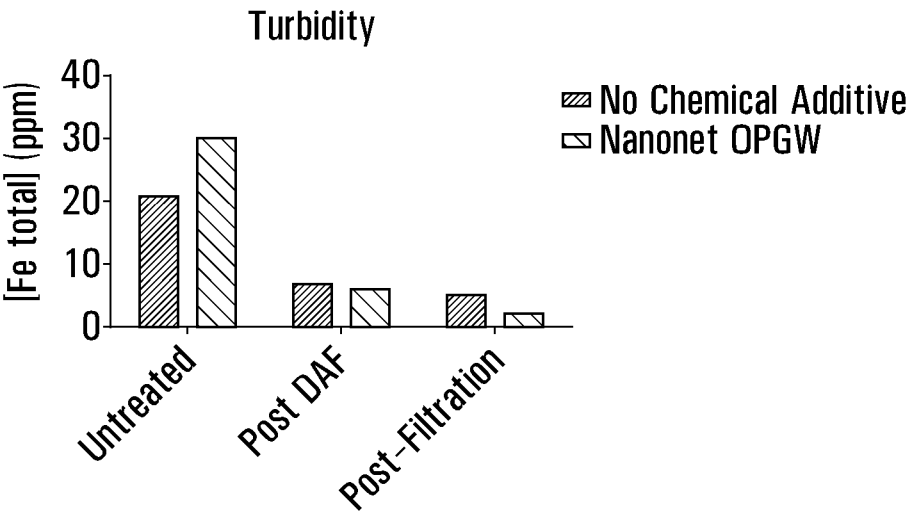


FIG. 12C



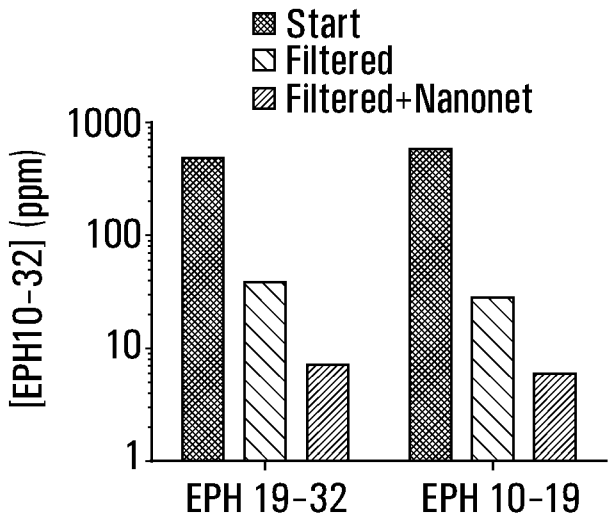


FIG. 13A

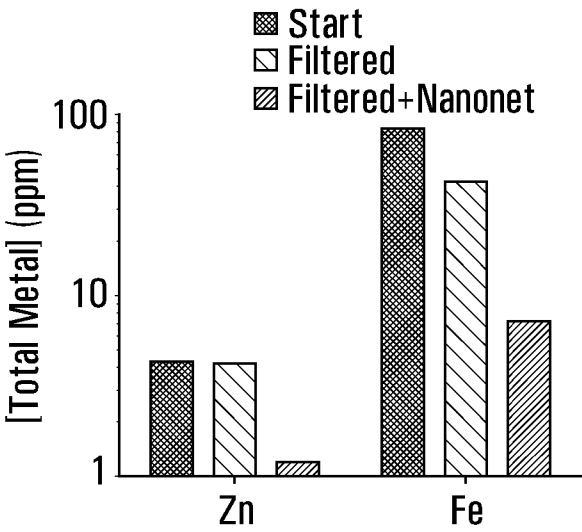


FIG. 13B

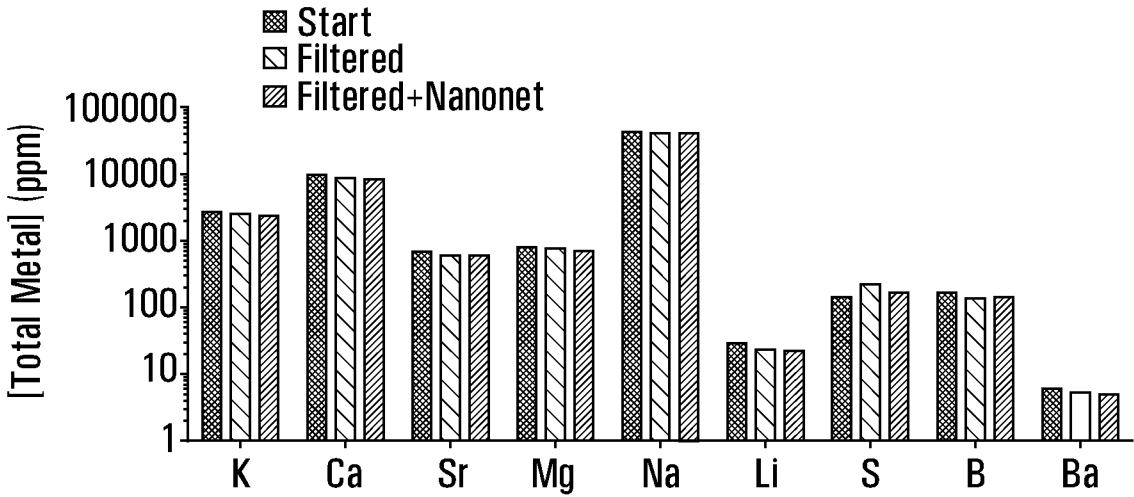


FIG. 13C

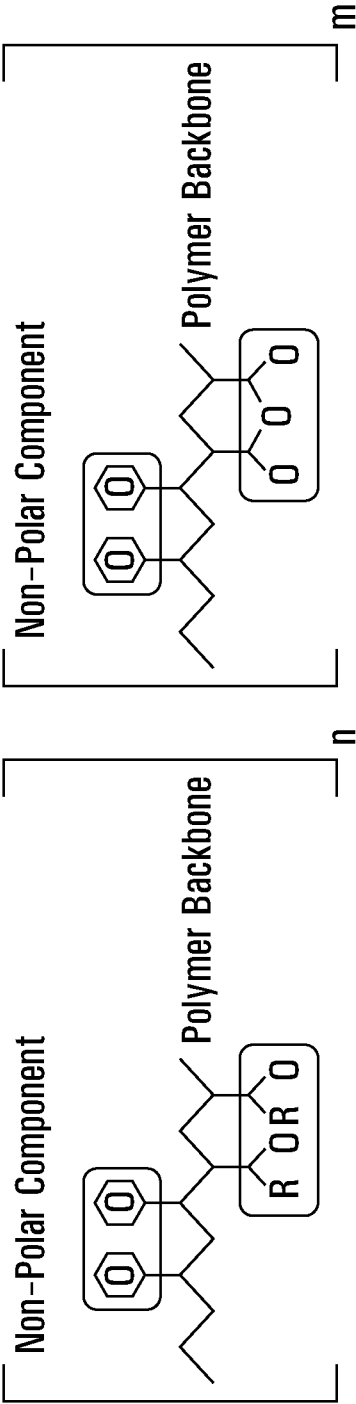
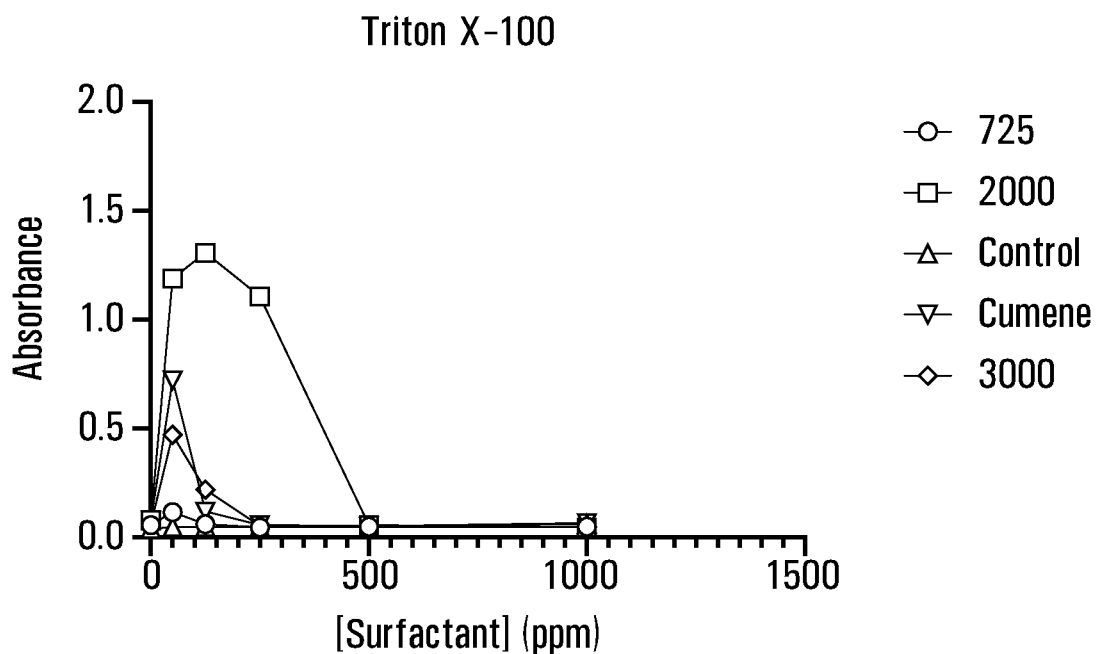
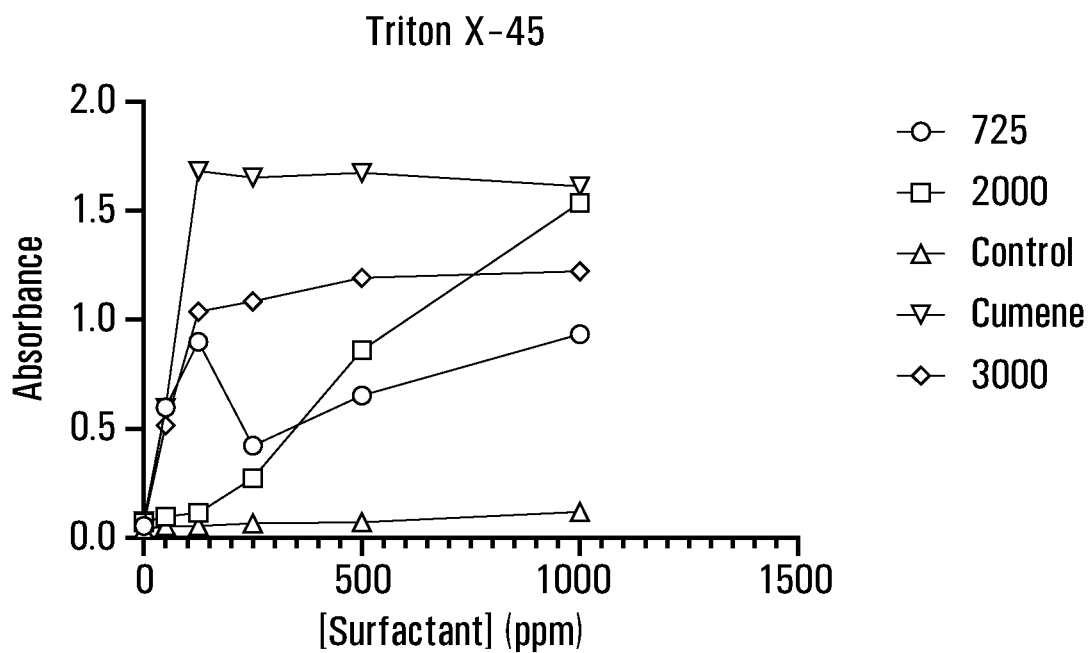


FIG. 14

15/29

**FIG. 15A****FIG. 15B**

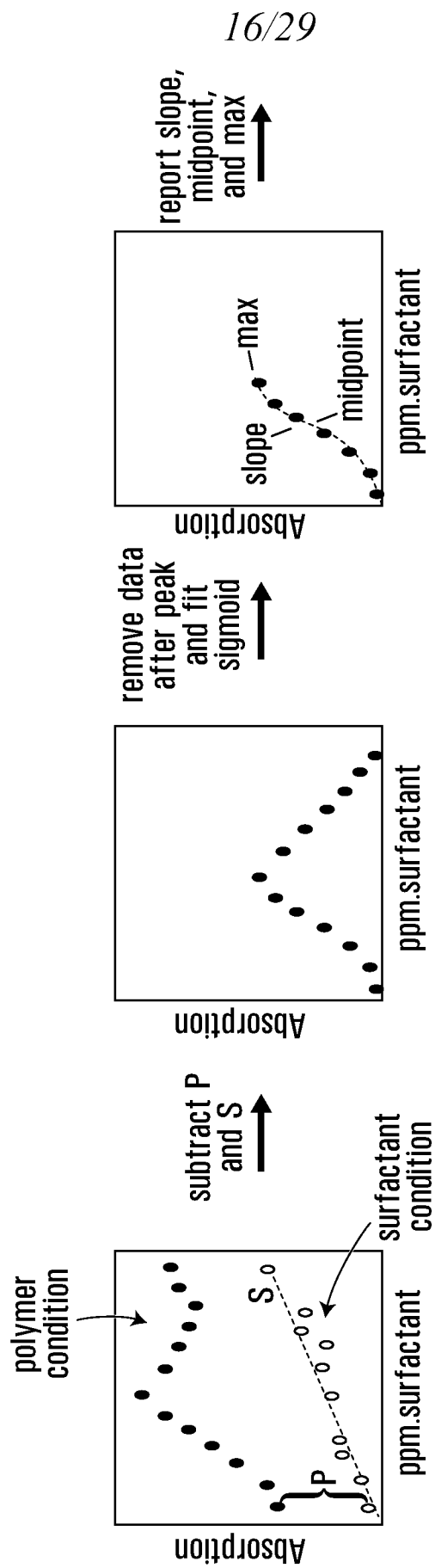
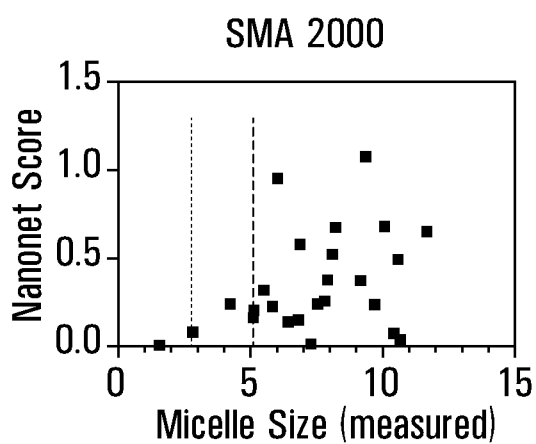
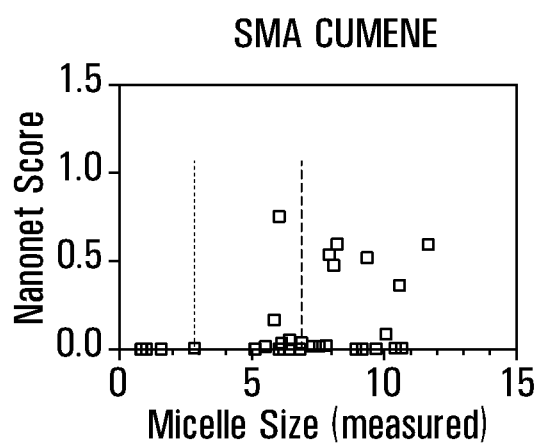
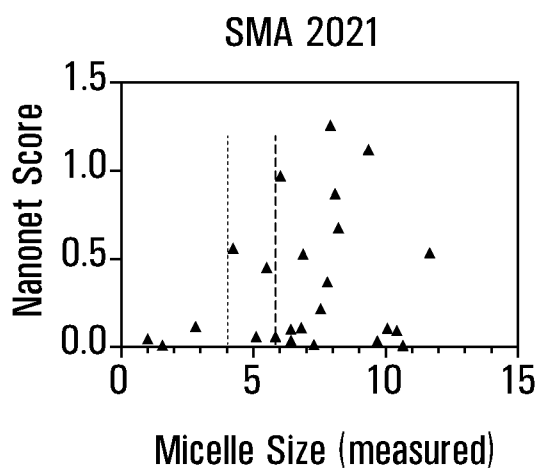
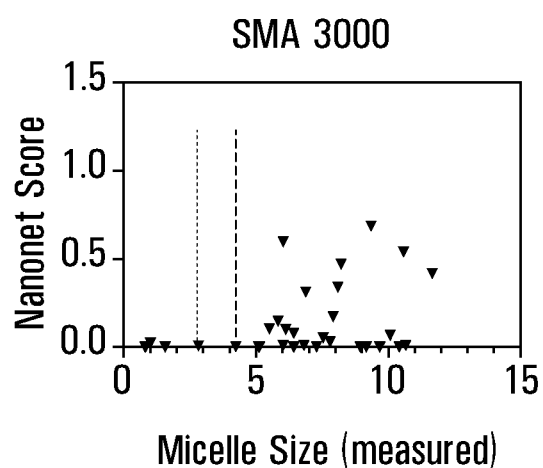
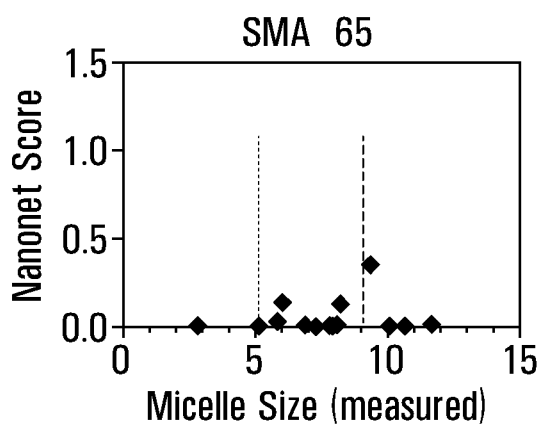
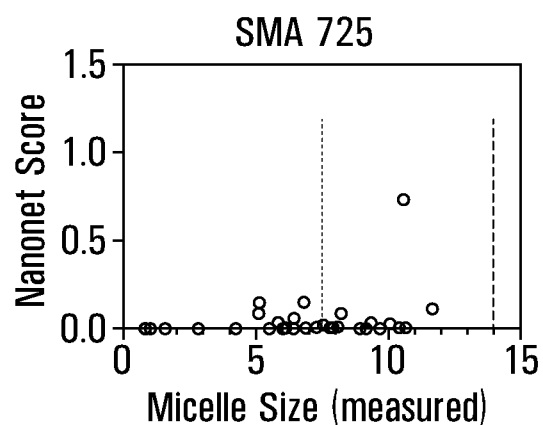


FIG. 16

17/29

**FIG. 17A****FIG. 17B****FIG. 17C****FIG. 17D****FIG. 17E****FIG. 17F**

18/29

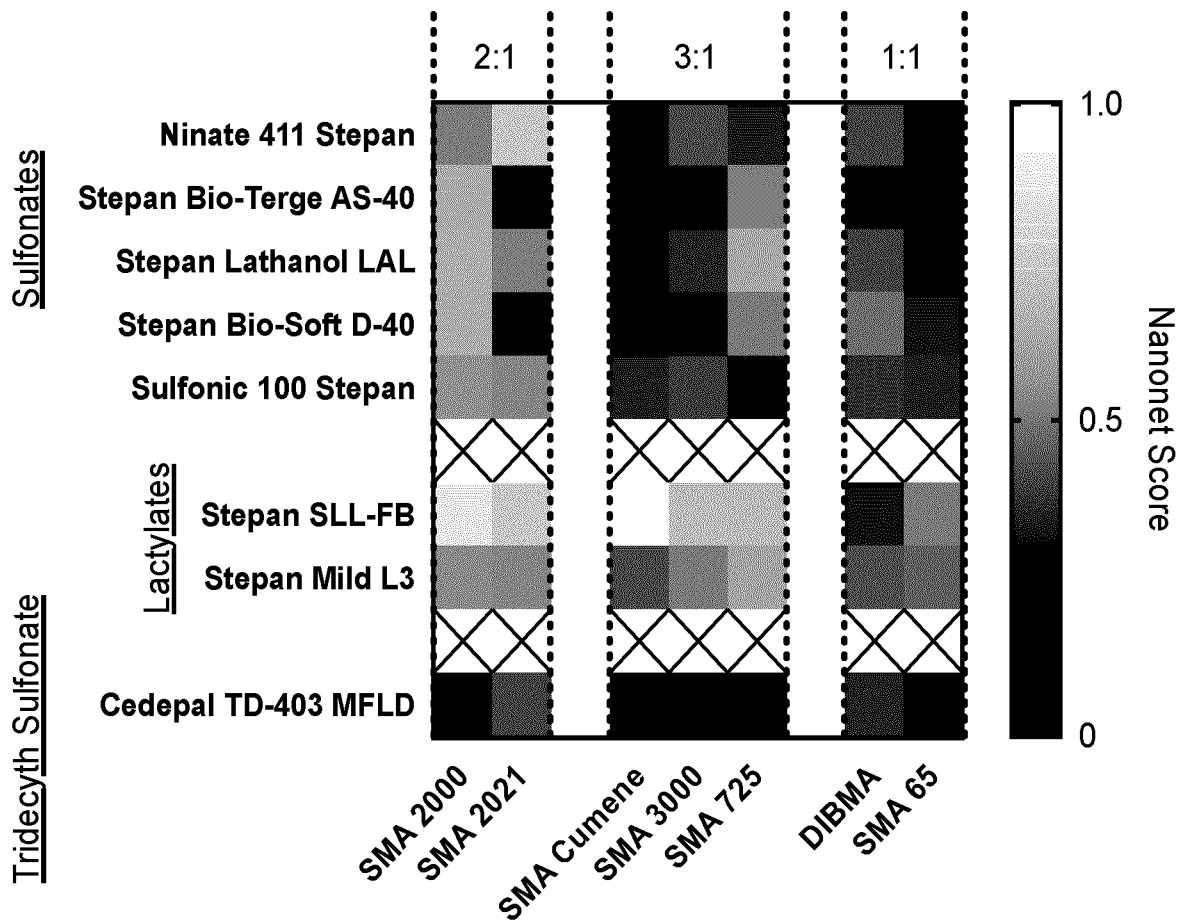


FIG. 18

19/29

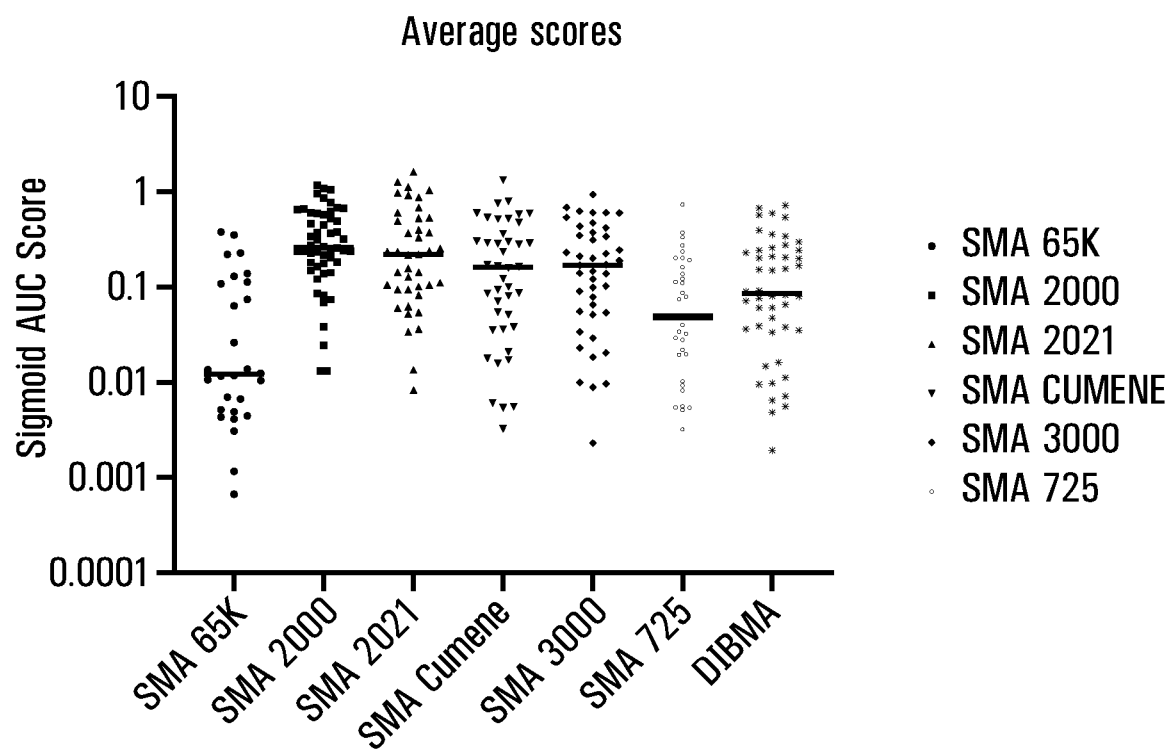
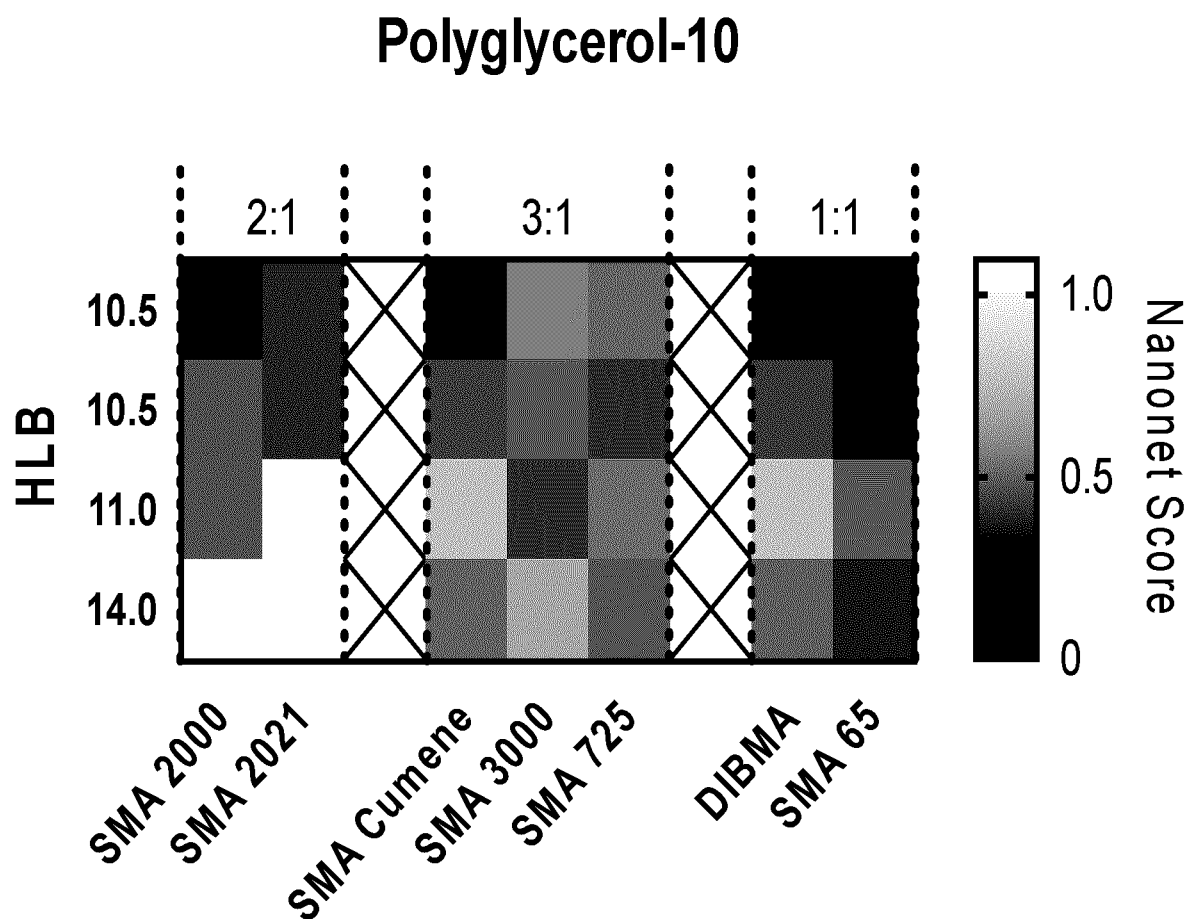


FIG. 19

20/29

**FIG. 20**

21/29

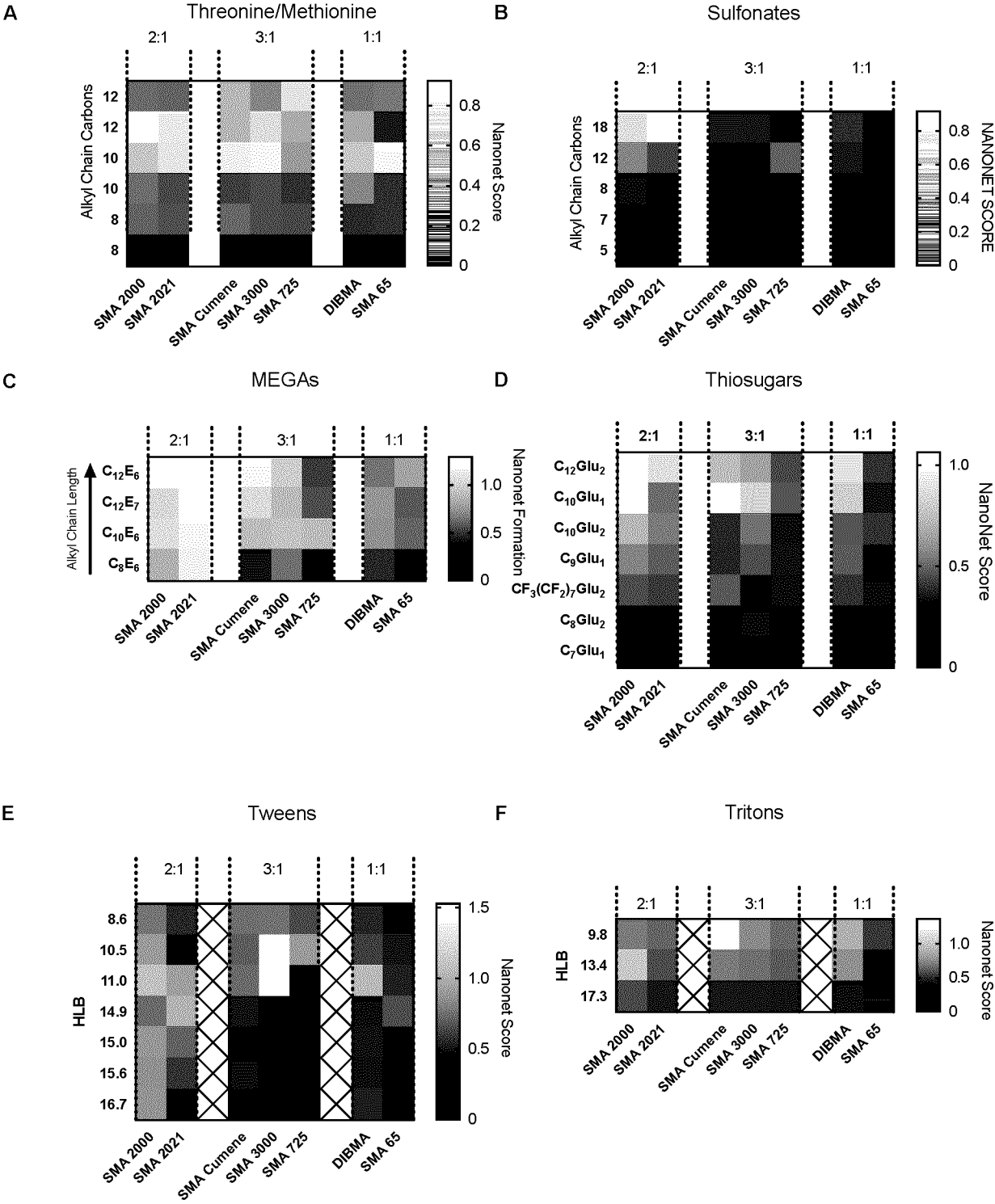
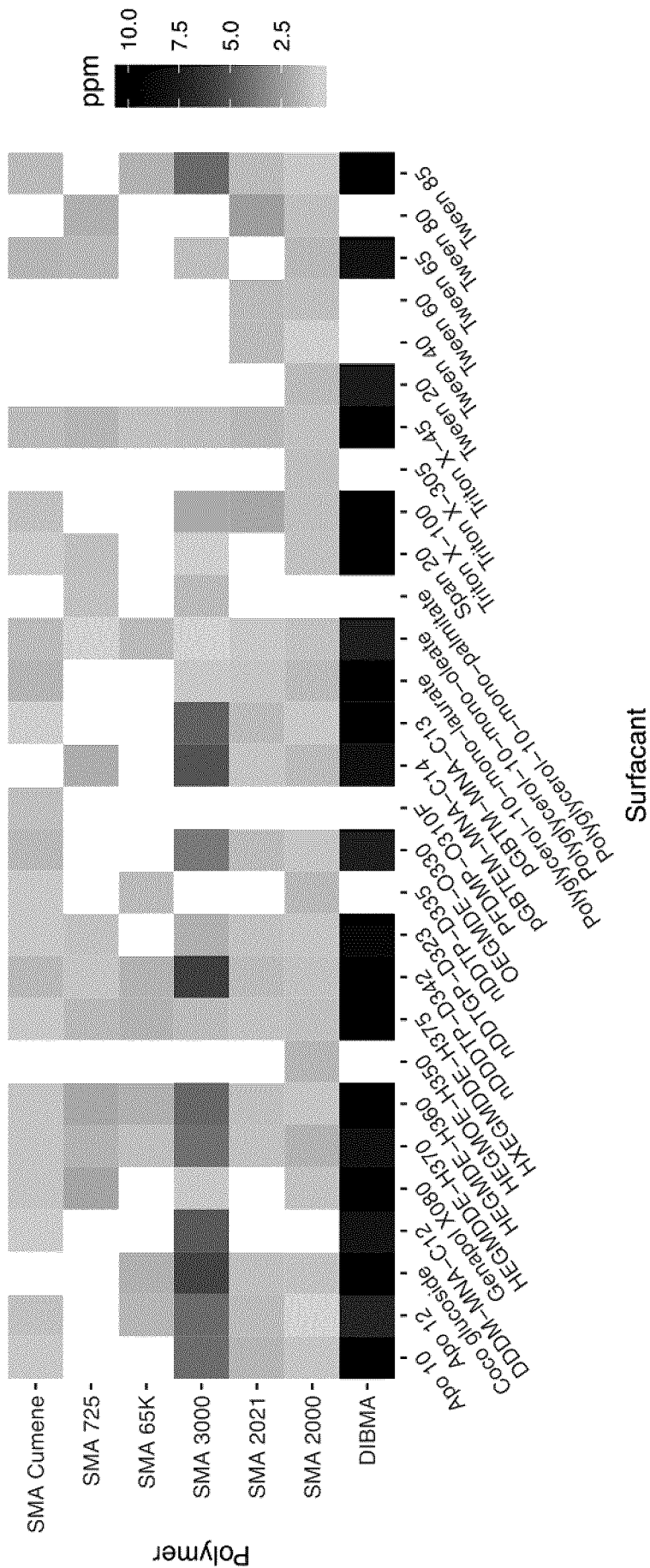


FIG. 21

FIG. 22



FIG. 23



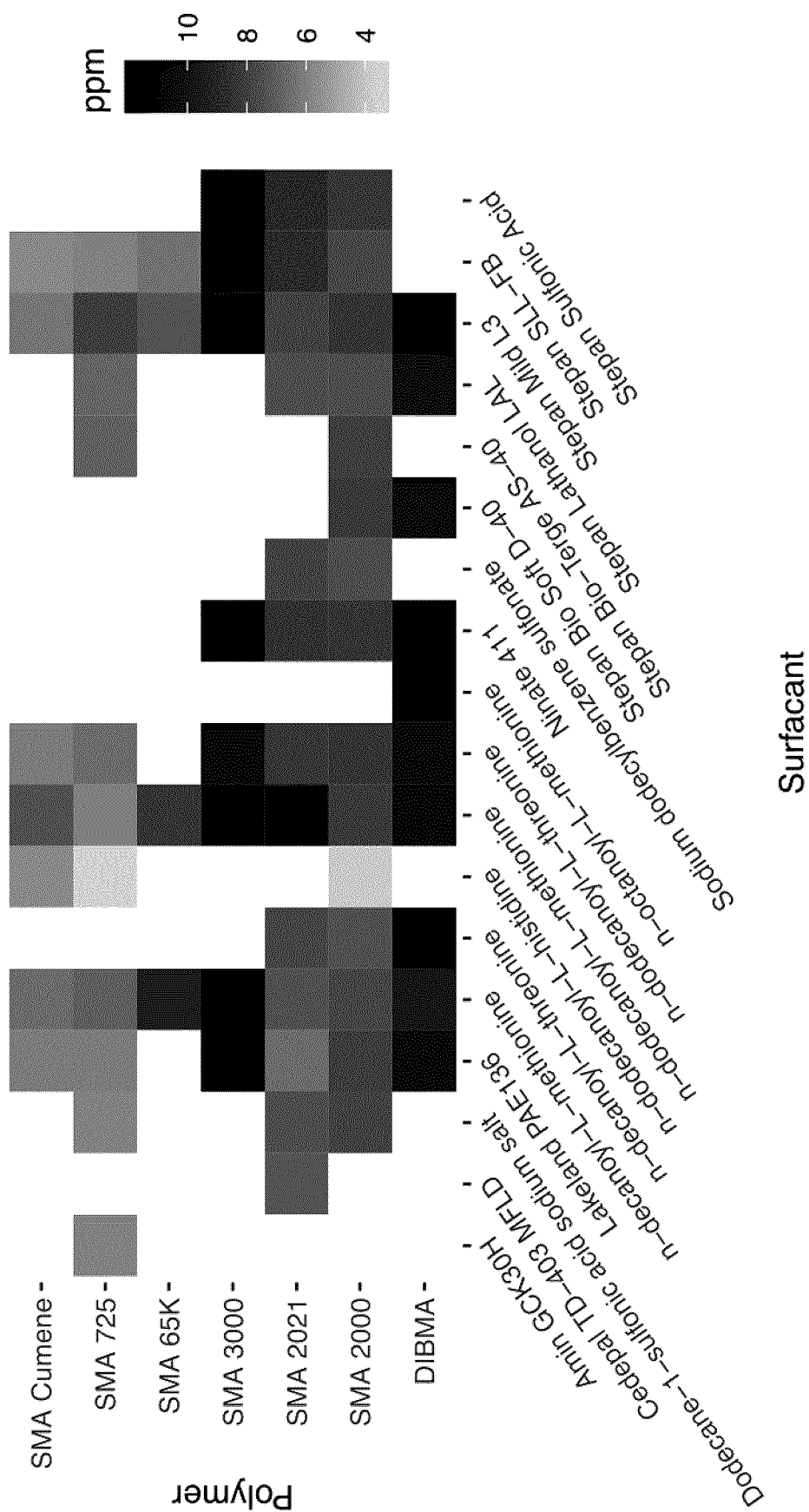
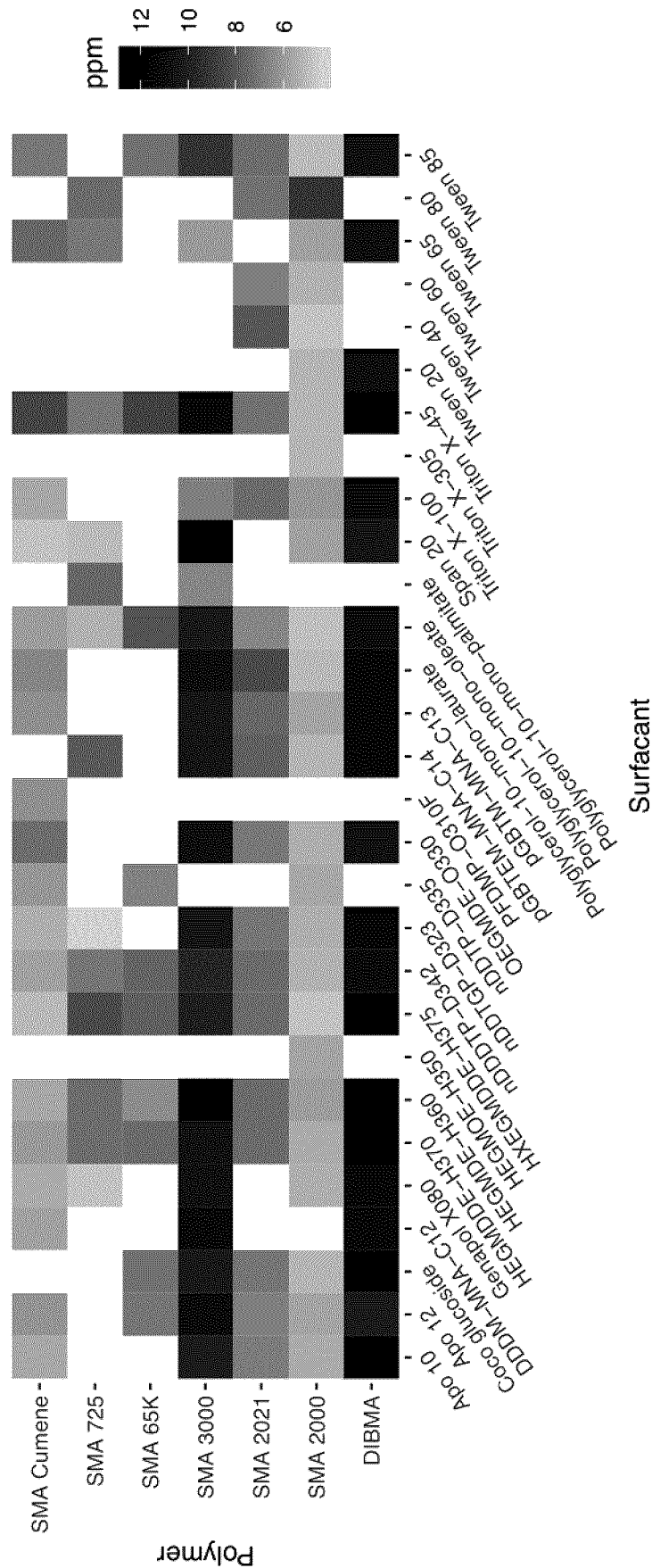
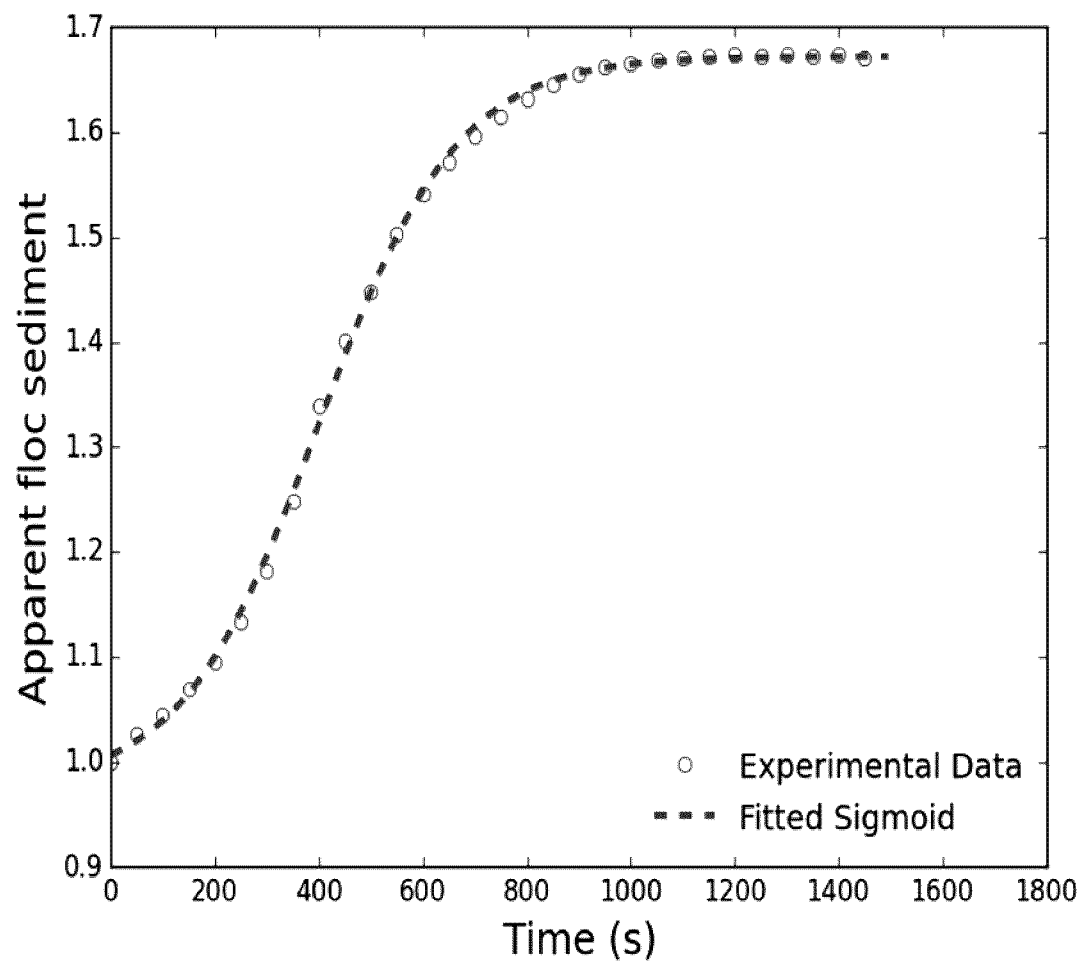


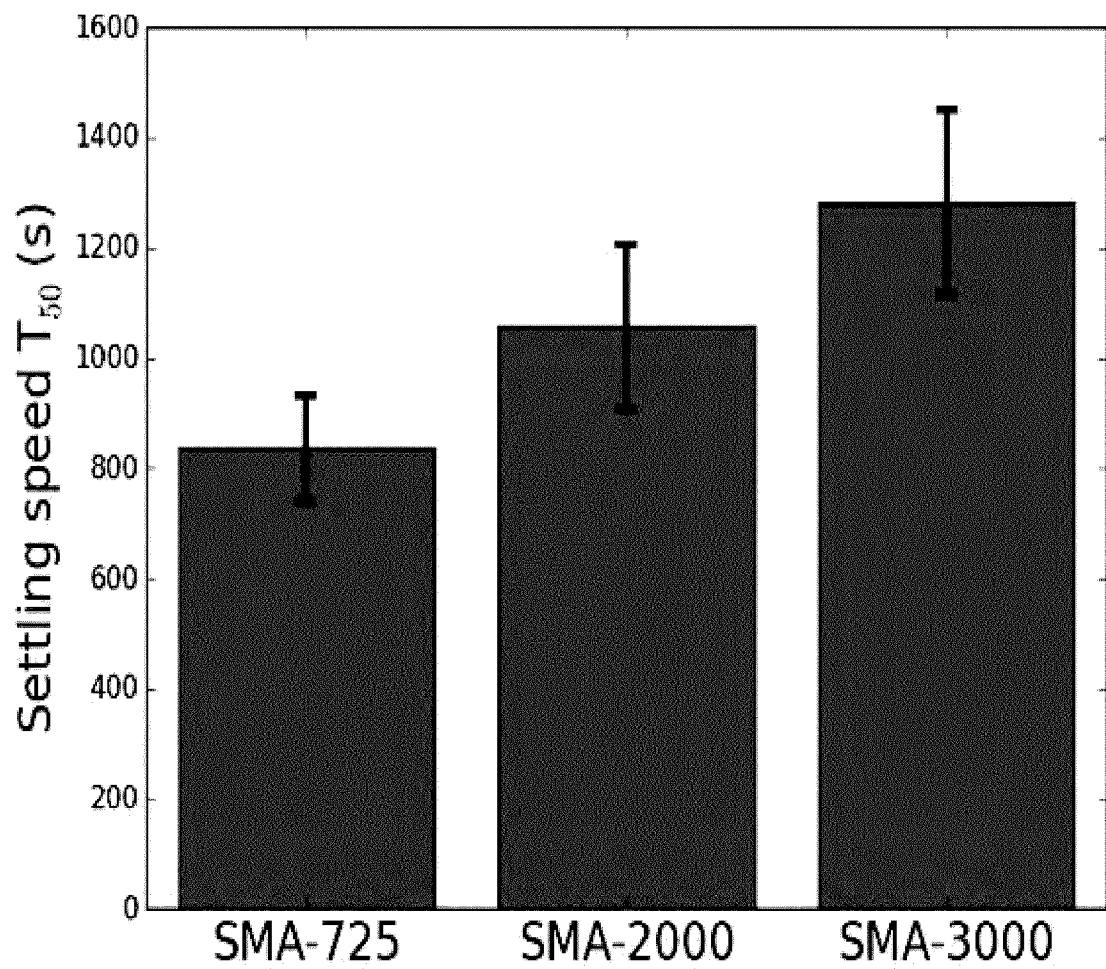
FIG. 25



26/29

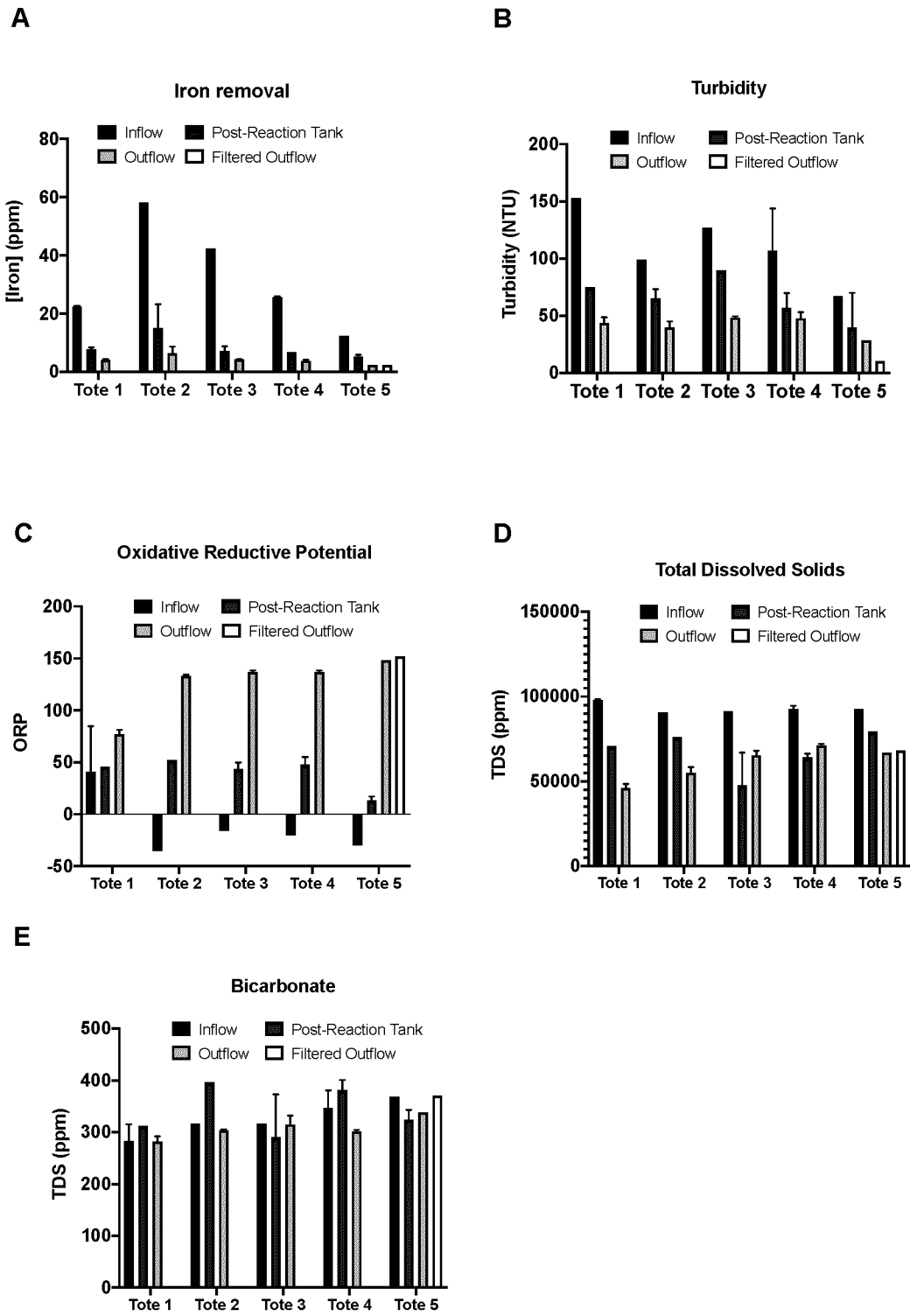
**FIG. 26**

27/29

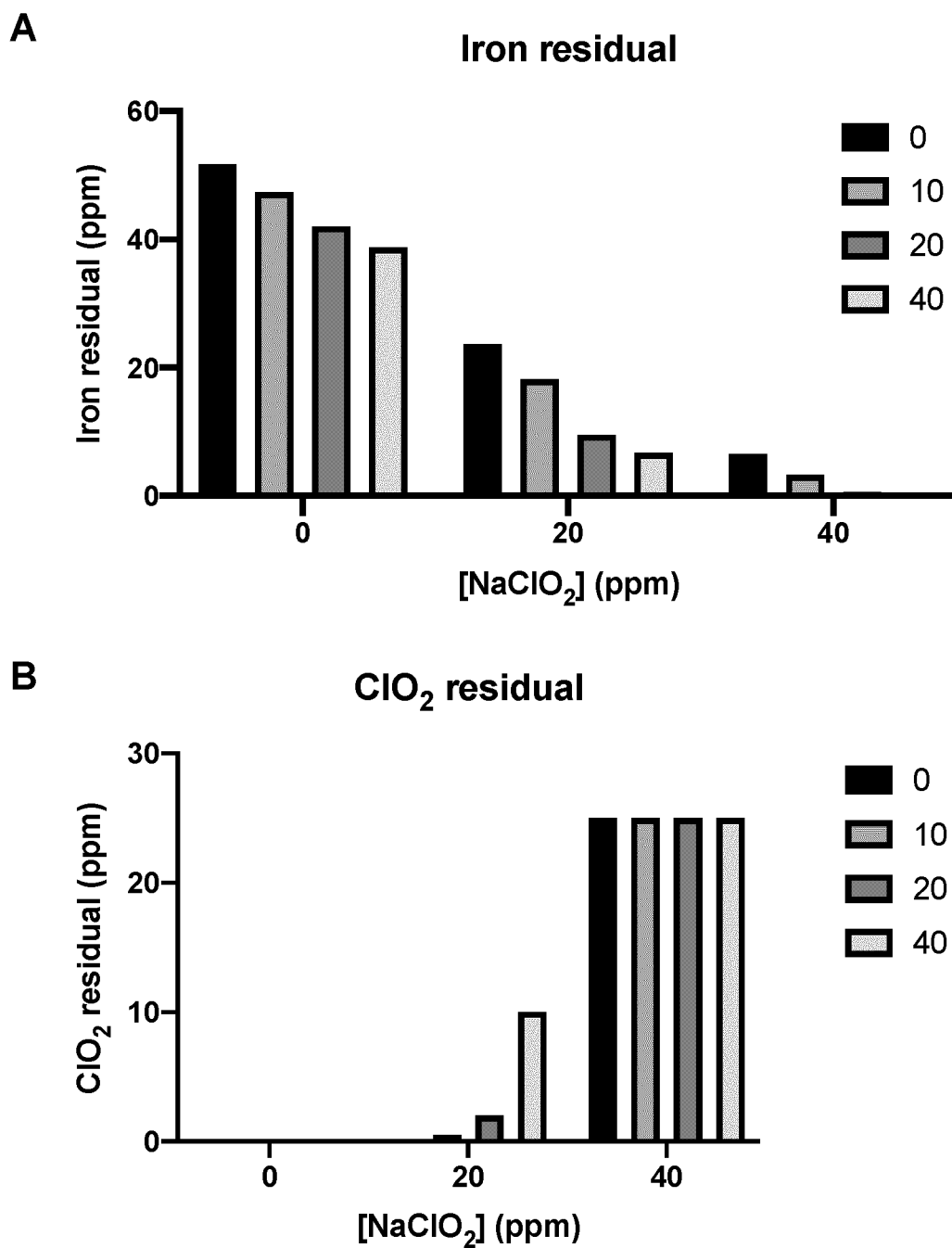
**FIG. 27**

28/29

FIG. 28



29/29

**FIG. 29**

INTERNATIONAL SEARCH REPORT

International application No.

PCT/CA2019/051739

A. CLASSIFICATION OF SUBJECT MATTER

IPC: **C02F 1/52** (2006.01), **B01J 20/22** (2006.01), **C02F 1/00** (2006.01), **C02F 1/26** (2006.01), **C02F 1/28** (2006.01)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC: C02F 1/52 (2006.01), B01J 20/22 (2006.01), C02F 1/00 (2006.01), C02F 1/26 (2006.01), C02F 1/28 (2006.01)

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic database(s) consulted during the international search (name of database(s) and, where practicable, search terms used)

Questel Orbit (FamPat)

Keywords: surfactant, polymer, precipit*, agent, nanonet

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US5391300 (WEBB et al.) 21 February 1995 (21-02-1995) *whole document	1-97
P, X	WO2018/218374 (CARLSON et al.) 06 December 2018 (06-12-2018) *whole document	1-97
A	US6524485 (DUBIN et al.) 25 February 2003 (25-02-2003) *whole document	1-97
A	US3580891 (RAINER) 25 May 1971 (25-05-1971) *whole document	1-97
A	US8115046 (BURNS et al.) 14 February 2012 (14-02-2012) *whole document	1-97

☒ Further documents are listed in the continuation of Box C.☒ See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"D" document cited by the applicant in the international application	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"E" earlier application or patent but published on or after the international filing date	"&" document member of the same patent family
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search
07 February 2020 (07-02-2020)Date of mailing of the international search report
27 February 2020 (27-02-2020)Name and mailing address of the ISA/CA
Canadian Intellectual Property Office
Place du Portage I, C114 - 1st Floor, Box PCT
50 Victoria Street
Gatineau, Quebec K1A 0C9
Facsimile No.: 819-953-2476

Authorized officer

Amy Westgate (819) 639-8587

INTERNATIONAL SEARCH REPORT

International application No.

PCT/CA2019/051739

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	CA2450592 (CHANG et al.) 09 June 2004 (09-06-2004) *whole document	1-97
A	WO2010/0017406 (HAAG et al.) 14 October 2010 (14-10-2010) *whole document	1-97

INTERNATIONAL SEARCH REPORT
Information on patent family members

International application No.

PCT/CA2019/051739

Patent Document Cited in Search Report	Publication Date	Patent Family Member(s)	Publication Date
US5391300A	21 February 1995 (21-02-1995)	None	
WO2018218374A1	06 December 2018 (06-12-2018)	CA3065594A1 CN110709356A IL270823D0	06 December 2018 (06-12-2018) 17 January 2020 (17-01-2020) 30 January 2020 (30-01-2020)
US6524485B1	25 February 2003 (25-02-2003)	None	
US3580891A	25 May 1971 (25-05-1971)	None	
US8115046B2	14 February 2012 (14-02-2012)	US2009245939A1 CA2650205A1 CA2650205C	01 October 2009 (01-10-2009) 18 July 2009 (18-07-2009) 29 March 2016 (29-03-2016)
CA2450592A1	09 June 2004 (09-06-2004)	AU2003266429A1 BR0305335A CN1510085A CN100376634C DE60315241D1 DE60315241T2 EP1428867A1 EP1428867B1 JP2004231941A KR20040050048A MXPA03011211A US2004110891A1	24 June 2004 (24-06-2004) 31 August 2004 (31-08-2004) 07 July 2004 (07-07-2004) 26 March 2008 (26-03-2008) 13 September 2007 (13-09-2007) 17 April 2008 (17-04-2008) 16 June 2004 (16-06-2004) 01 August 2007 (01-08-2007) 19 August 2004 (19-08-2004) 14 June 2004 (14-06-2004) 18 June 2004 (18-06-2004) 10 June 2004 (10-06-2004)
WO2010117406A1	14 October 2010 (14-10-2010)	TW201038513A TW1419861B US2011073802A1 US9090495B2	01 November 2010 (01-11-2010) 21 December 2013 (21-12-2013) 31 March 2011 (31-03-2011) 28 July 2015 (28-07-2015)